

# Exploring the potential of soapstock over a glycerol in vitamin K2 production by *Bacillus subtilis* natto: a comparative analysis

Faranak Ansari<sup>1</sup>, Hoda Nouri<sup>2</sup> and Hamid Moghimi<sup>1\*</sup>

# **Abstract**

**Background** Vitamin K2 is an essential nutrient for blood coagulation and cardiovascular health and mainly produced by bacteria strain like *B. subtilis*. researchers have explored producing strain improvement, cultivation mode, environmental optimization, increased secretion, and using cheaper carbon and nitrogen sources in order to increase vitamin K2 productivity. This study examines the impact of varioius concentration of soapstock, which is a by-product of vegetable oil refning, as an alternative carbon source with lower pirce, in the fermentation medium instead of glycerol on the microbial synthesis of vitamin K2 using *B. subtilis* natto ATCC 23857.

**Results** The results demonstrate that when the glycerol in fermentation medium was substituted with soapstock, by 75% concentartion, the fermentation process produced a yield of 158.16 mg/L of vitamin K2 after 72 h; This was 3.8 times more than the control medium containing glycerol. When the entire culture medium was replaced with wastewater, the vitamin K2 concentration reached 21.18 mg/L, 52% of the control medium's concentration. If the carbon sources in the fermentation medium consisted of 20% soapstock and 47.4 g/L glycerol (maintaining the same fnal glycerol concentration as the control medium), the vitamin K2 concentration reached 35.7 mg/L or 85.8% of the control medium. The analysis of soapstock fermentation medium characteristics reveals that after fermentation with *B. subtilis*, the COD of soapstock fermentation medium was dramatically reduced from 259,500 mg/L to 57,830 mg/L.

**Conclusions** Using soapstock as an alternative carbon source for fermentation did not negatively impact the bioprocess and increased vitamin K2 production. Therefore, this research introduces an alternative carbon resource for vitamin K2 production and paves the way for the biorefnement of soapstock.

**Keywords** *Bacillus subtilis* natto, Carbon source, Crude glycerol, Soapsotck, Vitamin K2

\*Correspondence:

Hamid Moghimi

hmoghimi@ut.ac.ir

<sup>1</sup> Department of Microbiology, School of Biology, College of Science, University of Tehran, Tehran 1417864411, Iran

<sup>2</sup> Department of Systems Biotechnology, Institute of Industrial and Environmental Biotechnology, National Institute for Genetic Engineering and Biotechnology (NIGEB), Tehran, Iran

# **Introduction**

Vitamin K2 is a member of the fat-soluble Vitamin K group, which is essential for blood coagulation, osteoporosis prevention, and cardiovascular health [[1–](#page-9-0)[3](#page-9-1)]. Also, known as menaquinone  $(MK)$  [[4](#page-9-2)], and comes in several forms, ranging from MK-2 to MK-14 [[5\]](#page-9-3), among all these, MK-7 is particularly noteworthy for its high bioavailability and extended half-life making it more efective than other vitamin K2 homologs [[6,](#page-9-4) [7](#page-9-5)]. Various microorganisms produce menaquinone, an important electron carrier in the respiratory chain of microbial cells [[8,](#page-9-6) [9\]](#page-9-7). Bacteria are the predominant producers [\[10](#page-9-8)]



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modifed the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by-nc-nd/4.0/>.

of menaquinone, and diferent strains can produce it in various forms. The production of vitamin  $K2$  in the MK-7 form by *B. subtilis* can reach up to 96% of the total production [\[11,](#page-9-9) [12\]](#page-9-10). *B. subtilis* is a suitable candidate strain for the microbial production of vitamin K2 owing to its consideration as (Generally Recognized As Safe) organism. Also, *B. subtilis* is notable for its high production of MK-7, compared to other menaquinone producers [[13](#page-9-11), [14](#page-9-12)]. Many experiments have been conducted to increase vitamin K2 productivity and accelerate its industrial production  $[14–16]$  $[14–16]$ . These include strain improvement [[17,](#page-9-14) [18\]](#page-9-15), diferent environmental optimization such as nitrogen sources, carbon sources, temperature [[15\]](#page-9-16), pH [[19\]](#page-9-17), bioflm formataion [[20\]](#page-9-18), dissolved oxygen [[21\]](#page-9-19), exploring diferent cultivation strategies and innovative bioreactor such as bioflm reactor  $[22]$  $[22]$  $[22]$ , increased secretion  $[23]$ , and using more afordable options for carbon and nitrogen sources [[24](#page-9-22), [25](#page-9-23)]. Carbon and nitrogen sources are infuential factors in vitamin K2 production. Result from previous study on determining potential carbon sources in mk-7 production reveals that among glycerol, glucose, sucrose and starch, glycerol is the most signifcant factor on MK-7 synthesis among all factors studied [[15\]](#page-9-16). MK-7 production often has low yields and expensive production costs; some products could take the place of pricey raw ingredients in it [\[26](#page-9-24)]. In order to reduce production costs, using a cheaper carbon source such as crude glycerol is considerable [[27](#page-9-25)]. Crude glycerol is a by-product of biodiesel production through transesterifcation, soap manufacturing via saponifcation, and hydrolysis reactions [\[28\]](#page-9-26). Another source of crude glycerol is soapstock. Soapstock is a by-product of refning crude or degummed vegetable oils. During this process, oils are treated with alkali to produce sodium soap. Enzyme production, biodegradation, and biodiesel production are some of the major uses of soapstock as a valuable biotechnological resource  $[29, 30]$  $[29, 30]$  $[29, 30]$  $[29, 30]$ . The use of alternative, economical carbon sources has attracted the attention of researchers in recent study, to utilize these alternative carbon sources for MK-7 production, a mutant strain of *Bacillus amyloliquefaciens* capable of digesting maize meal was developed, resulting in a high MK-7 content [[31](#page-9-29)]. Wang et al. used liquefied soybean flour and corn flour as primary carbon sources, achieving an MK-7 production yield of 18.9 mg/mL in shaking flask culture  $[32]$  $[32]$ . This study used soapstock as an alternative carbon source instead of glycerol for the frst time to develop an economical fermentation medium for vitamin K2 in MK-7 form. This approach not only reduces production costs but also advances the biorefnery process and promotes a circular economy.

# **Material and methods**

# **Bacterial strain and culture conditions**

*B. subtilis* natto ATCC 23857 was used in this study as a producing microorganism and was stored on nutrient agar plates and fresh plates cultivated monthly. One loop of bacteria from the fresh nutrient agar plate was inoculated to pre-culture media containing beef extract 5 g/L, glucose 30 g/L, peptone 40 g/L, NaCl 5 g/L, and yeast extract 5 g/L. For soapstock's pre-culture medium, soapstock was diluted with distilled water in a 1:9 (v/v) ratio. The fermentation process was carried out under shaking conditions at 120 rpm and 37 °C for 24 h in a shaking incubator.

The control fermentation medium for vitamin K2 production was based on a basal medium. This medium consists of 50 g/L of glycerol, 30 g/L of soybean peptone, 0.6 g/L of yeast extract, 0.3 g/L of  $K_2HPO_4$ , 0.3 g/L of  $MgSO<sub>4</sub>$ .7H<sub>2</sub>O, and 0.1 g/L of CaCl.H<sub>2</sub>O, used as a control medium according to Zhang's study  $[27]$ . The soapstock fermentation media contained 1.3 to 13 g/L of soapstock, 30 g/L of soybean peptone, 0.6 g/L of yeast extract, 0.3 g/L of  $K_2HPO_4$ , 0.3 g/L of MgSO<sub>4</sub>.7H<sub>2</sub>O, and 0.1 g/L of  $CaCl.H<sub>2</sub>O$ . The control and soapstock fermentation media were seeded at a rate of 5% with a 24-h pre-culture of soapstock or control media. These flasks were then placed in a rotary shaker under the conditions of 210 rpm and 37°C because vitamin K2 production was boosted under higher agitation rate conditions [[21](#page-9-19)], and this continued until the death phase was detected. Samples were taken every 12 h to measure biomass, pH, absorbed glycerol, and vitamin K2 yield. All media's pH were adjusted at 7 before cultivation.

# **Soapstock characteristics**

Soapstock was provided by Varamin oil-extracting factory No.1 Iran. The glycerol content of soapstock was measured at 13 g/L measured by spectrophotometric methods, pH 10.38 was measured by pH meter, COD 25050 mg/L was measured with the COD kits, TDS 2400 mg/L was measured by the TDS meter, and fatty acid measured to be 16% (w/v) by liquid–liquid extraction.

# **Soapstock preparation**

In the frst attempt to produce vitamin K2 using soapstock, glycerol was partially replaced in the control medium by adding soapstock at the following concentrations: 10%, 20%, and 30%. In all batches the substitution of glycerol with an equivalent of its concentration in soapstock, allowed the fnal glycerol content to remain at 50 g/L. More details are provided in Table [1](#page-2-0) based on the varying results from diferent glycerol concentrations, this research conducted to fnd out how difrent

Name	Glycerol provided from soap stock concentration (g/L)	Control concentration (g/L)	Total glycerol concentration (g/L)
Batch $10\% + Gly$		48.7	50
Batch $20% + Gly$	2.6	47.4	50
Batch $30\% + G$ ly	3.9	46.1	50

<span id="page-2-0"></span>**Table 1** Glycerol and soapstock mixing ratio

soapstock concentrations afected the formation of vitamin K2. Therefore, in the next step, glycerol was eliminated, and soapstock was used as the sole carbon source, diluted with distilled water in the following ratios: 10:90, 20:80, 30:70, 50:50, 75:25, and ultimately, 100% soapstock was used in the fermentation medium.

# **Measurement of bacterial growth and residual glycerol concentration**

The spectrophotometric approach was used to detect free glycerol based on the oxidation of glycerol and the production of formaldehyde after it reacted with acetylacetone, resulting in a yellow compound measured at 410 nm  $[33]$  $[33]$  using a Shimadzu UV-1601 device. The dry cell weight was determined by precipitating cells from a 10 mL fermentation broth sample and drying them overnight at 105 °C [\[34](#page-9-32)].

# **Determination of MK‑7**

After bacterial growth, 10 mL fermentation broth was provided to sterile centrifuge tubes, then biomass and supernatant were separated using a centrifuge set at 6000 Rpm for 10 min. After that, supernatant was transferred to a sanitized fask in preparation for the vitamin K2 extraction process.

For extracting the vitamin K2 from the aqua phase of the medium the mixture of n-hexane and isopropanol was added to the supernatant  $(4:2:3, v/v)$  and the organic phase shaped. Phases were separated by centrifuging the extraction mixture for 10 min at 6000 Rpm after it had been incubated for an hour at 30°C in a shaker incubator. In the next step 5mL of solvent layer extracted from the fask and transferd to a rotatory evaporator for 10 min at 40 °C. Then evaporator flask washed with 1mL of HPLCgrade ethanol to obtain MK-7 extract [[35\]](#page-9-33).

# **HPLC analysis**

Vitamin K2 was quantifed using an Agilent 1260 infnity high-performance liquid chromatography system. The following were the HPLC's operating conditions for the chromatographic column: SB-C18 4.6\*250, SB-C18 guard cartridge 4.6\*12.5/pk; mobile phase: 97% methanol: 3% water; flow rate: 1.0 mL/min; column temperature:  $25^{\circ}$ C; and detection wavelength:  $268$  nm  $[36]$  $[36]$ . The MK-7 calibration curve was linear between 5 mg/L and 100 mg/L ( $R^2$ =0.997).

#### **Soapstock parameters determination**

Soapstock parameters, including chloride, nitrite, phosphate, ammonium, sulfte, and total hardness, were measured before and after the fermentation with Vaheb analytical kits. Also, the Total Dissolved Solid (TDS) was measured by the TDS meter, and the Total Suspended Solids (TSS) was determined After fltering and drying at 105°C following the standard method 2540 B [\[37](#page-9-35)]. The chemical oxygen demand (COD) was measured with the COD kit, which is close to the standard method 5220 B-open reflux titrimetric  $[38]$  $[38]$ , and using the Hanna HI83314 COD photometer for reading. The determination of total fatty acid was performed by the liquid–liquid extraction with n-hexane, as recommended by EPA Method 1664 [[39\]](#page-9-37).

# **Statistical analysis**

The statistical software used for testing the result was SPSS Version 27.0.1, and the analysis was conducted in triplicate, also in this study, one-way ANOVA and Tukey's HSD tests were employed and set with a 95% confdence level, and the signifcance level was reported as *p*≥0.05.

# **Result and discussion**

# **Determination of kinetics of biomass, glycerol, and vitamin K2 concentration**

To examine the efect of soapstock concentration on vitamin K2 production, the kinetics of biomass, glycerol consumption, and vitamin K2 concentration were studied in both a control fermentation medium and a soapstock fermentation medium (Fig. [1](#page-3-0)).

According to the kinetics shown in Fig. [1a](#page-3-0), *B. subtilis,* was in its exponential growth phase from 12 h until 36 h in the control medium. while, in the soapstock medium, it took 48 h to observe this phase, and most of glycerol consumption occurred during this phase in the both medium. Biosynthesis of vitamin K2 began after 24 h of fermentation in the control medium. however, during the frst 24 h of fermentation in the soapstock medium, no vitamin K2 was detected, and it required



<span id="page-3-0"></span>**Fig. 1** Comparison of kinetics of biomass, glycerol consumption, and vitamin K2 production in control medium and soapstock fermentation medium. **a** Dry weight, **b** Residual glycerol concentration, and **c** Vitamin K2 concentration

48 h of fermentation to reach its maximum concentration. The biomass reached its highest concentration of 20.8 g/L at the 120 th hour with soapstock medium and compared to the control medium it was 3.7 fold higher. It indicates that *B. subtilis* was provided with a secondary carbon source from the soapstock medium, which is a fatty acid. This resulted in increased biomass growth even after most of the glycerol in the medium had been consumed. The ability of *B. subtilis* strain to degrade hydrocarbons [\[40](#page-9-38)] and bioremediate oily wastewater has been reported [\[41\]](#page-10-0). Additionally, the lipolytic activity of *B. subtilis* improved by using inexpensive vegetable oils [[42\]](#page-10-1). Compared to stationiry phase, it has been reported, MK-7 production mainly observed in growth phase [\[15](#page-9-16)], while the highest concentration of vitamin K2 in the control medium reached at  $41$  mg/L in  $72$  h. This results were in agreement with litature report indicating that MK-7 production continues in spite of glycerol depletion [[43\]](#page-10-2). However, Zhang's results showed no signifcant differences in bacterial growth or MK-7 synthesis between pure and crude glycerol-containing media, suggesting that crude glycerol can be replaced with pure glycerol as a lower-cost carbon source [\[27](#page-9-25)]. Whereas in the soapstock medium, the MK-7 concentration reached 21.8 mg/L in the 48 th hour of fermentation and decreased after the logarithmic phase. This drop in production might be due to the diferences in the components of these two media, as glycerol's efficiency in transferring through the cell

membrane contributes to some advantages in the MK-7 production process [\[44\]](#page-10-3).

#### **Soapstock as a partial replacement for glycerol**

Based on the result obtained from the comparison between the control medium containing glycerol and soapstock medium, both partial replacement of glycerol with soapstock and dilution of soapstock were examined. In the frst step, a partial replacement for glycerol was such that, the total glycerol concentration in the fermentation medium matched that of the control medium, and it contained both glycerol and soapstock in a specifc ratio that maintaining a total glycerol concentration of 50  $g/L$  (Table [1](#page-2-0)); The kinetics of glycerol consumption and biomass production were assessed to determine the optimal batch for vitamin K2 analysis, as shown in Figs. [2](#page-4-0) and [3](#page-5-0).

In batch 10%+Gly, exponential growth was observed at 36 h, similar to the control fermentation medium, and glycerol consumption increased rapidly until the 48th hour. Batch 30%+Gly the death phase occurred in the 96th hour of fermentation, with the highest biomass of 7.07 g/L observed at 60 h (Fig. [2](#page-4-0)a,b). Batch  $20\% + Gly$ showed a more balanced glycerol consumption and the



<span id="page-4-0"></span>**Fig. 2** Comparison of the kinetics of biomass, glycerol consumption, and vitamin K2 production in batches 10%+Gly, 20%+Gly, and 30%+Gly. **a** Dry weight. **b** Glycerol consumption. **c** Kinetics of biomass, glycerol concentration and vitamin K2 concentration in batch 20%+Gly



<span id="page-5-0"></span>**Fig. 3** Kinetics of biomass, glycerol consumption, and vitamin K2 production in diferent soapstock concentrations. **a** Kinetics of biomass, glycerol consumption with 10% (v/v) soapstock, **b** Kinetics of biomass, glycerol consumption and vitamin K2 concentration with (v/v) 20% soapstock, **c** Kinetics of biomass, glycerol consumption with (v/v) 30% soapstock, **d** Kinetics of biomass, glycerol consumption and vitamin K2 concentration with (v/v) 50% soapstock, **e** Kinetics of biomass, glycerol consumption and vitamin K2 concentration with (v/v) 75% soapstock, **f** Kinetics of biomass, glycerol consumption and vitamin K2 concentration with (v/v) 100% soapstock

highest biomass concentration, compared to the other batches. Consequently, it was the preferred choice for measuring vitamin K2 production due to the result from literature suggesting addition of excessive glycerol has no efect on MK-7 production, as microorganisms cannot utilize it [\[45](#page-10-4)]. According to reports, using glycerol and another carbon source at the same time can cause glycerol to act as an activator during the production of MK-7. [\[16\]](#page-9-13). In batch  $20\% + Gly$ , the maximum production of vitamin K2 was 18.57 mg/l in the 96th hours of fermentation (Fig. [2](#page-4-0)c). Glycerol may act as an enhancer in MK-7 production without impacting cell growth [\[46](#page-10-5)]; This result indicates that soapstock addition improved cell growth while not signifcantly afecting MK-7 production compared to the control medium.

# **Efect of diferent soapstock concentrations on vitamin K2 production**

Considering that soapstock addition as a carbon source infuenced both biomass and vitamin K2 production compared to the control medium, further experiments were conducted to assess the impact of varying soapstock concentrations without added glycerol. The kinetics of biomass, vitamin K2 content, and glycerol consumption are shown in Fig. [3](#page-5-0).

In batch 20% soapstock, with 2.6 g/l glycerol, the highest production of vitamin K2 was measured at the 144th hour from the initiation of fermentation, with a concentration of 9.44 mg/l, also fermentation time was extended from 96 to 144 h due to an increase in soapstock concentration, which provided more access to carbon sources and delayed the death phase of *B. subtilis* natto (Fig. [3b](#page-5-0)). Batch 30% was not analyzed for vitamin K2 production due to its similarity to the fermentation process of the previous culture (Fig.  $3c$ ). In batch 50%, similar to the control fermentation medium, the highest production of vitamin K2 of 35.7 mg/L was achieved at 72 h, the measurement of vitamin K2 concentration continued until 120th hour, at the point which it decreased to 21.2 mg/L (Fig. [3](#page-5-0)d). Batch 75% soapstock, with 9.75 g/L of glycerol, achieved the highest vitamin K2 concentration of  $158.16$  mg/L. This is maximum vitamin K2 yeild among all batches (Fig.  $3e$ ). The addition of glycerol enhances MK-7 biosynthesis by optimizing viscosity and water activity, which improves mass transfer [[47](#page-10-6)] consequently, the increase in MK-7 observed in this batch may be attributed to these effects. Batch 100%, with 13  $g/L$ of glycerol, reached a peak vitamin K2 concentration of 18.21 mg/L (Fig. [3](#page-5-0)f). After 36 h of fermentation, *B. subtilis* natto exited the exponential growth phase and entered the stationary phase. Compared to the control, vitamin K2 production was enhanced in the soapstock fermentation medium, especially with a 75% soapstock concentration. In a similar study, crude glycerol was used as a carbon source to produce vitamin K2 by *B. subtilis* Z-15 economically and confrmed that the usage of crude glycerol didn't afect either the synthesis of vitamin K2 or the growth of *B. subtilis* and using crude glycerol instead of control resulted in a 70% cost saving, too  $[27]$  $[27]$ . The impact of using wheat starch wastewater instead of glycerol as an extra carbon source has been investigated. This substitution successfully reduced the cost of the carbon source by 33%, the price of the nitrogen source by 7%, and the cost of the phosphate source by 100%  $[48]$  $[48]$ . The vitamin K2 production yield in diferent alternative carbon sources is provided in Table [2](#page-6-0). All these studies successfully took advantage of utilizing alternative carbon sources strategy as a cost-efective substrate for economical MK-7 production. This research has the highest concentration of vitamin K2 compared to other studies using alternative carbon sources, being 3.5 times higher than Zhang's research  $[27]$  $[27]$  $[27]$  and 2.5 times higher than Xu's  $[31]$  $[31]$ . The maximum value for vitamin K2 production was reported to be 226 mg/L, using glycerol as the carbon source [\[21](#page-9-19)], which is 1.4 times higher than the rate in this study.

Vitamin K2 purifcation is complex and costly, and it becomes even more complex when soapstock is employed in its production bcause of intensifying the purification challenges. There are potential solutions to this issue, including the utilization of complete fermented medium as a product. This strategy successfully implemented by Novin et al. by introducing a fermented dairy product containing vitamin K2, and efectively eliminating the downstream process for vitamin K2 purifcation  $[31]$  $[31]$ . This fermented product can be used as an animal supplement with some pre-treatment for soapstock.

<span id="page-6-0"></span>



<span id="page-7-0"></span>

vitamin K2 contributes to animal health similarly to humans, with defciencies causing coagulation and skeletal issues [\[43](#page-10-2)].

A quantitative comparison of vitamin K2 production has been obtained from this research outcome through the employment of Monod kinetic analysis. This data provides a systematic comparison of the results due to ability of the Monod equation for relating vitamin K2 production to biomass growth and glycerol consumption. Calculations for productivity, cell yield coefficient, vitamin K2 yield coefficient, specific growth rate, specific rate of glycerol consumption, and specifc rate of vitamin K2 generation are presented in Table [3](#page-7-0). Batch 75% showed the highest specifc rate of vitamin K2 formation, vitamin K2 yield coefficient, and productivity. The second vitamin K2 productivity recorded in the control medium, while batch  $20\% + Gly$  had the highest cell yield coefficient, indicating that the combination of glycerol and soapstock promoted greater biomass growth. Studies suggest that some carbon sources, like sucrose, are better for biomass conversion rather than increasing MK-7 production, as observed in 100% soapstock media  $[38]$  $[38]$ . The highest specifc rate of glycerol consumption was found in the control medium. Catabolite repression in *Bacillus* reduces gene expression for other carbon sources when a rapidly metabolizable one, like glycerol, is present  $[49, 50]$  $[49, 50]$  $[49, 50]$  $[49, 50]$  $[49, 50]$ . The maximum value of μm was recorded in the pure fermentation medium and also in the 75% soapstock; however, the specifc rate of vitamin K2 production and productivity was greater in the 75% soapstock batch compared to the control fermentation medium. From these fndings, the participation of another parameter in vitamin K2 production, except μm, could be concluded. In addition, comparing the specifc rate of glycerol consumption results revealed that the control fermentation medium had the greatest value, while the rates for the 20%, 50%, 75%, and 100% batches were the same. This emphasizes that the parameter leading to an increase in vitamin K2 is not related to glycerol and is possibly related to improved in situ recovery or permeability of cell membrane balance. The presence of certain fatty acids in the soapstock seems to create conditions that allow it to function as a surfactant, regulating the permeability of the bacterial membrane so that it facilitates the secretion of more extracellular vitamin K2 and might improve the recovery of vitamin K2 during the extraction process [\[21,](#page-9-19) [45](#page-10-4), 51. The productivity of batches of 50% and 100% was similar and closer to that of batch 75%, suggesting these soapstock concentrations are closer to the optimum for membrane permeability and vitamin K2 recovery. The productivity of both Gly+20% and 20% batches was similar, as was the specifc rate of vitamin K2 formation, indicating that glycerol concentration was not the primary factor afecting vitamin K2 production; otherwise, the vitamin K2 production in the  $Gly + 20%$  batch would have exceeded that of the 20% batch, indicating the efectiveness of the surfactant in vitamin K2 recovery. The vitamin K2 yield coefficient also showed a resemblance between these two batches, although glycerol had been suggested for increasing the yield of vitamin K2 production per cell [\[46](#page-10-5)]. When the soapstock concentration reached 50%, the vitamin K2 yield coefficient increased, implying that it could produce more vitamin K2 for the consumption of the same substrate unit. With a 75% soapstock concentration, the production of K2 reached its highest value. However, the biomass efficiency in converting the substrate to vitamin K2 was likely lowest at 100% soapstock concentration, it might be the efect of

<span id="page-7-1"></span>**Table 4** Comparison of vitamin K2 yield in soapstock concentration

Parameter	Control	Batch $20% + Gly$	Batch 20%	Batch 50%	Batch 75%	<b>Batch 100%</b>
Vitamin K2 yield (mg/L)	41.58	18.57	9.35	35.7	158.16	21.18
Initial glycerol concentration (g/L)	50	50	2.6	6.5	9.75	
Biomass (g/L)	5.25	6.51	6.33	13.8	14.4	15.2
Fermentation time $(h^{-1})$		96	144	72	72	48

Row	Parameter	Soapstock concentration (mg/L)	After 72h fermentation concentration Difference (%) (mq/L)	
	COD	259,500	57,830	77.7
$\overline{2}$	<b>TSS</b>	44,000	34,600	21.3
3	<b>TDS</b>	2920	1240	57.5
$\overline{4}$	Nitrite	0.1	0.05	50
5	Phosphate	400	300	25
7	Total hardness	256/5	171	33.2
8	pH	10.38	7.7	25.1
9	Sulfites	3200	2600	18.7
10	Hg	$\overline{\phantom{0}}$		$\Omega$

<span id="page-8-0"></span>**Table 5** Soapstock medium parameters before and after fermentation by *B. subtilis* natto

improper cell membrane permeability balance, while a slight increase in surface tension positively afected the cell membrane's state and composition, enhancing MK-7 concentration [\[43](#page-10-2)].

Table [4](#page-7-1) summarizes the vitamin K2 yield comparisons in diferent soapstock-containing media, along with the fermentation times spent for its production and the biomass values at those points. Result from this table indicates that the initial glycerol concentration does not have any signifcant efect on vitamin vitamin K2 production.

Statistical analysis of the results showed that addition of soapstock signifcantly afected vitamin K2 production. The groups showing the most similar results were media containing  $100\%$  soapstock,  $20\%$  soapstock + glycerol, and 20% soapstock; whereas the greatest difrence in K2 production was found in the batch with 75% soapstock. In biomass production, the media with 20% soapstock+glycerol, 20% soapstock, and 30% soapstock were the most compareable batches and the control group was closer to the 10% soapstock medium, however the medium with 100% soapstock exhibited the greatest diference.

# **Efect of fermentation with** *B. subtilis* **natto on soapstock medium parameters**

The soapstock medium was characterized by measuring the concentration of some parameters, as follows: chloride, nitrite, phosphate, ammonium, sulfte, total hardness, Hg, COD, TDS, and TSS, both before inoculation and after 72 h of fermentation (Table [5](#page-8-0)).

The COD of soapstock reduced from  $259,500$  mg/L to 57,830 mg/L indicating a 77% reduction. TDS diminished from 2920 mg/L to 1240 mg/L (57% reduction), and TSS dropped from 44,000 mg/L to 34,600 mg/L (21% reduction). Mercury was not detected in the culture medium with 100% soapstock before or after fermentation. In ordr to examine the efect of fermentation process on pH, this medium was not adjusted on 7 before the cultivatoin. The initial pH of soapstock was 10.38 (alkaline), which dropped to 7.7 (neutral) after 72 h of fermentation. Developing cleaner vitamin K2 production has been investigated lately, and it has been reported that there is great potential to reduce the pollution of wheat starch wastewater  $[48]$ . The result from this study could successfully imply MK-7 production and soapstock biorefning.

# **Conclusion**

The results of study indicate that using soapstock as an alternative carbon source, instead of pure glycerol, not only did not negatively impact the fermentation process but also increased vitamin K2 production by 3.8 times with 75% soapstock. The increase is attributed to fatty acids in soapstock, which serve as a supplementary carbon source, promoting biomass growth and thereby increasing vitamin K2 production. Additionally, these fatty acids function as surfactants, regulating bacterial membrane permeability and enhancing extracellular vitamin K2 production. Also, during the fermentation process, the COD of the soapstock was reduced by 77%; consequently, this innovative use of soapstock would take the edge of the environmental pollution of soapstock industrial plants and introduce soapstock as a potential carbon source for the vitamin K2 production and in-situ recovery of it.

# **Abbreviations**

MK Menaquinone HPLC High-performance liquid chromatography Total dissolved solid TSS Total suspended solids COD Chemical oxygen demand ANOVA Analysis of variance

#### **Acknowledgements**

Not applicable.

#### **Author contributions**

F. A. carried out the experiment and wrote the original draft of the manuscript. H.N. and H. M. edit the manuscript. The work was supervised and designed by H. N. and H.M.; and all authors read and approved the fnal version of the manuscript.

#### **Funding**

This research did not receive any specifc grant from funding agencies in the public, commercial, or not-for-proft sectors.

#### **Availability of data and materials**

No datasets were generated or analysed during the current study.

#### **Declarations**

#### **Ethics approval and consent to participate**

Ethics approval is not applicable as this article does not describe any studies involving human participants or animals.

#### **Consent for publication**

Not applicable.

## **Competing interests**

The authors declare no competing interests.

Received: 26 September 2024 Accepted: 20 December 2024

#### **References**

- <span id="page-9-0"></span>1. Schwalfenberg GK. Vitamins K1 and K2: the emerging group of vitamins required for human health. J Nutr Metab. 2017;2017:6254836.
- 2. Frandsen NE, Gordeladze JO. Vitamin K2 and bone health. In: Vitamin K2-vital for health and wellbeing. New York: IntechOpen; 2017. p. 101–23.
- <span id="page-9-1"></span>3. Hariri E, Kassis N, Iskandar JP, Schurgers LJ, Saad A, Abdelfattah O, Bansal A, Isogai T, Harb SC, Kapadia S. Vitamin K(2)—a neglected player in cardiovascular health: a narrative review. Open Heart. 2021;8: e001715.
- <span id="page-9-2"></span>4. Meganathan R. Biosynthesis of menaquinone (vitamin K2) and ubiquinone (coenzyme Q): a perspective on enzymatic mechanisms. In: Vitamins & hormones, vol. 61. New York: Academic Press; 2001. p. 173–218.
- <span id="page-9-3"></span>5. Capozzi A, Scambia G, Migliaccio S, Lello S. Role of vitamin K2 in bone metabolism: a point of view and a short reappraisal of the literature. Gynecol Endocrinol. 2020;36:285–8.
- <span id="page-9-4"></span>6. Sato T, Inaba N, Yamashita T. MK-7 and its effects on bone quality and strength. Nutrients. 2020;12:965.
- <span id="page-9-5"></span>7. Sato T, Schurgers LJ, Uenishi K. Comparison of menaquinone-4 and menaquinone-7 bioavailability in healthy women. Nutr J. 2012;11:93.
- <span id="page-9-6"></span>8. Bentley R, Meganathan R. Biosynthesis of vitamin K (menaquinone) in bacteria. Microbiol Rev. 1982;46:241–80.
- <span id="page-9-7"></span>9. Bentley R, Campbell I, Robins D, Kelsey M. Biosynthesis of bacterial menaquinones (vitamins K2). Biochemistry. 1971;10:3069–78.
- <span id="page-9-8"></span>10. Tani Y, Asahi S, Yamada H. Vitamin K\_2 (Menaquinone): screening of producing microorganisms and production by favobacterium meningosepticum. J Ferment Technol. 1984;62:321–7.
- <span id="page-9-9"></span>11. Chollet M, Guggisberg D, Portmann R, Risse M-C, Walther B. Determination of menaquinone production by Lactococcus spp. and propionibacteria in cheese. Int Dairy J. 2017;75:1–9.
- <span id="page-9-10"></span>12. Morishita T, Tamura N, Makino T, Kudo S. Production of menaquinones by lactic acid bacteria. J Dairy Sci. 1999;82:1897–903.
- <span id="page-9-11"></span>13. Westers L, Westers H, Quax WJ. *Bacillus subtilis* as cell factory for pharmaceutical proteins: a biotechnological approach to optimize the host organism. Biochim Biophys Acta (BBA) Mol Cell Res. 2004;1694:299–310.
- <span id="page-9-12"></span>14. Ren L, Peng C, Hu X, Han Y, Huang H. Microbial production of vitamin K2: current status and future prospects. Biotechnol Adv. 2020;39: 107453.
- <span id="page-9-16"></span>15. Berenjian A, Mahanama R, Talbot A, Biffin R, Regtop H, Valtchev P, Kavanagh J, Dehghani F. Efficient media for high menaquinone-7 production: response surface methodology approach. New Biotechnol. 2011;28:665–72.
- <span id="page-9-13"></span>16. Wu W-J, Ahn B-Y. Statistical optimization of medium components by response surface methodology to enhance menaquinone-7 (vitamin K 2) production by *Bacillus subtilis*. J Microbiol Biotechnol. 2018;28:902–8.
- <span id="page-9-14"></span>17. Song J, Liu H, Wang L, Dai J, Liu Y, Liu H, Zhao G, Wang P, Zheng Z. Enhanced production of vitamin K2 from *Bacillus subtilis* (natto) by mutation and optimization of the fermentation medium. Braz Arch Biol Technol. 2014;57:606–12.
- <span id="page-9-15"></span>18. Liu Y, Wang J, Huang J-B, Li X-F, Chen Y, Liu K, Zhao M, Huang X-L, Gao X-L, Luo Y-N. Advances in regulating vitamin K2 production through metabolic engineering strategies. World J Microbiol Biotechnol. 2024;40:8.
- <span id="page-9-17"></span>19. Chen X, Shang C, Zhang H, Sun C, Zhang G, Liu L, Li C, Li A, Du P. Efects of alkali stress on the growth and menaquinone-7 metabolism of *Bacillus subtilis* natto. Front Microbiol. 2022;13: 899802.
- <span id="page-9-18"></span>20. Berenjian A, Chan NL-C, Mahanama R, Talbot A, Regtop H, Kavanagh J, Dehghani F. Efect of bioflm formation by *Bacillus subtilis* natto on menaquinone-7 biosynthesis. Mol Biotechnol. 2013;54:371–8.
- <span id="page-9-19"></span>21. Berenjian A, Mahanama R, Talbot A, Regtop H, Kavanagh J, Dehghani F. Designing of an intensifcation process for biosynthesis and recovery of menaquinone-7. Appl Biochem Biotechnol. 2014;172:1347–57.
- <span id="page-9-20"></span>22. Berenjian A, Mahdinia E, Demirci A. Sustainable menaquinone-7 production through continuous fermentation in bioflm bioreactors. Bioprocess Biosyst Eng. 2024;47:1107.
- <span id="page-9-21"></span>23. Ranmadugala D, Ebrahiminezhad A, Manley-Harris M, Ghasemi Y, Berenjian A. High level of menaquinone-7 production by milking menaquinone-7 with biocompatible organic solvents. Curr Pharm Biotechnol. 2018;19:232–9.
- <span id="page-9-22"></span>24. Zhao C, Wan Y, Tang G, Jin Q, Zhang H, Xu Z. Comparison of diferent fermentation processes for the vitamin K2 (Menaquinone-7) production by a novel Bacillus velezensis ND strain. Process Biochem. 2021;102:33–41.
- <span id="page-9-23"></span>25. Słowik-Borowiec M, Potocki L, Oklejewicz B, Broda D, Podbielska M, Szpyrka E. Preparation of vitamin K2 MK-7 in a process of fermentation of diferent seeds and cereals by bacteria *Bacillus subtilis*. Acta Univ Cinbinesis Ser E Food Technol. 2021;25:93.
- <span id="page-9-24"></span>26. Dajanta K, Chukeatirote E, Apichartsrangkoon A. Improvement of thua nao production using protein-rich soybean and *Bacillus subtilis* TN51 starter culture. Ann Microbiol. 2012;62:785–95.
- <span id="page-9-25"></span>27. Zhang C, Wu D, Ren H. Economical production of vitamin K2 using crude glycerol from the by-product of biodiesel. Sci Rep. 2020;10:5959.
- <span id="page-9-26"></span>28. Tan H, Aziz AA, Aroua M. Glycerol production and its applications as a raw material: a review. Renew Sustain Energy Rev. 2013;27:118–27.
- <span id="page-9-27"></span>29. dos Santos RR, Muruci LNM, Santos LO, Antoniassi R, da Silva JPL, Damaso MCT. Characterization of diferent oil soapstocks and their application in the lipase production by Aspergillus niger under solid state fermentation. J Food Nutr Res. 2014;2:561–6.
- <span id="page-9-28"></span>30. Shah KKR, Patel GB. Biodegradation of soap stock: as an alternative renewable energy resource and reduce environmental pollution environmental pollution. In: Arora S, Kumar A, Ogita S, Yau Y-Y, editors. Innovations in environmental biotechnology. Singapore: Springer Nature Singapore; 2022. p. 653–76.
- <span id="page-9-29"></span>31. Xu J-z, Zhang W-g. Menaquinone-7 production from maize meal hydrolysate by Bacillus isolates with diphenylamine and analogue resistance. J Zhejiang Univ Sci B. 2017;18:462.
- <span id="page-9-30"></span>32. Wang H, Liu H, Wang L, Zhao G, Tang H, Sun X, Ni W, Yang Q, Wang P, Zheng Z. Improvement of menaquinone-7 production by *Bacillus subtilis* natto in a novel residue-free medium by increasing the redox potential. Appl Microbiol Biotechnol. 2019;103:7519–35.
- <span id="page-9-31"></span>33. Bondioli P, Della Bella L. An alternative spectrophotometric method for the determination of free glycerol in biodiesel. Eur J Lipid Sci Technol. 2005;107:153–7.
- <span id="page-9-32"></span>34. Bratbak G, Dundas I. Bacterial dry matter content and biomass estimations. Appl Environ Microbiol. 1984;48:755–7.
- <span id="page-9-33"></span>35. Mahdinia E, Demirci A, Berenjian A. Production and application of menaquinone-7 (vitamin K2): a new perspective. World J Microbiol Biotechnol. 2017;33:1–7.
- <span id="page-9-34"></span>36. Orlando P, Silvestri S, Marcheggiani F, Cirilli I, Tiano L. Menaquinone 7 stability of formulations and its relationship with purity profle. Molecules. 2019;24:829.
- <span id="page-9-35"></span>37. APHA AWWA W. Standard methods for the examination of water and wastewater 20th edition. American Public Health Association, American Water Work Association, Water Environment Federation, Washington, DC 1998.
- <span id="page-9-36"></span>38. Carranzo IV. Standard methods for examination of water and wastewater. In: Anales de hidrología médica. Universidad Complutense de Madrid. 2012; pp. 185.
- <span id="page-9-37"></span>39. .S. Environmental Protection Agency. Method 1664, revision B: n-hexane extractable material (HEM; oil and grease) and silica gel treated *n*-hexane extractable material (SGT-HEM; non-polar material) by extraction and gravimetry. Retrieve from [http://www.epa.gov/environmental-topics/](http://www.epa.gov/environmental-topics/water-topics) [water-topics.](http://www.epa.gov/environmental-topics/water-topics) 2010.
- <span id="page-9-38"></span>40. Nwaogu L, Onyeze G, Nwabueze R. Degradation of diesel oil in a polluted soil using *Bacillus subtilis*. Afr J Biotech. 1939;2008:7.
- <span id="page-10-0"></span>41. Hussein SF, Goran SM. Bioremediation of oily wastewater by using of bacteria (*Bacillus subtilis*). Zanco J Pure Appl Sci. 2020;32:206–23.
- <span id="page-10-1"></span>42. Takaç S, Marul B. Efects of lipidic carbon sources on the extracellular lipo lytic activity of a newly isolated strain of *Bacillus subtilis*. J Ind Microbiol Biotechnol. 2008;35:1019–25.
- <span id="page-10-2"></span>43. Mahdinia E, Demirci A, Berenjian A. Efects of medium components in a glycerol-based medium on vitamin K (menaquinone-7) produc tion by *Bacillus subtilis* natto in bioflm reactors. Bioprocess Biosyst Eng. 2019;42:223–32.
- <span id="page-10-3"></span>44. Luo M-M, Ren L-J, Chen S-L, Ji X-J, Huang H. Effect of media components and morphology of Bacillus natto on menaquinone-7 synthesis in sub merged fermentation. Biotechnol Bioprocess Eng. 2016;21:777–86.
- <span id="page-10-4"></span>45. Hu X-c, Liu W-m, Luo M-m, Ren L-j, Ji X-j, Huang H. Enhancing menaqui none-7 production by Bacillus natto R127 through the nutritional factors and surfactant. Appl Biochem Biotechnol. 2017;182:1630–41.
- <span id="page-10-5"></span>46. Sato T, Yamada Y, Ohtani Y, Mitsui N, Murasawa H, Araki S. Efficient production of menaquinone (vitamin K2) by a menadione-resistant mutant of *Bacillus subtilis*. J Ind Microbiol Biotechnol. 2001;26:115–20.
- <span id="page-10-6"></span>47. Zhang C, Wu D, Ren H. Economical production of vitamin K(2) using crude glycerol from the by-product of biodiesel. Sci Rep. 2020;10:5959.
- <span id="page-10-7"></span>48. Zhang C, Ren H, Zhong C. Economical production of vitamin K2 using wheat starch wastewater. J Clean Prod. 2020;270: 122486.
- <span id="page-10-8"></span>49. Fisher SH, Sonenshein AL. Control of carbon and nitrogen metabolism in *Bacillus subtilis*. Annu Rev Microbiol. 1991;45:107–35.
- <span id="page-10-9"></span>50. Sonenshein AL, Hoch JA, Losick R. *Bacillus subtilis* and its closest relatives: from genes to cells. 11th ed. Amer Society for Microbiology: Washington; 2002.
- <span id="page-10-10"></span>51. Liu Y, Zheng Z-M, Qiu H-W, Zhao G-H, Wang P, Liu H, Wang L, Li Z-M, Wu H-F, Liu H-X. Surfactant supplementation to enhance the production of vitamin K2 metabolites in shake fask cultures using Escherichia sp. mutant FM3-1709. Food Technol Biotechnol. 2014;52:269–75.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in pub lished maps and institutional afliations.