
Tubulins in *C. elegans**

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Table of Contents

1. Tubulin, microtubules, and tubulin mutations in <i>C. elegans</i>	2
2. The multi-tubulin hypothesis and cellular and developmental roles for microtubules	4
2.1. The mitotic and neuronal tubulins: TBA-1, TBA-2, TBB-1, TBB-2, and BEN-1	7
2.2. The sensory neuron axonemal tubulins: TBA-5, TBA-6, TBA-9, and TBB-4	8
2.3. The mechanosensory tubulins: MEC-12 and MEC-7	9
2.4. α - and β - tubulins with undiscovered roles	9
3. Beyond α - and β -tubulin	10
3.1. TBG-1	10
3.2. Other tubulins	10
4. Cellular control of microtubules	10
4.1. Tubulin biogenesis and homeostasis	10
4.2. Post-translational modification	11
4.3. Microtubule-associated proteins (MAPs)	11
5. Concluding remarks and future directions	14
6. Tables	15
6.1. Table 1. Tubulin alleles that have been classified genetically, have an identified molecular change, or are represented by a strain at the CGC	15
6.2. Table 2. Summary of the expression and function of <i>C. elegans</i> tubulins	19
6.3. Table 3. Summary of the expression and function of regulators, modifiers, and proteins associated with <i>C. elegans</i> tubulin and microtubules	21
7. Acknowledgements	23
8. References	23

Abstract

The *C. elegans* tubulin family is composed of nine α -, six β -, and one γ -tubulin. Tubulins are highly conserved, functioning as α - β heterodimers that assemble into microtubules. These cylindrical and ubiquitous components of the cytoskeleton are critical for nearly all cellular and developmental processes. *C. elegans* has provided a model for the study of microtubules in multiple settings including separation of chromosomes, cellular polarity, and neuronal sensation. Tubulins and microtubules interact with a **long** list of other cellular

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proteins that regulate tubulin homeostasis, modify microtubule dynamics, and control incorporation into or disassociation of higher-order cellular structures such as spindles or ciliary axonemes. A collection of enzymes modifies tubulins, often at the variable carboxyl-terminal tail, adding another layer of regulation to microtubule structure and function. Genetic and cytological studies in *C. elegans* have revealed roles for tubulin and its associated proteins in numerous contexts from embryogenesis to adult behavior.

1. Tubulin, microtubules, and tubulin mutations in *C. elegans*

Tubulins are ~450 amino acid globular proteins of the cytoplasm that comprise microtubules (Figure 1). The sequences of both α - and β -tubulins (Figure 2), which form heterodimers, indicate that they are part of a large family of proteins that bind to and hydrolyze guanosine triphosphate (GTP). Tubulin-like proteins appear early in evolution, and they have remained highly conserved in the eukaryotic lineage. Tubulins are related to the FtsZ proteins of eubacteria. Like tubulins, FtsZ proteins polymerize and depolymerize with a GTPase cycle and play a role in cell division (Ingerson-Mahar and Gitai, 2012). Genes for FtsZ homologs are widely distributed in prokaryotic lineages, but closer relatives to eukaryotic tubulins are found in a few archaeal species (Yutin and Koonin, 2012). Amongst eukaryotes, the α - and β -tubulins are nearly identical to each other within their families, as most of the sequence divergence is located in the carboxyl-terminal tails which emanate from the outer surface of the microtubule and can impart specific functions to tubulins, perhaps through interactions with motor proteins and/or other microtubule-associated proteins (Hsu et al., 2014; Nogales et al., 1999; Popodi et al., 2008).

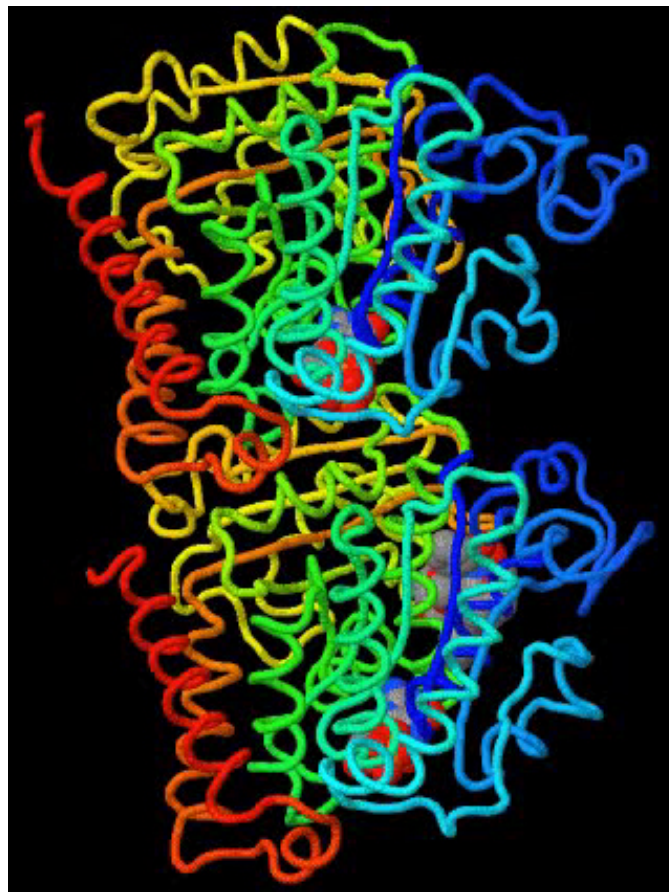


Figure 1. Rainbow decorated three-dimensional structure of the tubulin dimer. Colors indicate position in the primary structure as in Figure 2. The amino-terminal GTP binding elements (Rossmann fold) span from blue to green and face the interior of the microtubule. Space-filling ligands include GTP bound to both α -tubulin (upper) and β -tubulin (lower), and paclitaxel (taxol) bound to β -tubulin. The external surface of the microtubule is composed of the yellow through red secondary structures. Carboxyl-terminal tails are not part of the crystal structure of the dimer. Animation was created using FirstGlance in Jmol (<http://firstglance.jmol.org>) and the PDB identification code 1TUB (Nogales et al., 1999).

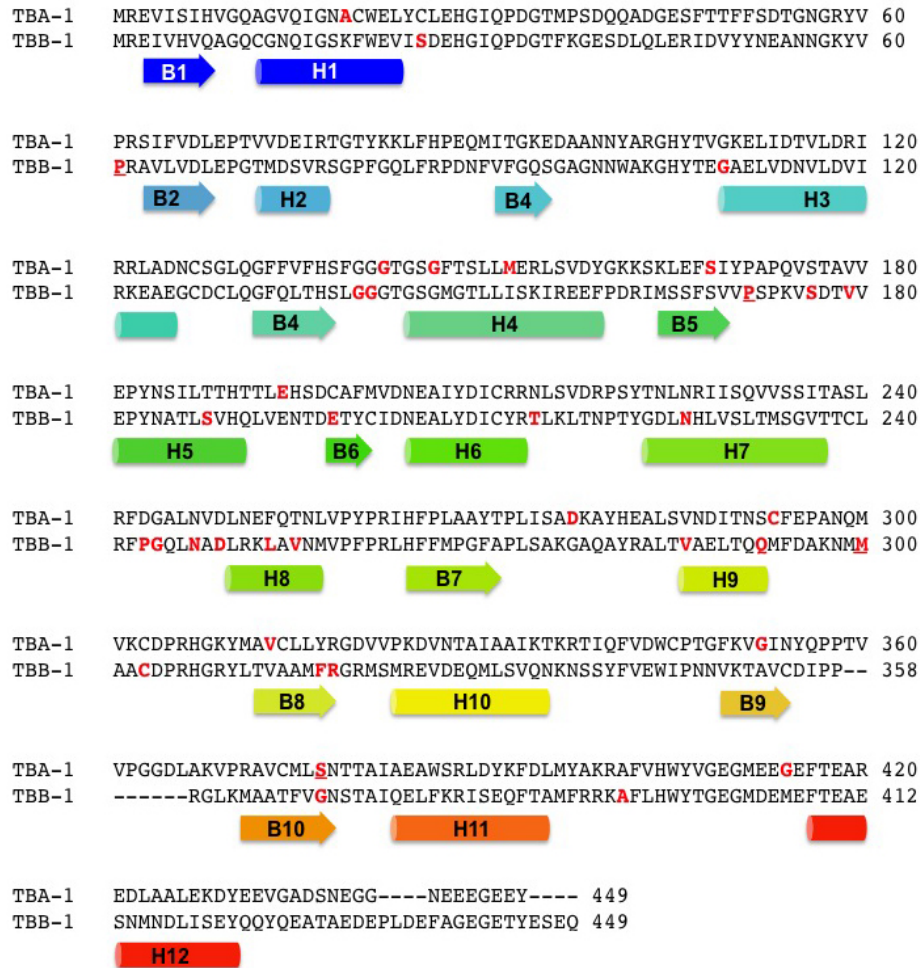


Figure 2. A pairwise alignment of TBA-1 and TBB-1 with dominant alleles and secondary structure. Genetically defined dominant, semi-dominant, and/or gain of function alleles are shown in red and underlined if they have been independently discovered more than once (Table 1). Colored α -helices (cylinders) and β -ribbons (arrows) as in Figure 1 are shown below the primary structure.

Microtubules are semi-rigid, cylindrical cytoskeletal polymers composed of repeating units of the α - and β -tubulin heterodimer. Heterodimers, which are formed soon after synthesis of monomers, associate head to tail into protofilaments. Kinetic and structural data suggest that the further assembly of heterodimers and short protofilaments into microtubules is explained by a lattice model. This model suggests that incorporation into a microtubule straightens soluble, curved heterodimers, while nucleotide status tunes the strength of contacts within the lattice of dimers. Hydrolysis of GTP to GDP in the exchangeable site (E-site) of β -tubulin within a microtubule causes compaction and other conformational changes in both subunits of the dimer that weaken the contacts holding the lattice together, making the microtubule unstable (Rice et al. 2008; Alushin et al., 2014). The structure and nucleotide hydrolysis capacity of tubulin imparts microtubules with dynamic instability, an intrinsic property of self-assembly and disassembly (Mitchison and Kirschner, 1984).

In vivo, microtubules are commonly nucleated by the γ -tubulin ring complex and other components of centrosomes (Teixido-Travesa et al., 2012), which pattern protofilaments into the helical pitch of a microtubule. Nematode microtubules are 11 protofilaments in most cells, while typical eukaryotic microtubules are 13 protofilaments. Touch receptor cells in nematodes have 15 protofilament microtubules (Chalfie and Thomson, 1982). Microtubules are polar polymers, with assembly and disassembly occurring preferentially at the plus end. Similar activities take place at the minus end, although at a slower rate. Polymer polarity is a hallmark of microtubules (and actin), and it often establishes and/or maintains cellular polarity (e.g., see Dubey et al. 2015).

Mutations in genes encoding tubulins in *C. elegans* have been discovered in screens for a variety of phenotypes. The forward genetic approach has produced a large collection of alleles for which there is a rich and

growing collection of phenotypic information. This allows for understanding the cellular role of tubulin in many different contexts (see [Section 2](#)). In addition, because the *C. elegans* genome has a fairly large family of tubulin genes (with many being non-essential), dominant mutations in tubulin have been isolated and propagated. Mapping of these alleles to the structure has provided insight into the function of tubulin, especially in the case of genetically explored and molecularly identified point mutations ([Figure 2](#) and [Table 1](#), [Section 6.1](#)).

Tubulin can be divided into three domains. Domain one spans the first 200 amino acids, and it includes the Rossman fold and nucleotide-binding portion that faces the lumen of the microtubule. Dominant mutations have been found in the glycine-rich stretch (GGGTG[T/S]G, sometimes called the tubulin signature motif) in between B4 and H4 of both α and β -tubulin ([Figure 1](#) and [Figure 2](#)). Dominant mutations of the first of a conserved set of prolines (aa170) that reside in the pocket around the purine base of GTP have been found twice, both in β -tubulin. Other dominant mutations are scattered in the amino-terminal half of both tubulins, often in between or near the ends of α helices or β ribbons. Two intragenic revertants of a dominant mutation in the conserved S169 have also been identified.

The second domain (sometimes called the activation domain) spans the remaining conserved amino acids up to the divergent C-terminal tails (CTTs, [Figure 3](#)). As noted by [Erickson, \(1998\)](#), dominant mutations near or in the highly conserved G244, N247, and D249 of the ‘synergy loop’ have only been found in β -tubulin, typically in *mec-7*, although the nearby L253 in H8 is altered in an allele of *tbb-4*. This collection of alleles is curious as the amino acids they change in β -tubulin form part of the binding pocket for the non-exchangeable GTP nucleotide in the dimer-dimer interface that is mostly bound by α -tubulin ([Figure 1](#) and [Figure 2](#)). More exploration will reveal whether this is specific to these worm tubulins or telling something broader about dimer formation and the non-exchangeable GTP. A mutation of the conserved methionine at position 300 has been independently isolated twice (both in *mec-7*), and the serine at position 377 is mutated to phenylalanine in two independent alleles of *tba-1*. Negative charges around a glycine (aa416 in *mec-12*, aa414 in [Figure 2](#)) near H12 modulate the interaction between microtubules and motor proteins such as dynein ([Hsu et al., 2014](#)). As is the case in the amino terminal portion of tubulin, many of the dominant mutations are in between or near the ends of the α -helices and/or β -ribbons. None have yet been found in a CTT.

alpha-tubulins	
TBA-1/F26E4.8/WP_CE09692	KDYEEVGADSNEGGNEEEGEEY-----
TBA-2/C47B2.3/WP_CE17563	KDYEEVGADSNEGG-EEEGEEY-----
MEC-12/TBA-3/C44B11.3/WP_CE24843	KDYEEVGVDSDMEDNG-EEGDEY-----
TBA-4/F44F4.11/WP_CE18680	KDYEEVGADSNEGL-EEDGEEY-----
TBA-5/F16D3.1/WP_CE09434	KDYEEVGVDSPD-P---NDEEY-----
TBA-6/F32H2.9/WP_CE34484	KDYEEIGEDELPPDDIDDQSYRGRSSGSR
TBA-7/T28D6.2/WP_CE16521	KDYEEVGADS-DAN-DNGDDEY-----
TBA-8/ZK899.4/WP_CE37468	KDYAEVSRDTADL--EEENDEF-----
TBA-9/F40F4.5/WP_CE30131	KDYEEVGLDAGEPDEEDDYSHY-----
beta-tubulins	
TBB-1/K01G5.7/WP_CE16197	YQQYQEATAEDEPLDEFAGEAGETYESEQ
TBB-2/C36E8.5/WP_CE00913	YQQYQEATAEDDVGYAEGEAGETYESEQ
MEC-7/TBB-3/ZK154.3/WP_CE15257	YQQYQEAAADEDAAEAFDGE-----
TBB-4/B0272.1/WP_CE00850	YQQYQEATADDE EGEF DEHDQDVE-----
BEN-1/TBB-5/C54C6.2/WP_CE33770	YQQYQEATAEEDGELDGTGDAE-----
TBB-6/T04H1.9/WP_CE36500	FQQY-EKVHSA-----

Figure 3. Multiple alignment of α -tubulin and β -tubulin C-termini. An alignment of the C-terminal tails (CTTs) of the nine α -tubulins and six β -tubulins. The sequence shown starts at the end of helix 12 ([Figure 2](#)). Tubulins within each family are nearly identical to each other throughout the rest of their structure. The axonemal motif in TBB-4 is shown in bold. For an alignment of complete sequences of all the tubulins in *C. elegans*, see [Gogonea et al., 1999](#).

2. The multi-tubulin hypothesis and cellular and developmental roles for microtubules

The *C. elegans* tubulin superfamily consists of nine α -tubulins, six β -tubulins, and a single γ -tubulin ([Table 2](#), [Section 6.2](#)). The presence of multiple, closely related α - and β -tubulin paralogs (also referred to as isoforms or isotypes) in eukaryotic genomes has historically suggested three hypotheses, which can also be applied to other families of paralogous proteins. First, multiple genes may be present to ensure a stable and redundant pool of

tubulin; subtle sequence changes are simply due to post-duplication genetic drift. Second, particular tubulins may be required for specific environmental conditions and are expressed in response to these conditions. Third, the subtle sequence changes of the monomers impart functional specificity to tubulin dimers, which can optimize the function of the microtubule cytoskeleton for its role in a particular cell type or developmental context (Cleveland, 1987; Wade, 2007; Wilson and Borisy, 1997). These explanations are not mutually exclusive as genetic and cytological evidence support redundancy and specificity among the members of the tubulin superfamily in *C. elegans* and other metazoans.

Worm tubulins have been studied for thirty years in multiple cellular and developmental contexts. *C. elegans* research made early contributions to establishing a linkage between mechanosensation and microtubule structure/function (Chalfie and Thomson, 1982; Fukushige et al., 1999). These pioneering genetic studies mostly supported the notion that certain tubulins play specific biological roles. Further support for this came from the early observation that certain monoclonal tubulin antibodies labeled specific subsets of neuronal cells (Siddiqui et al., 1989). More recently, non-motile primary cilia at the tip of the dendrite of many sensory neurons have provided a model to study intraflagellar transport processes (see WormBook chapter [The sensory cilia of *Caenorhabditis elegans*](#); Müller et al., 2011), and they give another example of the use of specific tubulins to optimize behavioral responses (Hao et al., 2011; Hurd et al., 2010).

As a core component of the metazoan cytoskeletal system, microtubules composed of the less diverged, more broadly expressed, and likely more redundant tubulins play numerous and essential roles at all phases of the cell and life cycle. The early *C. elegans* embryo has become a model to study particular processes such as the assembly and function of the spindle (see [Figure 4](#) and WormBook chapter [Cell division](#); Müller-Reichert et al., 2010; Lu and Mains, 2005; Lu et al., 2004; Cheeranbathur et al., 2013), microtubule dynamics (Bajaj and Srayko, 2013), or the accumulation of spindle proteins after nuclear envelope breakdown (Hayashi et al., 2012). In addition, the early embryo is also a venue to study the role of microtubules in the developmental processes of establishing polarity through asymmetric cell divisions and axis formation, although it is disputed whether centrosomes or the microtubules that emanate from them are the initial symmetry breaking entities (for a history of the evidence both ways, see Wallenfang and Seydoux, 2000; Cowan and Hyman, 2004; Tsai and Ahringer, 2007; Motegi et al. 2011; Bienkowska and Cowan, 2012).

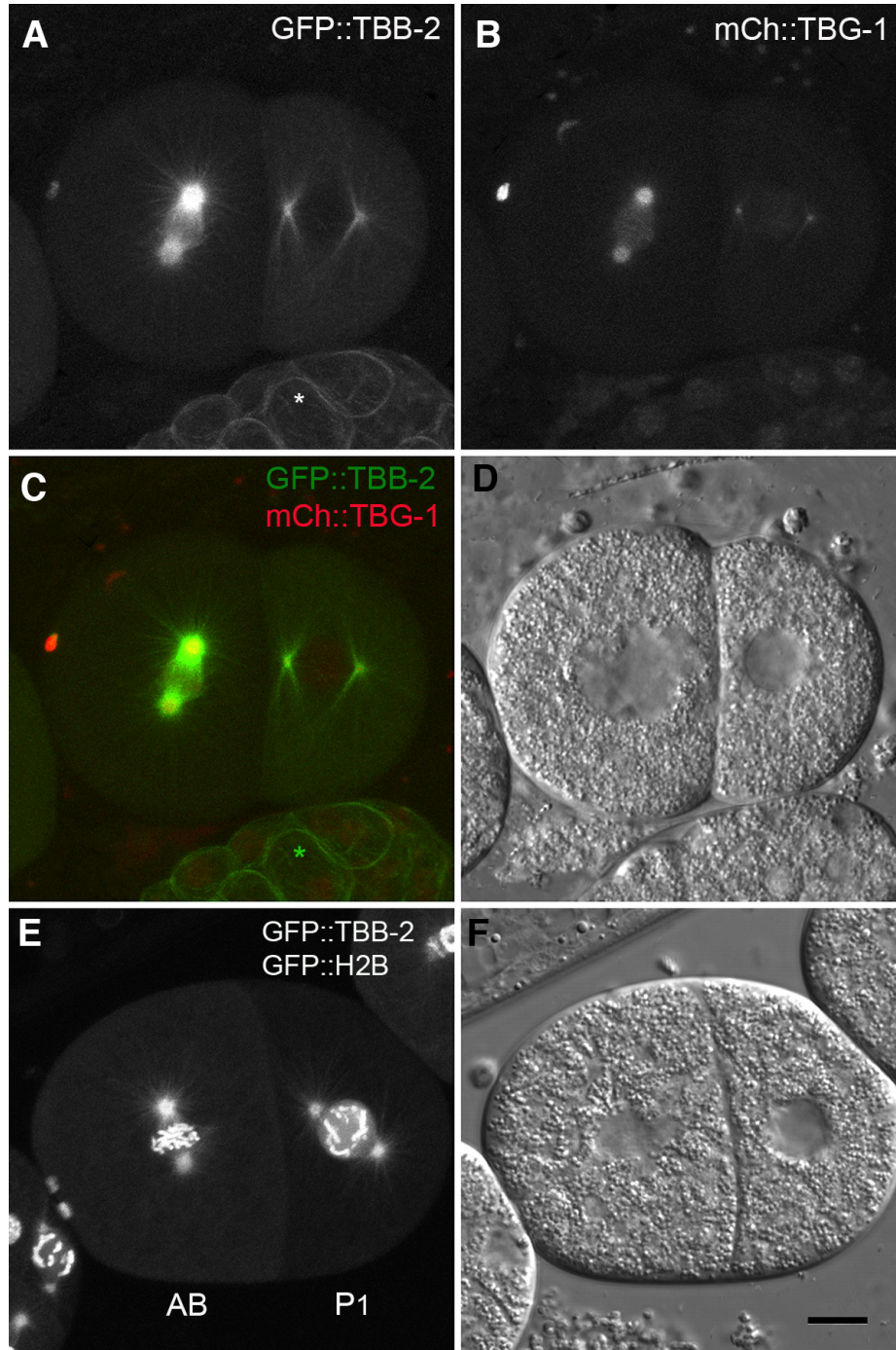


Figure 4. Translational fusions to the promoter of *pie-1* allow for live visualization of tubulins and other components of mitosis in embryonic blastomeres. Strain SA250 contains GFP::TBB-2 as shown in panel A and mCh::TBG-1 in panel B, along with mCh::HIS-48 (histone 48), which appears more strongly in later blastomeres (Toya et al., 2010). A merge of A and B is shown in panel C, and a DIC image is shown in panel D. Strain TY3558 contains GFP::TBB-2 and GFP::H2B (histone 2B) as shown in panel E, with corresponding DIC image in panel F (this strain is not referenced). In all panels, anterior is to the left. Note the asymmetric distribution of one or both of these components at the two-cell stage as seen by a brighter cytoplasm in AB relative to P1 (panel E) and that interphase blastomeres in older embryos show a cortical tubulin network (asterisks in panels A and C). Both strains are available at the *Caenorhabditis* Genetics Center (CGC). Scale bar is 10 μ m.

Evidence for both redundancy and specificity among *C. elegans* tubulins has come from studies of the tubulins that function in somatic tissues in processes such as neuronal polarity, synaptogenesis, and plasticity (Baran et al., 2010; Ou and Shen, 2011; Driscoll et al., 1989; Kurup et al., 2015). The broadly expressed tubulins, perhaps with some contribution from the more specialized family members, likely function as the predominant microtubule tracks

for the movement, positioning, and anchoring of intracellular membranous organelles such as yolk granules (McNally et al., 2010) and nuclei (Tapley and Starr, 2013; see [Section 4.3.15](#) for description of *NOCA-1*). In a process involving the polarity protein *PAR-3* microtubules and components of centrosomes relocate during establishment of intestinal epithelial polarity (Feldman and Priess, 2012). Under control of the formin *EXC-6* (*excretory cell* abnormal) microtubules help organize tubulogenesis (Shaye and Greenwald, 2015). In the absence of centrosomes, microtubules can polymerize from and help remodel epidermal junctions, where they function alongside the *LET-502* (Rho kinase) pathway during embryonic elongation (Quintin et al., 2016; Wang et al. 2015). Microtubules also form the tracks for dynein-based movement of chromosomes during oocyte cell division (Muscat et al., 2015). During differentiation of reproductive muscles microtubules undergo regulated changes in their behavior that are controlled by regulatory proteins and/or compartmentation of soluble monomer (Lacroix et al., 2014; Lacroix et al., 2016).

2.1. The mitotic and neuronal tubulins: *TBA-1*, *TBA-2*, *TBB-1*, *TBB-2*, and *BEN-1*

Two α - (*TBA-1* and *TBA-2*) and two β - (*TBB-1* and *TBB-2*) tubulins show some degree of redundancy and are major components of the mitotic spindle microtubules and of the axonal microtubules in motor neurons. A third β -tubulin, *BEN-1*, functions redundantly in neurons.

2.1.1. *TBA-1* and *TBA-2*

TBA-1 is expressed in the germline and is involved in meiotic and mitotic spindle assembly in the early embryo. It is partially redundant with *TBA-2* as RNAi depletion of both results in embryonic lethality while RNAi or deletion of either causes little effect on embryonic viability (Lu and Mains, 2005; Phillips et al., 2004; Honda et al. 2017). *TBA-1* is also broadly expressed in excitatory motor neurons ([Table 2](#), [Section 6.2](#)), where it functions with *DLK-1* (dual leucine zipper kinase), a mitogen-activated protein kinase kinase kinase, in synaptogenesis and remodeling events (Kurup et al., 2015). Dominant or semi-dominant missense alleles, but not deletion alleles, cause defects in cell division events and neuronal function. These molecular and cellular defects underlie embryonic lethality (O'Rourke et al. 2011) and uncoordination, respectively (Baran et al., 2010). Suppressors of the neuronal phenotypes caused by the combination of a dominant allele of *TBA-1* and loss of *DLK-1* include a neomorphic allele of *TBB-2* and novel missense alleles of kinesin, dynein/dynactin, and the casein kinase *TBK-3* (tau tubulin kinase) (Kurup et al. 2017).

The primary structure of *TBA-2* is very closely related to that of *TBA-1* (98.5% identical, 99.5% similar), which likely provides the basis for redundancy. Sequence similarity extends through their CTTs, which are also the most similar to α -CTTs of other organisms. *TBA-2* is expressed in the germline and in certain neurons ([Table 2](#), [Section 6.2](#)), overlapping with *TBA-1* in the *VB* and *DB* motor neurons and the *PLM* sensory neurons (Fukushige et al., 1993). Like *TBA-1*, *TBA-2* functions in the mitotic spindle of the early embryo as dominant missense alleles cause embryonic lethality (Lu and Mains, 2005; Phillips et al., 2004). Both *TBA-1* and *TBA-2* are also expressed in touch receptor neurons, but they do not seem to be required for the structure or function of those cells (Lockhead et al. 2016). However, minor sequence differences between *TBA-1* and *TBA-2*, which are mostly at the amino-terminus, result in differential sensitivity to the actions of the microtubule severing protein, katanin (*MEI-1/MEI-2*) and in differences in microtubule dynamics in the early embryo (Lu and Mains, 2005; Honda et al. 2017).

2.1.2. *TBB-1* and *TBB-2*

Like the broadly expressed α -tubulins, *TBB-1* and *TBB-2* (97.6% identical and 98.9% similar) both function in a partially redundant way in the mitotic spindle in embryonic blastomeres, and they interact differently with *MEI-1/MEI-2* katanin (Lu et al., 2004; Wright and Hunter, 2003). Similarly, they are partially redundant in motor neurons as loss of either causes mild axonal outgrowth and synaptic defects similar to the dominant mutation of *tba-1* (loss of both is lethal). However, loss of either enhances the defects caused by a dominant allele of *tba-1* (Baran et al., 2010). Both are expressed in many unidentified neurons and other tissues, but rarely does loss of function of one cause any significant defect (Lockhead et al. 2016).

TBB-2 functions during microtubule-dependent processes in the early embryo such as spindle placement and rotation, where dominant missense alleles cause stronger defects than do null alleles (Ellis et al., 2004; Wright and Hunter, 2003, O'Rourke et al., 2011). Loss of *TBB-2* causes an up-regulation of *TBB-1* in the early embryo, providing a genetic mechanism for redundancy (Ellis et al., 2004). Quantitative expression analysis in the early embryo indicates that *TBB-2* is more prevalent than *TBB-1*, and substitution experiments suggest that the particular tubulins present and their concentration contribute to diverse microtubule behaviors (Honda et al. 2017).

Large-scale RNAi screens indicate that **TBB-2** is also required for P-granule localization (Updike and Strome, 2009). Along with **TBA-2**, **TBB-2** is required for correct sexual specification of the male gonad (Kalis et al., 2010). It is broadly expressed in many somatic tissues and the germline (Hunt-Newbury et al., 2007; Lu et al., 2004; McKay et al., 2003), and regulated by the formin **EXC-6**, it also plays a role in excretory cell morphogenesis (Shaye and Greenwald, 2015).

2.1.3. BEN-1

Benomyl is an anti-mitotic benzimidazole compound that acts to inhibit dynamic instability of microtubules (Gupta et al., 2004; Singh et al., 2008). When wild-type *C. elegans* are reared in its presence, growth is slowed, movement is uncoordinated, and fewer neuronal processes emanate from ventral cord motor neurons. The *ben-1* locus was identified in a screen for mutations that offer resistance to benomyl (Driscoll et al., 1989). Deletion of *ben-1* and certain missense mutations confer resistance while causing no other gross abnormalities. Mutations that cause either resistance or hypersensitivity to benomyl have been linked to a β -tubulin in both nematodes and fungi (Nakaune and Nakano, 2007). *ben-1* is broadly expressed in the nervous system (Figure 5), which supports the idea that it is involved in the neuronal control of locomotion. However, no cytological phenotypes have been reported. The structure of the CTT of **BEN-1** is similar to **TBB-4** (Figure 3), although it lacks a recognizable axonemal motif (see Section 2.2.4).

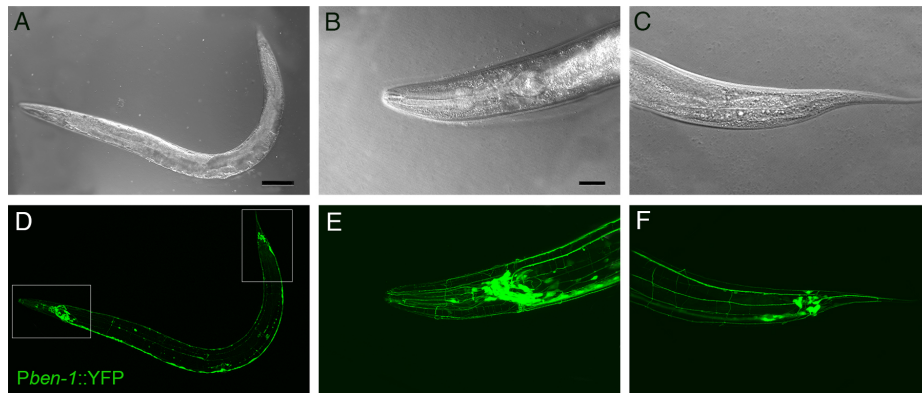


Figure 5. *ben-1* is expressed in multiple neurons. A transcriptional fusion (after Boulin et al., 2006) of the *ben-1* promoter to yellow fluorescent protein (YFP, from pPD136.64/L4817 in the Fire Lab vector kit 1999) reveals broad expression in sensory, motor, and likely interneurons. Scale bars = 100 μ m in panels A and D and 10 μ m in B, C, E, and F. Boxes in D indicate views shown in E and F. Strain to be available at the CGC.

2.2. The sensory neuron axonemal tubulins: **TBA-5**, **TBA-6**, **TBA-9**, and **TBB-4**

C. elegans sense their environment with an array of chemo- and mechanosensory neurons such as the anterior amphid neurons and posterior phasmid neurons. The non-motile cilia at the tips of the dendrites in these neurons are built on a 9+0 axoneme structure, and they have become a model in which to study homologs of the ciliopathy genes in humans (see WormBook chapter *The sensory cilia of *Caenorhabditis elegans**;

2.2.1. TBA-5

TBA-5 is an axonemal α -tubulin expressed in amphid and phasmid sensory neurons where it is localized to cilia. It was isolated in a screen to identify mutants in ciliary dye-filling. A missense allele causes a **Dyf** (dye-filling defective) phenotype and shortened cilia while loss-of-function (deletion) mutations do not show defects in dye-filling (Hao et al., 2011). **TBA-5**, **TBB-4**, and perhaps additional axonemal tubulins are transported by intraflagellar transport (IFT) mechanisms involving a number of microtubule motor proteins (Hao et al., 2011). No behavioral defects have been associated with the perturbation of *tba-5*.

2.2.2. TBA-6

TBA-6 was identified as a ciliary tubulin by combining serial analysis of gene expression (SAGE) and microarray datasets. It is co-expressed with **PKD-2** (a polycystin) in ciliated, male-specific sensory neurons (CEMs and RnBs) in addition to inner labial neurons found in both sexes. Mutant analysis indicated that it is required, along with **TBB-4**, for the normal structure of **CEM** cilia and the normal localization of **PKD-2** to the distal tips of RnB

cilia (Hurd et al., 2010). *TBA-6* is also required for multiple aspects of the specialized structure and function of male-specific CEM neurons including axoneme ultrastructure, motor protein velocity, and extracellular vesicle function (Silva et al., 2017). The combination of these abnormalities likely underlies the male mating behavior defects shown by mutants (Hurd et al., 2010). *TBA-6* mRNA is presumably expressed in the germline as it is found in the one-cell embryo. It is also expressed in other somatic tissues (Baleanu-Gogonea and Siddiqui, 2000), but deletion mutants are fertile and without obvious morphological abnormalities. Structurally, *TBA-6* lacks a recognizable α -tubulin axonemal CTT found in other ciliary α -tubulins.

2.2.3. *TBA-9*

TBA-9 was also identified as a candidate ciliary α -tubulin by combining SAGE and microarray datasets (Hurd et al., 2010). It is expressed in many ciliated sensory neurons in the head of both sexes, often with *TBB-4* (Table 2, Section 6.2). It is required for many normal behaviors including male mating, retraction after nose touch, locomotory rate/posture, exploratory behavior, and the basal slowing response upon contact with food. Deletion causes only subtle defects in the localization of signaling proteins (D. D. Hurd, unpublished results), although it is required for the normal distribution of *TBB-4*. Structurally, it lacks a recognizable axonemal CTT found in other ciliary α -tubulins (Hurd et al., 2010).

2.2.4. *TBB-4*

TBB-4 is a broadly expressed ciliary β -tubulin orthologous to ciliary β -tubulins in other species (Hao et al., 2011). It has a recognizable axonemal motif in its CTT, and it is expressed in many, but not all ciliated sensory neurons, including certain amphid, certain labial, and both classes of male tail ray neurons (Table 2, Section 6.2) where it is required for the normal localization of signaling proteins, such as *PKD-2*. *TBB-4* is required for normal male mating and nose-touch behaviors (Hurd et al., 2010). It is enriched in the ciliary axoneme, and is transported by intraflagellar transport. Some missense mutations are more severe than null (deletion) mutations (Hao et al., 2011).

2.3. The mechanosensory tubulins: *MEC-12* and *MEC-7*

The *MEC-12* α -tubulin and the *MEC-7* β -tubulin were originally discovered in screens for mutants that did not respond to mechanical stimulation along the body (Chalfie and Sulston, 1981). They are both expressed in the six body wall touch receptor neurons (TRNs) sometimes referred to as the microtubule cells (*ALML/R*, *PLML/R*, *AVM*, and *PVM*). These sensory neurons contain cross-linked 15-protofilament microtubules that terminate near the plasma membrane (Chalfie and Thomson, 1982). *MEC-12* is the only *C. elegans* α -tubulin with a lysine as aa40, a position that is frequently modified via post-translational acetylation (see Section 4.2.1). Antibodies to acetylated tubulin strongly label the touch receptor cells (Akella et al., 2010; Shida et al., 2010; Siddiqui et al., 1989).

MEC-12 and *MEC-7* are required for the 15 protofilament microtubules in the TRNs, for mechanoreceptor currents, and for touch sensitivity (Bounoutas et al., 2009; O'Hagan et al., 2005). Mutations often cause the appearance of 11 protofilament microtubules, which can alter the distribution and/or expression of the components of the mechanosensation signal transduction pathway (Bounoutas et al., 2011; Bounoutas et al., 2009; Huang et al., 1995). Mutations in these two tubulins that cause the most severe defects are not null, but rather dominant or semi-dominant missense (antimorphic or neomorphic) alleles (Hsu et al., 2014; Savage et al., 1994; Kirszenblat et al., 2013). Loss of *PTL-1* (see Section 4.3.1) causes an enhancement of touch insensitivity in double mutant combinations with certain alleles of *mec-12* or *mec-7* (Gordon et al., 2008).

TBA-7 is also expressed in the TRNs, but is not strictly required for their behavioral function nor their characteristic 15 protofilament microtubules. Instead it plays a role in microtubule dynamics in these cells as loss of function increases stability, causes ectopic neurite growth and resistance to cochicine in a manner similar to *KLP-7* (Zheng et al., 2017).

2.4. α - and β - tubulins with undiscovered roles

2.4.1. *TBA-4*

TBA-4 has not been studied in detail. Data from RNAi experiments have suggested that *TBA-4* may play a role in embryonic development or larval growth (Simmer et al., 2003). Its anatomic expression resembles that of *tba-7* (see Table 2, Section 6.2, and Figure 6).

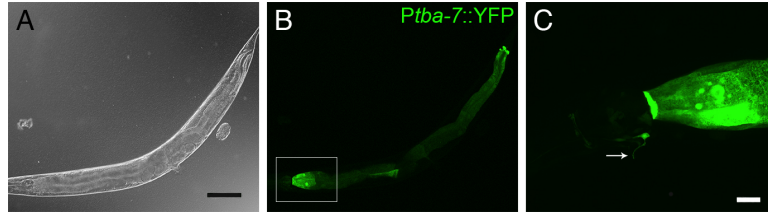


Figure 6. *tba-7* is primarily expressed in the intestine. A transcriptional fusion of the *tba-7* promoter to YFP reveals expression predominantly in the gut (see Table 2). It is also expressed in the pore cell of the excretory system (arrow in panel C, which is a high magnification view of the junction of the posterior pharynx and anterior intestine). Scale bars = 100 μ m in panel A and 10 μ m in B and C. Box in panel B indicates view in panel C. Strain to be available at the CGC.

2.4.2. TBA-8

The primary structure of TBA-8 is notable for its diverged CTT that has a phenylalanine as the last amino acid (Figure 3). Nearly all other α -tubulins have a tyrosine that is often removed after translation. Transcriptional reporters indicate that it is expressed in seam cells and unidentified neurons (D. D. Hurd and D. S. Portman, unpublished results). No loss of function analysis has been undertaken.

2.4.3. TBB-6

TBB-6 is the most structurally diverged β -tubulin as it lacks most of what is considered to be a CTT (typically acidic amino acids, see Figure 3). It appears to have arisen recently in the *Caenorhabditis* lineage (Saunders et al. 2013). Its anatomic expression closely resembles that of *tba-7* (Table 2, Section 6.2). Neither a deletion mutation nor RNAi depletion causes an overt phenotype (D. D. Hurd, unpublished results).

3. Beyond α - and β -tubulin

3.1. TBG-1

γ -tubulin functions in the nucleation and anchoring of microtubules. Antibody staining and translational GFP fusions show that γ -tubulin is a component of centrosomes (Bobinnec et al., 2000; Strome et al., 2001). It is also found at the base of cilia in amphid and phasmid sensory neurons and the apical cortex in the intestinal epithelium (Bobinnec et al., 2000). RNAi depletion of γ -tubulin causes polyploid germline cells and early blastomeres that contain a disorganized, ineffective mitotic spindle (Bobinnec et al., 2000; Strome et al., 2001). Too much γ -tubulin at the centrosome is caused by loss of the RNA-binding protein ATX-2 (*ataxin-2*), and this also brings about multiple defects in cell division (Stubenvoll et al., 2016). γ -tubulin is not absolutely required for microtubule nucleation, but instead is required for proper placement of microtubule minus ends within centrosomes and normal centriole structure (O'Toole et al., 2012).

3.2. Other tubulins

The *C. elegans* genome does not contain orthologs of other tubulins (δ -, ϵ - or η -tubulin), which are found in some single-celled eukaryotes and vertebrates. These tubulins are perhaps involved in basal body and centriole function (Dutcher, 2003; Oakley, 2000).

4. Cellular control of microtubules

Microtubules underlie a variety of essential cellular processes and are regulated by diverse mechanisms at many levels including biogenesis, heterodimer formation, nucleation, polymerization, depolymerization, post-translational modification, and association with other proteins (see Table 3, Section 6.3).

4.1. Tubulin biogenesis and homeostasis

Newly synthesized tubulin (and actin) monomers pass through a series of chaperone complexes prior to being incorporated into dimers and eventually microtubules. The first of these is the prefoldin complex, which accepts nascent tubulin monomers from the ribosome and delivers them to the CCT (chaperonin-containing TCP-1) complex. The CCT complex finalizes folding of monomers and presents them to the tubulin folding cofactors A-E for dimerization. Tubulin degradation is thought to be mediated by the ubiquitin-proteasome pathway and other regulators/sensors of tubulin homeostasis, such as parkin or cofactor E-like proteins (Lundin et al., 2010).

PFD-1 through PFD-6 are subunits of the prefoldin complex, and CCT-1 through CCT-8 compose the CCT complex. Both transgenic fusion proteins and antibody staining indicate that they are expressed in nearly all cells with the most intense expression (of transgene) in body wall muscle, pharyngeal muscle, vulval muscle, hypodermal cells, and the somatic gonad. Inhibition by RNAi in the early embryo indicates that prefoldin, CCT, and TXDC-9 function are required for multiple events in the early embryo that are dependent on microtubule function in addition to distal tip cell migration (Leroux and Candido, 1997; Lundin et al., 2008; Ogawa et al., 2004; Srayko et al., 2005). Post-embryonic RNAi depletion of *cct-5* has shown that it is required for proper levels of tubulin and normal organization of intestinal actin and microvilli (Saegusa et al., 2014). The tubulin folding cofactors have not been studied in detail. Parkin has been implicated in the ubiquitination of tubulin in other systems, but the role for the broadly expressed *C. elegans* ortholog (PRD-1) in tubulin homeostasis has not been investigated (Springer et al., 2005).

4.2. Post-translational modification

Eukaryotic tubulins can be covalently modified after translation in a number of ways. These include tubulin-specific modifications such as removal/replacement of a terminal tyrosine (α -tubulin), addition of an acetyl group to an amino-terminal lysine (α -), addition of glycines or glutamates to residues in the CTT (both α - and β -). Tubulins also undergo more general modifications such as phosphorylation/dephosphorylation (β -) and palmitoylation (α -). The most intensely studied modifications are those that are unique to tubulins (Hammond et al., 2008; Janke and Bulinski, 2011).

4.2.1. Acetylation

MEC-12 is the only α -tubulin with a lysine at position 40, suggesting that it is the only acetylated α -tubulin in *C. elegans*. Supporting this idea, antibodies that recognize acetylated tubulin intensely stain the body wall TRNs, and neurons implicated in nose mechanosensation that express MEC-12 (Shida et al., 2010; Siddiqui et al., 1989).

Acetylation is accomplished by two α -tubulin acetyltransferases (aTATs), MEC-17 and ATAT-2. They are expressed in TRNs and are required for acetylation of tubulin on the luminal side of the microtubule wall in these cells. Acetylated α -tubulin is required for optimal touch sensation (Akella et al., 2010; Shida et al., 2010) and the structure and mechanical properties of the specialized 15 protofilament microtubules found in touch receptor neurons, although MEC-17 might play an additional role in microtubule structure (Cueva et al., 2012; Topalidou et al., 2012, Davenport et al., 2014).

4.2.2. Glutamylolation/deglutamylolation

Tubulins are glutamylated by tubulin tyrosine ligating enzymes (TTLLs). Some members of this family initiate while some elongate chains of glutamate. These chains are added to a glutamate residue in the primary structure of tubulin CTTs. TTLL-4 glutamylates microtubules in the amphid channel cilia and certain labial cilia (See Table 3, Section 6.3). Individual TTLL mutations cause little overt phenotypic effect, but a triple mutant of TTLL-4, TTLL-5, and TTLL-11 eliminates glutamylolation and causes reduced male mating efficiency (Chawla et al., 2016).

Certain members of the cytosolic carboxypeptidase (CCPP) family remove glutamate chains from tubulin. Both CCPP-1 and CCPP-6 have been shown to suppress poly- or hyper-glutamylolation (Kimura et al., 2010; O'Hagan et al., 2011). Proper levels of glutamylolation are required for axonemal microtubule ultrastructure, normal velocity of motor proteins, localization of signaling proteins, and certain behaviors that require input from ciliated neurons (O'Hagan et al., 2011).

4.3. Microtubule-associated proteins (MAPs)

Microtubules are found in their numerous cellular contexts in either highly dynamic and transient structures, such as the spindle during mitosis, more stable structures, such as the axoneme supporting a cilia, or in variably stable structures, such as neuronal microtubules during development or after injury. These higher order structures and/or changes in dynamic behavior are regulated by a collection of microtubule-associated proteins that can promote growth/stabilize, promote shrinkage/destabilize, sever, bundle, bind to plus or minus ends, or perform some combination of these functions.

4.3.1. PTL-1

tau and other similar MAPs bind to and stabilize microtubules. In vertebrates, dysfunction of *tau* causes disassociation from microtubules and self-aggregation, which underlies a broad class of neuronal pathologies known

collectively as tauopathies (Sabbagh and Dickey, 2016). *C. elegans* has a single identified protein that is similar to mammalian *tau* (Goedert et al., 1996; McDermott et al., 1996). PTL-1 (protein with *tau*-like repeats) is most highly conserved in both structure and function to *tau* and other MAPs in the carboxyl-terminal microtubule binding repeats (Hashi et al., 2016; Gordon et al., 2008). It is most prominently expressed in mechanoreceptor cells. Loss of PTL-1 causes no overt phenotypic effect, but *ptl-1* mutations dominantly enhance the loss of touch sensation caused by lack of either MEC-7 or MEC-12 (Goedert et al., 1996; Gordon et al., 2008). Loss of both PTL-1 and spectrin (UNC-70) also causes a rearrangement of microtubule bundles, which sensitizes TRN neurites to mechanical damage (Krieg et al. 2017). PTL-1 is required for the normal behavior of microtubule motor proteins and the distribution of synaptic components in touch receptor cells (Tien et al., 2010).

4.3.2. ELP-1

EMAP (echinoderm microtubule-associated protein) family members bind to and can either stabilize (Houtman et al., 2007) or destabilize microtubules (Eichenmüller et al., 2002). Consensus domains across family members include ~6 WD40 repeats and another domain specific to the EMAP-like proteins, the ELP domain. *C. elegans* has a single member of the family, ELP-6 (Suprenant et al., 2000), which has a predicted amino terminal coiled domain found in some of the vertebrate orthologs (Hueston et al., 2008). It has been shown to bind to microtubules and is expressed in a number of cell types including body wall muscle, sensory neurons (IL1, male ray neurons, TRNs), vulval muscles, the spermathecal valve cell, intestinal cells, and seam cells (Hueston et al., 2008). Loss of ELP-1 through mutation or RNAi causes defects in touch reception, and in combination with loss of the dystrophin homolog DYS-1, causes paralysis and death (Hueston and Suprenant, 2009).

4.3.3. TAC-1

The transforming acidic coiled-coil (TACC-1) family of proteins functions in the regulation of microtubule length by promoting growth (Mortuza et al, 2014). The *C. elegans* protein TAC-1 consists solely of the TACC-1 domain, while orthologous proteins have more complex and larger structures (Bellanger and Gönczy, 2003; Le Bot et al., 2003). TAC-1 accumulates in the centrosomes of dividing blastomeres (dependent upon ZYG-9) where it physically associates and functions with ZYG-9 and ZYG-8 to regulate microtubule length, pronuclear migration, and spindle positioning as a polymerizing factor (Bellanger et al., 2007; Bellanger and Gönczy, 2003; Srayko et al., 2003).

4.3.4. ZYG-8

Doublecortin kinases bind to microtubules to stabilize and/or straighten them (Jean et al., 2012). ZYG-8 is the ortholog of the human protein DCLK (doublecortin-like kinase), as it contains a doublecortin domain and a kinase domain similar to calcium/calmodulin-regulated kinases. ZYG-8 localizes to microtubules in the mitotic spindle in early blastomeres (and in cultured mammalian cells), where it functions to promote microtubule growth required for proper spindle placement (Gönczy et al., 2001). It is also expressed in certain neurons where it is required for process outgrowth, locomotion, and touch sensitivity (Bellanger et al., 2012). ZYG-8 physically associates with TAC-1 through its doublecortin domain (Bellanger et al., 2007) and with EFA-6 during regeneration after axonal injury (see Section 4.3.8).

4.3.5. ZYG-9

The XMAP215 (*Xenopus* microtubule-associated protein) family of microtubule binding proteins is composed of three TOG (tumor overexpressed gene) domains that interact with tubulin dimers and promote assembly (Al-Bassam and Chang, 2011). ZYG-9 is the *C. elegans* member, originally discovered in screens for mutations that alter polarity in the early embryo (Kemphues et al., 1986). It was subsequently shown to be required for spindle function (Matthews et al., 1998). ZYG-9 localizes to the centrosome in dividing blastomeres (dependent upon TAC-1), where it functions with ZYG-8 and TAC-1 to regulate the length of microtubules (Bellanger et al., 2007; Bellanger and Gönczy, 2003; Srayko et al., 2003).

4.3.6. CLS-1, CLS-2, and CLS-3

The CLASP (cytoplasmic linker associated protein) family of microtubule regulators also contains TOG domains. They function in microtubule stabilization by promoting rescue of shrinking microtubules (Al-Bassam and Chang, 2011). CLS-1, CLS-2, and CLS-3 are the members of the CLASP family in *C. elegans*. RNAi depletion causes defects in nuclear rotation, maintenance of spindle length, and spindle placement in the one-cell embryo, likely due to fewer astral microtubules. CLS-2 is found on the mitotic spindle, centrosomes, and kinetochores in dividing cells in early embryogenesis and in the cytoplasm of interphase cells (Espirito et al, 2012).

4.3.7. CHE-12

Crescerins (*crescere*: to grow) are another family of TOG domain proteins that bind to and regulate microtubules in cilia. This family is represented by CHE-12 in *C. elegans*, and is part of a larger family that includes the CLASPs (CLS-1, -2 and -3) and ch-TOG/XMAP215 (ZYG-9) proteins. CHE-12 localizes to the cilia of the amphid channel neurons and the phasmid neurons. Loss of function through mutation causes altered cilia structure, inability to take up lipophilic dyes, and failure in chemotaxis (Bacaj et al., 2008; Das et al., 2015).

4.3.8. EFA-6

Members of the EFA-6 (exchange factor for ARF6 family GTPases) protein family function to limit microtubule growth. These proteins are composed of a Sec7 guanine nucleotide exchange factor domain, a pleckstrin homology (PH) domain, and regions of coiled-coil. *C. elegans* EFA-6 is found in the cortex of blastomeres where loss of function through mutation or RNAi treatment causes an increase in the total number of microtubules, an increase in the fraction of the population that is growing, and an increase in microtubule length (Chen et al., 2011; O'Rourke et al., 2010). EFA-6 is also found dynamically rearranging from a smooth distribution to punctae of microtubule minus ends during axotomy-induced regeneration in PLMs. This is controlled by an 18 amino acid N-terminal motif through which it also makes physical association with TAC-1 and ZYG-8. Loss-of-function mutations cause neurite overgrowth after injury (Chen et al., 2015).

4.3.9. RMD-1, RMD-2, RMD-3, RMD-4, RMD-5, and RMD-6

RMD-1 through -6 is a family of proteins that might function in the regulation of microtubule dynamics. They all contain predicted coiled-coil domains and have orthologs in vertebrates, although little is known about the vertebrate homologs. Only RMD-1 has been studied in detail; it appears to function in the attachment of microtubules to kinetochores as depletion through RNAi causes delayed anaphase and incorrectly segregated DNA during mitosis. RMD-1 localizes to the spindle of the one-cell embryo and binds to microtubules *in vitro* (Oishi et al., 2007).

4.3.10. MEI-1 and MEI-2

Katanins are a family of microtubule severing proteins (Roll-Mecak and McNally, 2010). MEI-1 is the p60 catalytic subunit (AAA ATPase) of *C. elegans* katanin, and MEI-2 is the p80 activator/regulator of MEI-1. They were originally identified in screens for defects in meiosis and mitosis (Clark-Maguire and Mains, 1994; Mains et al., 1990). They are maternally expressed and dependently colocalize in the meiotic spindle of oocytes (Srayko et al., 2000). It was originally thought that microtubule severing activity was required for functional meiotic spindles (Srayko et al., 2006), but recent analysis of specific point mutations suggests that MEI-1 might also bind/bundle microtubules (McNally and McNally, 2011; Connolly et al., 2014; McNally et al., 2014). Levels of MEI-1 decrease after meiosis due to the activity of the MEL-26 E3 ubiquitin ligase (Johnson et al., 2009; Pintard et al., 2003).

4.3.11. SPAS-1

Like katanins, spastins are microtubule severing proteins in the AAA ATPase family of enzymes (Roll-Mecak and McNally, 2010). SPAS-1 was identified via homology in a search for proteins related to spastin, the human protein that is defective in a form of spastic paraplegia. SPAS-1 is reportedly expressed in a broad variety of tissues in worms. A precise *in vivo* function is not evident from studies of a deletion mutation, which causes pleiotropic defects in growth, vulval development, and reproduction (Matsushita-Ishiodori et al., 2007). Biochemically, SPAS-1 forms hexamers, and its ATPase activity is stimulated by microtubules. Conserved basic amino acids interact with acidic CTTs of tubulin, and a conserved tryptophan in SPAS-1 is required for microtubule severing (Matsushita-Ishiodori et al., 2009).

4.3.12. KLP-7

KLP-7 is a kinesin-like protein of the MCAK (mitotic centromere-associated kinesin) family, which functions to depolymerize microtubules in many systems (Sanhaji et al., 2011). It is found at the kinetochore in the blastomeres of the early embryo (Oegema et al., 2001). KLP-7 is phosphorylated by Aurora kinases, and it regulates nucleation rate; loss of *klp-7* expression through RNAi or mutation increases the number of astral and midzone microtubules while decreasing growth rate (Srayko et al., 2005; Han et al., 2015). KLP-7 also functions to establish a bipolar meiotic spindle in the absence of centrosomes (Connolly et al., 2015). In adult touch receptor neurons,

KLP-7 functions to maintain a dynamic population of microtubules in normal cells and to inhibit microtubule upregulation in axotomized cells (Ghosh-Roy et al. 2012).

4.3.13. EBP-1, EBP-2, and EBP-3

Microtubule end-binding proteins (also known as tip-interacting proteins or +TIPs) recognize and bind to a structural feature at the plus ends of microtubules (Jiang and Akhmanova, 2011). EBP-1 and EBP-2 fusion proteins (driven by the *pie-1* promoter) decorate the growing plus ends of microtubules in the spindle of early embryos (Motegi et al., 2006; Srayko et al., 2005). EBP-1 is required for efficient axon regrowth in PLM neurons after laser axotomy (Chen et al., 2011). Perturbation by mutation or RNAi does not cause severe phenotypes, perhaps due to redundancy, and analysis of anatomic expression from native promoters for these three paralogs has not been reported.

4.3.14. PTRN-1

Members of the CAMSAP (calmodulin-regulated spectrin-associated protein)/Nezha/patronin family decorate and stabilize minus ends of microtubules (Akhmanova and Hoogenraad, 2015). The proteins share a conserved structure, which includes a calponin homology domain, a coiled-coil domain(s), the namesake CAMSAP (also called CKK) domain, and a carboxyl-terminal microtubule binding domain. PTRN-1 in *C. elegans* is broadly expressed in neurons and has been shown to be associated with microtubules in puncta in nerve and muscle cells (Marcette et al. 2014; Richardson et al., 2014; Chuang et al. 2014). It is not required in general for normal movement, sensation, or reproduction. The effects of loss-of-function mutations, such as overgrowth or ectopic neurite formation, are more readily observed in microtubule-sensitized situations such as exposure to the depolymerizing drug colchicine, or in specific developmental contexts such as neuronal remodeling in touch receptor neurons or after injury. Loss of DLK-1 suppresses the subtle defects caused by loss of PTRN-1, suggesting they may work in the same pathway (Marcette et al. 2014; Richardson et al., 2014; Chuang et al. 2014).

4.3.15. NOCA-1

Ninein proteins function to anchor and perhaps stabilize and/or nucleate microtubules through association with the minus end (Srivatsa et al. 2015). The *C. elegans* homolog of vertebrate ninein-like proteins is NOCA-1, encoded by a complex locus that produces several variants. It was originally identified as a gene that phenocopied γ -tubulin defects. A deletion causes abnormal positioning of germline nuclei and associated sterility, with no other obvious phenotypes. NOCA-1 works with γ -tubulin and PTRN-1 to establish non-centrosomal microtubule arrays that position nuclei and/or provide structural integrity (along with LET-502) to the hypodermis (Quintin et al., 2016). NOCA-1 and γ -tubulin colocalize, and NOCA-1 and PTRN-1 bind to microtubules in extracts (Wang et al. 2015).

4.3.16. CYLC-1 and CYLC-2

Cylicin was originally discovered in the cytoskeletal calyx that surrounds part of the nucleus in the head of mammalian sperm; structurally it is characterized by numerous small repeats of the charged amino acids KKD/E (Hess et al. 1993). *C. elegans* has two highly conserved proteins, CYLC-1 and CYLC-2, that are related to mammalian cylicin and potentially to stathmin. Inhibition by RNAi causes altered tubulin concentrations and misregulated growth of microtubules in a region of uterine muscle cells, suggesting that these proteins function in the intracellular compartmentation of tubulin levels (Lacroix et al., 2016). No anatomic expression pattern has yet been reported.

5. Concluding remarks and future directions

As a genetic and cytological model in which to study a family of closely related structural proteins, *C. elegans* provides evidence for both redundancy and specificity in the use of α - and β -tubulins. In support of redundancy, mutagenesis screening has produced a collection of missense alleles that change conserved amino acids in tubulin (e.g., in the *tba-1*, *tba-5*, *tbb-2*, *mec-12*, *mec-7*, and *tbb-4* genes) that often cause more severe defects than deletions/nulls. This means that it is often more deleterious to incorporate an altered tubulin into a microtubule than it is to incorporate a wild-type version of another tubulin. In addition, genetic and/or RNAi elimination of multiple mitotic tubulins (*tba-1*, *tba-2*, *tbb-1*, and *tbb-2*) is required to disrupt embryogenesis, further supporting redundancy among tubulins. Nevertheless, specific expression patterns (Figure 7), the presence of variant CTTs, and the quantifiable cytological and behavioral defects caused by the loss of single tubulins (e.g., *tba-5*, *tba-6*, *tba-9*, *mec-12*, *tbb-4*, or *mec-7*) all support the idea that tubulins can be specialized for certain developmental and physiological contexts. Whether tubulins in *C. elegans* are regulated in response to environmental change has not been explored.

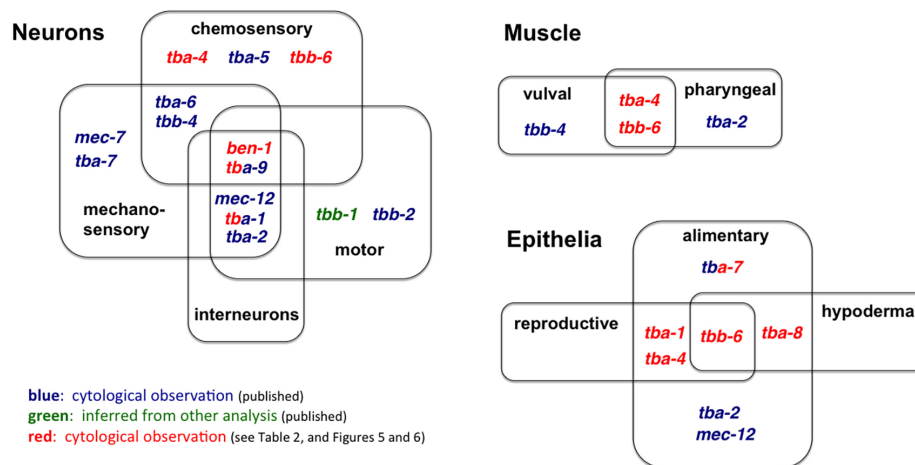


Figure 7. Summary of the somatic expression of the α - and β -tubulins. Venn diagrams depict the anatomic expression of the tubulin family members in non-germline tissues/organs. The type of data used to place a particular tubulin in a particular area is depicted using color. See text for references.

C. elegans provides the ability to understand tubulin biology from the molecular to the organismal level. Worm tubulins will continue to be discovered playing roles in diverse and unforeseen biological processes, whether through *de novo* mutagenesis screening, systems level analysis, or novel approaches. At the molecular level, new alleles can continue to shed insight into the structure and function of the tubulin dimer and the myriad other proteins that interact with tubulin, either soluble or assembled. Both forward and reverse genetic screens and searching for modifiers of existing phenotypes, especially among paralogs expressed in the same tissues and/or with regulatory factors, allows understanding of tubulin usage and regulation within an organism. Technological advances (e.g., studying transport velocities of cellular components in neurons or other tissues), a strength of the nematode model system, should provide a higher resolution and more precise quantitative description of the role of tubulin in metazoan biology.

6. Tables

6.1. Table 1. Tubulin alleles that have been classified genetically, have an identified molecular change, or are represented by a strain at the **CGC**

Gene	Allele	Protein change	Strain	Mutation class	Reference
<i>tba-1</i>	<i>or346</i>	S377F	EU1135	Dominant	Philips et al., 2004; O'Rourke et al., 2011
<i>tba-1</i>	<i>or594</i>	S377F	EU1161	Semi-dominant	O'Rourke et al., 2011
<i>tba-1</i>	<i>ju89</i>	G414R		Dominant	Baran et al., 2010
<i>tba-2</i>	<i>sb116</i>	E22K		Intragenic revertant of <i>sb51</i>	Lu, C. et al., 2005
<i>tba-2</i>	<i>sb117</i>	E69K		Intragenic revertant of <i>sb51</i>	Lu, C. et al., 2005
<i>tba-2</i>	<i>sb51</i>	S168Y	HR505	Dominant	Lu, C. et al., 2005
<i>tba-2</i>	<i>sb27</i>	E194K	HR973	Dominant	Lu, C. et al., 2005
<i>tba-2</i>	<i>sb25</i>	E277K	HR899	Dominant	Lu, C. et al., 2005
<i>mec-12</i>	<i>gk170196</i>	P32S		Recessive, Neomorph	Zheng et al., 2017
<i>mec-12</i>	<i>u76</i>	D69N		Recessive, Antimorph	Fukushige et al., 1999; Zheng et al., 2017
<i>mec-12</i>	<i>u1016</i>	E97K		Recessive, Antimorph	Zheng et al., 2017

Gene	Allele	Protein change	Strain	Mutation class	Reference
<i>mec-12</i>	<i>u950, gk672907</i>	S140F		Recessive, Antimorph	Zheng et al., 2017
<i>mec-12</i>	<i>gk600523</i>	G142E		Recessive	Zheng et al., 2017
<i>mec-12</i>	<i>u1021, e1607</i>	G144S		Semi-dominant (haplo-insufficient), Recessive, Antimorph	Hsu et al., 2014; Bountonas et al. 2011; Bountonas et al., 2009; Zheng et al., 2017
<i>mec-12</i>	<i>u94</i>	G148S		Semi-dominant	Gu et al., 1996; Bounoutas et al., 2009
<i>mec-12</i>	<i>u67</i>	M154I		Temperature-sensitive dominant	Gordon et al., 2008; Bounoutas et al., 2009
<i>mec-12</i>	<i>u204</i>	S178F			Chalfie and Au 1989; Bounoutas et al., 2009
<i>mec-12</i>	<i>u279</i>	T179I			Gu et al., 1996
<i>mec-12</i>	<i>u50, e1605</i>	H192Y	CB3284	Recessive, Loss of function	Bountonas et al., 2009; Bountonas et al., 2011
<i>mec-12</i>	<i>gk915672</i>	E196K		Recessive, Neomorph	Zheng et al., 2017
<i>mec-12</i>	<i>u1041</i>	G246E		Recessive, Loss of function	Zheng et al., 2017
<i>mec-12</i>	<i>u917</i>	V260I		Recessive, Neomorph	Zheng et al., 2017
<i>mec-12</i>	<i>u159</i>	C295Y		Semi-dominant	Gu et al., 1996; Bounoutas et al., 2009
<i>mec-12</i>	<i>gk515972</i>	V323I		Recessive, Neomorph	Zheng et al., 2017
<i>mec-12</i>	<i>u241, u1019</i>	G354E		Dominant, Gain of function, Recessive, Antimorph	Fukushige et al, 1999; Bountonas et al., 2011; Bountonas et al., 2009; Zheng et al., 2017
<i>mec-12</i>	<i>u63</i>	E415K		Recessive, Loss of function	Fukushige et al., 1999; Bounoutas et al., 2011; Zheng et al., 2017
<i>mec-12</i>	<i>gm379</i>	G416E		Semi-dominant, Recessive, Loss of function	Hsu et al., 2014
<i>tba-5</i>	<i>qj14</i>	A19V		Gain-of-Function	Hao et al., 2011
<i>tbb-2</i>	<i>or600</i>	G140E	EU1588	Semi-dominant	O'Rourke et al., 2011
<i>tbb-2</i>	<i>or362</i>	G141E	EU858	Semi-dominant, Dominant	Ellis et al, 2004; Lu et al., 2004
<i>tbb-2</i>	<i>qt1</i>	E198K	HC48	Gain-of-Function	Wright et al., 2003
<i>tbb-2</i>	<i>t1623</i>	V313M	GE2255	Semi-dominant	Wright et al., 2003
<i>tbb-2</i>	<i>sb26</i>	E439K	HR1060	Semi-dominant, Gain of function	Lu, et.al., 2004; Shaye et al., 2015
<i>mec-7</i>	<i>e1506</i>	M1I	CB3270	Recessive	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	<i>u319</i>	S25F		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017

Gene	Allele	Protein change	Strain	Mutation class	Reference
<i>mec-7</i>	u305 , u1020	G34S		Recessive, Loss of function	Savage et al., 1994; Sze et al., 1997; Zheng et al., 2017
<i>mec-7</i>	u58 , u223	P61L		Semi-dominant, Antimorph	Savage et al., 1994
<i>mec-7</i>	u249	P61S		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u430	A97V		Recessive, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u9 , u440	W101stop		Recessive	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u222	G109E		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u429 , u433	G141E		Recessive, Loss of function	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	gk595364	G141R		Recessive, Loss of function	Zheng et al., 2017
<i>mec-7</i>	u275	G148R		Recessive, Loss of function	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	gk895768	T149I		Recessive, Neomorph	Zheng et al., 2017
<i>mec-7</i>	gk286003	R162Q		Recessive, Neomorph	Zheng et al., 2017
<i>mec-7</i>	e1343 , u127 , u957	P171L	CB1477 ; CB2217	Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u911	P171S		Dominant, Antimorph	Zheng et al., 2017
<i>mec-7</i>	u1056	S172P		Recessive, Loss of function	Zheng et al., 2017
<i>mec-7</i>	u48	S176F		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u449	V179A		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u10	S188F		Recessive, Loss of function	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u225	T214P		Recessive, Loss of function	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	ky852	P220S			Kirszenblat et.al. 2013
<i>mec-7</i>	u262	N226Y		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	gk286002	P234S		Recessive, Loss of function	Zheng et al., 2017
<i>mec-7</i>	u283	P243L		Dominant, Antimorph	Savage et al., 1994; Bountonas et al., 2011: 21368137; Zheng et al., 2017
<i>mec-7</i>	u129 , u958	G244S		Dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	n434	N247I	TU300	Dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017

Gene	Allele	Protein change	Strain	Mutation class	Reference
<i>mec-7</i>	u162	D249N		Dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u427	V255M		Semi-dominant	Savage et al., 1994
<i>mec-7</i>	e1505	G269D		Recessive, Loss of function	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u142	Q279stop		Recessive	O'Hagan et.al 2005; Zheng et al., 2017
<i>mec-7</i>	u178, u431	Q280stop		Recessive	Sze et al., 1997
<i>mec-7</i>	e1527	V286D	CB3276	Dominant	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u451	Q292P		Semi-dominant, Antimorph	Savage et al., 1994
<i>mec-7</i>	u445	M300V		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u98	M300T		Semi-dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u1058	A302T		Semi-dominant, Neomorph	Zheng et al., 2017
<i>mec-7</i>	u278	C303Y		Semi-dominant, Neomorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	gk286000	A314V		Semi-dominant, Antimorph	Zheng et al., 2017
<i>mec-7</i>	e1522	F317I		Recessive, Loss of function	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u234	R318Q		Recessive, Loss of function	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	u88	R318stop		Recessive	Sze et al., 1997
<i>mec-7</i>	u955	A352T		Dominant, Antimorph	Zheng et al., 2017
<i>mec-7</i>	u910, gk373602	P357L		Dominant, Antimorph	Zheng et al., 2017
<i>mec-7</i>	u956	P358L		Dominant, Antimorph	Zheng et al., 2017
<i>mec-7</i>	u143	G369R		Semi-dominant	Savage et al., 1994
<i>mec-7</i>	u428	G369E		Recessive, Loss of function	Gu et al., 1996; Zheng et al., 2017
<i>mec-7</i>	u1017	L377F		Recessive, Neomorph	Zheng et al., 2017
<i>mec-7</i>	u1059	R380S		Recessive, Neomorph	Zheng et al., 2017
<i>mec-7</i>	u18	A393T		Dominant, Antimorph	Savage et al., 1994; Zheng et al., 2017
<i>mec-7</i>	gk285997	A393V		Dominant, Antimorph	Zheng et al., 2017
<i>mec-7</i>	u173	G402R		Recessive	Savage et al., 1994
<i>mec-7</i>	u170	E407L		Recessive, Neomorph	Savage et al., 1994; Zheng et al., 2017

Gene	Allele	Protein change	Strain	Mutation class	Reference
<i>mec-7</i>	<i>u1060</i>	E410K		Semi-dominant, Antimorph	Zheng et al., 2017
<i>mec-7</i>	<i>u382</i>	E410:Tc5		Recessive	Savage et al., 1994
<i>mec-7</i>	<i>u388</i>	E410:Tc5		Recessive	Savage et al., 1994
<i>mec-7</i>	<i>u443</i>			Recessive	Savage et al., 1994
<i>mec-7</i>	<i>u448</i>			Recessive	Savage et al., 1994
<i>mec-7</i>	<i>u453</i>			Recessive	Savage et al., 1994
<i>mec-7</i>	<i>u80</i>			Recessive	Savage et al., 1994
<i>mec-7</i>	<i>u136</i>			Semi-dominant	Savage et al., 1994
<i>tbb-4</i>	<i>sa127</i>	L253F	SP1742	Gain of function	Hao et al., 2011
<i>ben-1</i>	<i>e1880</i>		CB3474	Semi-dominant	Driscoll et al., 1989
<i>ben-1</i>	<i>e1910</i>			Semi-dominant	Driscoll et al., 1989
<i>ben-1</i>	<i>e1911</i>			Semi-dominant	Driscoll et al., 1989
<i>ben-1</i>	<i>u102</i>			Semi-dominant	Driscoll et al., 1989
<i>ben-1</i>	<i>u116</i>			Semi-dominant	Driscoll et al., 1989
<i>ben-1</i>	<i>u134</i>			Semi-dominant	Driscoll et al., 1989
<i>ben-1</i>	<i>u462</i>			Semi-dominant	Driscoll et al., 1989
<i>ben-1</i>	<i>u463</i>			Semi-dominant	Driscoll et al., 1989
<i>tbb-6</i>	<i>oxTi207</i>		EG7952		
<i>tbq-1</i>	<i>t1465</i>	A401V	GE2959		Hannack et al., 2002

6.2. Table 2. Summary of the expression and function of *C. elegans* tubulins

Locus	Gene Model	Expression	Cellular, developmental, behavioral roles	Selected references
<i>tba-1</i>	F26E4.8	ALM, PLM, AVM, PVM, VA/VB/DA/DB motor neurons, multiple other neurons, <i>germline</i> , early embryo	cellular architecture, meiosis/mitosis, neuronal pathfinding, synaptogenesis	Lu and Mains, 2005; Phillips et al., 2004; Baran et al., 2010
<i>tba-2</i>	C47B2.3	pharynx, intestine, VB/DB motor neurons, PLM, ALA, other anterior neurons, <i>germline</i> , early embryo	cellular architecture, meiosis/mitosis	Lu and Mains, 2005; Phillips et al., 2004
<i>mec-12</i>	C44B11.3	ALM, PLM, AVM, PVM, other neurons	sensory neuron dendritic microtubules, mechanosensation	Chalfie and Sulston, 1981; Bounoutas et al., 2009; O'Hagan et al., 2005; Bounoutas et al., 2011
<i>tba-4</i>	F44F4.11	intestine and other tissues	unknown/redundant	D. D. Hurd, unpublished (expression)

Locus	Gene Model	Expression	Cellular, developmental, behavioral roles	Selected references
<i>tba-5</i>	F16D3.1	PHA, PHB, amphid neurons, perhaps other sensory neurons	sensory neuron axonemal microtubules, chemosensation and/or mechanosensation	Hao et al., 2011
<i>tba-6</i>	F32H2.9	IL2, CEM, HSN, RnB, HOB	sensory neuron axonemal/dendritic microtubules, chemosensation and/or mechanosensation	Hurd et al., 2010
<i>tba-7</i>	T28D6.2	ALM, PLM, AVM, PVM, intestine, and excretory pore cell (Figure 5)	microtubule dynamics	Figure 6 and Zheng et al., 2017
<i>tba-8</i>	ZK899.4	seam cells, neurons	unknown/redundant	D. D. Hurd and D. S. Portman, unpublished (expression)
<i>tba-9</i>	F40F4.5	CEP, OLQ, ASI, ADF, AFD, AWA, AWC, ASE, ADE, URX, RIG, PDE, PQR, RnA, HOA, various motor neurons, other neurons, and neuronal support cells	axonal/axonemal/dendritic microtubules, chemosensation and/or mechanosensation	Hurd et al., 2010
<i>tbb-1</i>	K01G5.7	germline, early embryo	cellular architecture, meiosis/mitosis	Lu et al., 2004; Wright and Hunter, 2003
<i>tbb-2</i>	C36E8.5	germline, early embryo, neurons, excretory cell	cellular architecture, meiosis/mitosis, neuronal pathfinding, synaptogenesis,	SLu et al., 2004; Wright and Hunter, 2003
<i>mec-7</i>	ZK154.3	ALM, PLM, AVM, PVM, FLP, PVD, BDU	sensory neuron dendritic microtubules, mechanosensation	Chalfie and Sulston, 1981; Bounoutas et al., 2009; O'Hagan et al., 2005; Bounoutas et al., 2011
<i>tbb-4</i>	B0272.1	CEP, OLQ, CEM, ASK, ADL, ASI, ASJ, ASH, AWB, AWA, AWC, ASE, ASG, FLP, HSN, PDE, PQR, PHA, PHB, RnA, RnB, HOA, vulval muscles	axonemal/dendritic microtubules, chemosensation and/or mechanosensation	Hurd et al., 2010; Hao et al., 2010
<i>ben-1</i>	C54C6.2	motor, sensory, and interneurons	axonal pathfinding, sensitivity to benzimidazole	Figure 5 and Driscoll et al., 1989

Locus	Gene Model	Expression	Cellular, developmental, behavioral roles	Selected references
<i>tbb-6</i>	T04H1.9	intestine and other tissues	unknown/redundant	D. D. Hurd, unpublished (expression)
<i>tbg-1</i>	F58A4.8	early embryo, sensory neurons, other tissues	microtubule organization/ nucleation, spindle organization, nuclei positioning, tissue integrity, embryonic elongation	Bobinnec et al., 2000; Strome et al., 2001; O'Toole et al., 2012; Quintin et al., 2016; Wang et al. 2015

6.3. Table 3. Summary of the expression and function of regulators, modifiers, and proteins associated with *C. elegans* tubulin and microtubules

Locus	Sequence	Expression	Molecular, cellular, developmental, and/or behavioral roles	Selected references
<i>pfid-1, pfid-2, pfid-3, pfid-4, pfid-5, pfid-6</i>	C08F8.1, H20J04.5, T06G6.9, B0035.4, R151.9, F21C3.5	ubiquitous	protein folding, α -tubulin levels, microtubule dynamics, cell division, cell migration	Lundin et al., 2008;
<i>cct-1, cct-2, cct-3, cct-4, cct-5, cct-6, cct-7, cct-8</i>	T05C12.7, T21B10.7, F54A3.3, K01C8.10, C07G2.3, F01F1.8, T10B5.5, Y55F3AR.3	ubiquitous	protein folding, α -tubulin levels, cell division, cell migration, actin localization, formation/maintenance of microvilli in intestine	Saegusa et al., 2014
<i>mec-17, atat-2</i>	F57H12.7, W06B11.1	both: ALM, AVM, PLM, and PVM <i>atat-2</i> only: OLQ, CEP, PVR	acetylation of α -tubulin, microtubule architecture and stability, mechanosensation	Cueva et al., 2012; Topalidou et al., 2012, Davenport et al., 2014
<i>ttll-4, ttll-5, ttll-9, ttll-11, ttll-12, ttll-15</i>	ZK1128.6, C55A6.2, F25C8.5, H23L24.3, D2013.9, K07C5.7	amphid and other sensory neurons, body wall muscles, motor neurons	glutamylolation of tubulins	Kimura et al., 2010; O'Hagan et al., 2011
<i>ccpp-1, ccpp-6</i>	F56H1.5, EEED8.6	ASE, ASG, ASH, ASI, ASK, ASL, ADF, ADL, AWA, AWB, AWC, AFD, IL2, CEM, RnB1-RnB9, HOB	deglutamylolation of tubulins, microtubule architecture and stability, motor protein activity, cilia stability	Kimura et al., 2010; O'Hagan et al., 2011
<i>ptl-1</i>	F42G9.9	ALM, AVM, PLM, PVM (early larva only), other neurons	motor protein activity, egg hatching, mechanosensation	Goedert et al., 1996; Gordon et al., 2008; Tien et al., 2010

Locus	Sequence	Expression	Molecular, cellular, developmental, and/or behavioral roles	Selected references
<i>elp-1</i>	F38A6.2	ALM, AVM, PLM, PVM, IL1, male tail ray neurons, body wall muscle, vulval muscles, spermathecal valve cell, intestine, and seam cells	associates with microtubules; mechanosensation, motility and egg-laying (along with <i>DYS-1</i>)	Hueston et al., 2008; Hueston and Suprenant, 2009
<i>tac-1</i>	Y54E2A.3	centrosomes of blastomeres	associates with <i>ZYG-8</i> and <i>ZYG-9</i> , polymerizes/stabilizes microtubules, pronuclear migration, spindle positioning	Bellanger and Gönczy, 2003; Srayko et al., 2003
<i>zyg-8</i>	Y79H2A.11	centrosomes of blastomeres, ALM, AVM, PLM, PVM, VD and DD motor neurons	associates with <i>TAC-1</i> and <i>ZYG-9</i> , polymerizes/stabilizes microtubules, pronuclear migration, spindle positioning, cell polarity, locomotion, mechanosensation	Gönczy et al., 2001; ; Bellanger et al., 2012
<i>zyg-9</i>	F22B5.7	centrosomes of blastomeres	associates with <i>TAC-1</i> and <i>ZYG-8</i> , polymerizes/stabilizes microtubules, pronuclear migration, spindle positioning	Matthews et al., 1998; Bellanger et al., 2007
<i>cls-1, cls-2, cls-3</i>	C07H6.3, R107.6, ZC84.3	centrosomes, kinetochores, and spindle of blastomeres	stabilize microtubules, spindle length and positioning	Espiritu et al, 2012
<i>che-12</i>	B0024.8	ASE, ASG, ASH, ASI, ASJ, ASK, PHA, PHB	stabilizes axonal microtubules, cilia structure, chemosensation	Bacai et al., 2008; Das et al., 2015
<i>efa-6</i>	Y55D9A.1	germline, blastomeres, neurons	associates with <i>TAC-1</i> and <i>ZYG-8</i> , inhibits axon regrowth after injury	O'Rourke et al., 2010; Chen et al., 2011; Chen et al., 2015
<i>rmd-1, rmd-2, rmd-3, rmd-4, rmd-5, rmd-6</i>	T05G5.7, C27H6.4, B0491.3, F36H12.11, T23B3.3, R13H9.1	germline, blastomeres	associates with microtubules, associates with <i>AIR-2</i> (aurora B kinase), kinetochore-microtubule attachment	Oishi et al., 2007
<i>mei-1, mei-2</i>	T01G9.5, F57B10.12	germline, blastomeres	severs and/or bundles microtubules, meiotic spindle formation and maintenance	McNally and McNally, 2011; Connolly et al., 2014; McNally et al., 2014

Locus	Sequence	Expression	Molecular, cellular, developmental, and/or behavioral roles	Selected references
<i>spas-1</i>	C24B5.2	ubiquitous	severs microtubules, reproduction	Matsushita-Ishiodori et al., 2009
<i>klp-7</i>	K11D9.1	kinetochores, blastomeres, neurons	depolymerizes microtubules, meiotic spindle assembly	Srayko et al., 2005; Ghosh-Roy et al. 2012; Connolly et al., 2015
<i>ebp-1, ebp-2, ebp-3</i>	Y59A8B.7, VW02B12L.3, Y59A8B.9	unknown	interact with plus ends of microtubules, axon regrowth after injury	Motegi et al., 2006; Chen et al., 2011
<i>ptrn-1</i>	F35B3.5	neurons	polymerizes microtubules, axon remodeling and regrowth after injury	Marcette et al. 2014; ; Richardson et al., 2014; ; Chuang et al. 2014:
<i>noca-1</i>	T09E8.1	hypodermis, neurons, intestine, pharynx	polymerizes (non centrosomal) microtubules, tissue integrity (with γ -tubulin and PTRN-1), nuclear positioning	Wang et al. 2015; Quintin et al., 2016

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