known for decades that IL-12 induces Th1 cells [interferon- $\gamma$  (IFN- $\gamma$ ) producers] and IL-4 induces Th2 cells (IL-4, IL-5 and IL-13 producers), it was only recently demonstrated that Th17 cells differentiate upon exposure to combinations of IL-1, IL-6 and transforming growth factor- $\beta$  (TGF- $\beta$ ). Interleukin-23, although not critical for the induction of the Th17 lineage, is required *in vivo* for the stabilization and proliferation of Th17 cells.<sup>4,8</sup> In addition to IL-17, Th17 cells and most 'Th17-like' innate cells produce IL-17F, tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ), IL-22 and IL-21; accordingly, in this review we will discuss the involvement of these related Th17 cytokines as they pertain to co-operative target gene regulation. We refer the reader to numerous reviews outlining mechanisms underlying Th17 differentiation.<sup>8-12</sup>

Interleukin-17 and other Th17 cytokines are linked to the pathogenesis of diverse autoimmune and inflammatory diseases (Table 1, Fig. 1). Conversely, IL-17 is essential for host defence against many microbes, particularly extracellular bacteria and fungi.<sup>13</sup> The IL-17 receptor is expressed ubiquitously, and hence most cells can potentially respond to this cytokine.<sup>14</sup> The target cell types best analysed are of non-immune origin, particularly epithelial and mesenchymal cells within diseased or inflamed tissues.<sup>15</sup> Studies have revealed IL-17-dependent activities in immune cells, particularly B lymphocytes and antigen-presenting cells (APC). Here, we aim to describe how IL-17 exerts its beneficial and its harmful properties via specific target gene regulation in the context of disease (Fig. 1).

# IL-17-mediated pathogenesis in autoimmune disease

Interleukin-17 mediates adverse effects in many autoimmune diseases. Before the discovery of the Th17 subset as a distinct CD4<sup>+</sup> effector population, it was considered that Th1, Th2 and B cells were the main mediators of pathology in autoimmunity. For example rheumatoid arthritis (RA) was widely accepted to be Th1-mediated, supported by the presence of IFN- $\gamma$  and TNF- $\alpha$  (then thought to be a Th1 cytokine) in synovial lesions and peripheral blood.<sup>16</sup> Similarly, inflammatory bowel disease (IBD) was described as a 'mixed' Th1 and Th2 pathology, with both IL-4 and IFN-y implicated.<sup>17</sup> However, during the late 1990s, studies pointed to IL-17 as a possible effector in RA and other diseases, despite the prevailing confusion as to whether IL-17 was a Th1 or 'Th0' cytokine.<sup>18,19</sup> The discovery of the Th17 cell as a bona fide T-cell subset led to a re-kindling of interest in this cytokine in the context of autoimmunity. Indeed, pre-clinical studies supporting a role for IL-17 in disease (outlined herein) led to current clinical trials designed to block IL-17, the IL-17 receptor (IL-17R) or its inducers (i.e. IL-23, IL-6) in autoimmunity.20-22

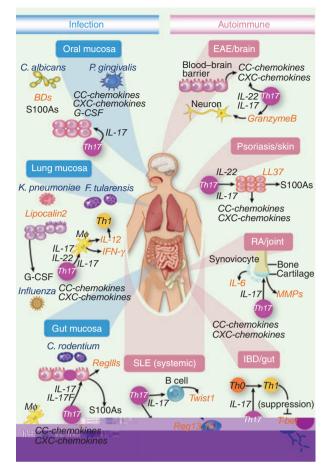


Figure 1. IL-17 signaling and target genes in various disease settings. In blue are representative mucosal infections where IL-17 plays a key role, along with key target genes involved in each. In pink are autoimmune diseases and the role of IL-17 and particular target genes therein.

## Rheumatoid arthritis

Many, if not most, autoimmune diseases are now connected in some manner to IL-17 or the Th17 pathway. In particular, RA has been intensively studied, starting even before the recognition of the Th17 subset. The key features of inflamed arthritic joints are proliferating synovial fibroblasts, joint and cartilage erosion, infiltrating CD4<sup>+</sup> T cells and autoantibody-producing plasma cells. In addition, increased numbers of innate immune cells (DC, granulocytes and macrophages), in some cases ectopic germinal centres (GC) are found within joints. Early studies showed that high levels of IL-17 were found in the rheumatoid synovium of patients with RA but not of controls or of patients with osteoarthritis. Consistently, adding IL-17 to an in vitro culture system stimulated bone resorption and collagen destruction.<sup>23</sup> Furthermore, neutralizing IL-17 or its receptor in collagen-induced arthritis mouse models resolved RA symptoms, IL-17Adeficient mice are protected from collagen-induced

which appears to promote spontaneous generation of autoreactive GC and hence elevated autoantibody production.<sup>45</sup>

Another animal model with lupus-like features is the  $Ets-1^{-/-}$  mouse, characterized by increased high-titre

expressed.<sup>43</sup> Interleukin-17 synergizes with TNF-α to promote induction of nearly all its target genes, and in many cases synergy has also been observed with IFN- $\gamma$  and IL-1 $\beta$  (reviewed in ref. <sup>27</sup>). As a consequence, IL-17 alone or together with other inflammatory cytokines in the inflamed joint mediates adverse events in RA.

#### Systemic lupus erythematosus

Systemic lupus erythematosus (SLE) is a multi-organ systemic autoimmune disease characterized by autoantibody production. Although traditionally considered mainly a B-cell disease, recent reports indicate that there is likely to be a role for IL-17 in lupus (reviewed in ref.44). Interleukin-17 is elevated in the serum of many SLE patients.45,46 The BXD2 mouse strain develops SLE-like features and spontaneous erosive arthritis with age.45 These mice produce pathogenic autoantibodies because of increased somatic hypermutation and enhanced classswitch recombination, mediated by over-expressed activation-induced cytidine deaminase in GC.47 In studies to elucidate the underlying mechanism of this phenotype, Hsu et al.45 unexpectedly discovered IL-17-dependent signalling in BXD2 B cells that led to enhanced autoantibody production and increased numbers of GCs. CD4 T cells from BXD2 mice were also more prone to differentiate into Th17 cells. The IL-17 functions in this setting by inducing genes encoding Regulator of G-protein signalling 13 and 16 (Rgs13 and Rgs16), which inhibit G-coupled protein receptors such as CXCR4. Hence, IL-17 disrupts trafficking of B cells within lymph nodes, antigens.<sup>52</sup> Recognition of gut microbes is mediated by Toll-like receptors (TLR) and also intracellular pattern recognition receptors of the nucleotide oligomerization domain (NOD) family. Strikingly, genome-wide association studies (GWAS) identified NOD2 (CARD15) as an IBD-associated gene,<sup>53,54</sup> leading to the current paradigm that IBD results from unregulated immune responses against commensal bacteria. In addition to NOD2, additional genes identified in GWAS studies have implicated the Th17 pathway, most notably the IL-23R,<sup>55</sup> which is expressed primarily on IL-17-producing cells. Other Th17-associated genes implicated in GWAS studies of IBD include

mune disease of the central nervous system. It is elicited by immunization of neuroantigens, such as myelin basic protein and proteolipid protein. As with many other autoimmune conditions, Th1 cells were long thought to be responsible for EAE pathology, despite the fact that IFN<sup>-/-</sup>, IFN $\gamma R^{-/-}$  and IL-12p35<sup>-/-</sup> mice were susceptible.<sup>77–79</sup> Landmark studies comparing the IL-12p35<sup>-/-</sup> and IL-23p19<sup>-/-</sup> mice showed clearly that the Th17 pathway was responsible for pathology.<sup>80</sup> Further evidence for the role of Th17 cells in driving EAE was shown in STAT6<sup>-/-</sup>/T-bet<sup>-/-</sup> doubly deficient mice, lacking Th1 and Th2 cells.<sup>81</sup> Furthermore, EAE in these mice could be ameliorated by treatment with anti-IL-17 antibodies. An elegant study noted that regulation of IL-17 pathogenic activity can be controlled by IL-10. Specifically, Th17 cells derived *ex vivo* following treatment with TGF- $\beta$  and IL-6 do not cause EAE upon adoptive transfer, whereas Th17 cells derived ex vivo with TGF- $\beta$ , IL-6 and IL-23 develop EAE.<sup>82</sup> Interleukin-23 suppresses IL-10, correlating with elevated expression of IL-17 target genes such as CXCL10 (IP10), CCL2, CXCL2 and CCL20.82 Both IL-17 and IL-22 also disrupt the tight junctions that form the blood-brain barrier, enabling Th17 cells to migrate into the central nervous system and cause neuronal damage.<sup>83</sup>

In summary, increasing numbers of autoimmune conditions implicate the Th17 pathway. Although the specific genes and mechanisms exhibit some variation depending on the location, chronicity and type of disease, there are many common threads in terms of IL-17-mediated gene expression. These findings have made blocking the IL-17 pathway an attractive target for anti-cytokine therapy.<sup>22</sup> On the flip side, the same pathways that promote disease in autoimmunity are beneficial in many infection settings, which will be considered in the following sections.

## Infection

In contrast to its adverse effects in autoimmunity, IL-17 plays a vital role in protecting the host from infection.<sup>13</sup> This is particularly evident at mucosal sites such as lung, gut and the oral cavity. Interleukin-17-producing cells are enriched at mucosal surfaces, and Th17 cells express the CCR6 receptor that targets them to mucosal areas.<sup>84,85</sup> Pro-inflammatory cytokines such as, IL-6, IL-1 $\beta$  and TNF- $\alpha$ , which mediate defensive responses are induced by IL-17. In particular, IL-6, acts in a positive feedback loop to further amplify Th17 differentiation and activate acute-phase responses and complement.<sup>28</sup> Interleukin-17 modulates neutrophils via cytokines that promote polymorphonuclear cell expansion and survival (G-CSF, granulocyte-macrophage CSF)86 as well as neutrophil chemoattractants (CXCL1, CXCL2 and CXCL5). Additionally, CXCL9, CXCL10 and CCL20 are target genes of IL-17, which have chemotactic activity for lymphocytes, DC and other immune cells, targeting them to mucosal surfaces; IL-17 induces CCL2 and CCL7, monocyterecruiting chemokines. Interestingly, IL-17 suppresses CCL5/RANTES, although the significance of this with respect to infection has not been determined.<sup>29,87</sup> Antimicrobial peptides, which contribute to host defence by direct killing of invading organisms, are strongly up-regulated by IL-17. Notably, some of the chemokines regulated by IL-17 also exhibit antimicrobial activity (e.g. CCL20, which binds the CCR6 receptor found on DC and Th17 cells).88,89 In the following sections, we will discuss IL-17-induced genes in representative infectious models at various mucosal sites. Not surprisingly, most of the genes identified in the context of infection are the same as those identified from autoimmune studies or cell lines, but some appear to be somewhat specific to a particular pathogen or infected tissue.

## Extracellular bacterial infections

# Klebsiella pneumoniae (lung mucosa)

The first report to describe the IL-17RA<sup>-/-</sup> mouse assessed pulmonary infection by an extracellular pathogen, Klebsiella pneumoniae.<sup>90</sup> Klebsiella pneumoniae is a Gram-negative bacterium that causes intra-abdominal and urinary tract infections, as well as hospital and community-acquired pneumonia. Infection with K. pneumoniae in IL-17RA<sup>-/-</sup> mice led to reduced survival and elevated bacterial burden. Neutrophil levels in lung were severely impaired, which was linked to reduction of CXC chemokines and G-CSF in bronchoalveolar lavage fluid. The same group demonstrated a requirement for IL-23 in defence against K. pneumoniae.<sup>91</sup> Mice that were IL-23<sup>-/-</sup> showed a significant reduction in CCL3, CXCL2, CXCL1, CXCL5 and IL-6. When it was recognized that IL-22 is also a Th17-derived cytokine, IL-22 was found to be an even more critical player than IL-17 in the setting of K. pneumoniae.<sup>92</sup> Although blocking both IL-17 and IL-22 significantly lowered G-CSF and CXCL1 expression in bronchoalveolar lavage fluid, the addition of IL-22 alone did not serve to increase G-CSF and CXCL1 expression. Rather, IL-22 affected mainly production of IL-6 and CCL3. In addition to stimulating chemokine expression, IL-17 and IL-22 induce lipocalin 2 (Lcn2, 24p3) in tracheal epithelial cells. The Lcn2 blocks catecholate-type siderophores of Gram-negative bacteria, preventing them from scavenging free iron,<sup>93,94</sup> and is potently regulated by IL-17.95 Surprisingly, however, IL-17 and IL-22 are apparently not necessary for Lcn2 induction in K. pneumoniae infections, whereas TLR4 and Myd88 are essential.<sup>96</sup>

## Citrobacter rodentium (gut mucosa)

Interleukin-17-expressing cells are particularly abundant in the gut, including classic  $\alpha\beta$ -T cells as well as more innate cells such as  $\gamma\delta$ -T and natural killer T cells. Citrobacter rodentium is a naturally occurring murine enteric pathogen, and is an extracellular Gram-negative organism considered a model for enteropathic attaching and effacing Escherichia coli infections. In model studies of C. rodentium infection, IL-17 did not play a significant protective role compared with IL-22 at early stages of disease.97 Infection with C. rodentium induces a variety of antimicrobial peptides, such as S100A8, S100A9, RegIII $\beta$  and RegIII $\gamma$ . RegIII $\beta$  and RegIII $\gamma$  are C-type lectins originally thought to selectively kill Gram-positive bacteria. Not surprisingly, adding back RegIIIy protected mice significantly. In contrast, direct comparison of IL-17A<sup>-/-</sup> versus IL-17F<sup>-/-</sup> mice suggested that IL-17 and IL-17F do contribute to protection from C. rodentium in later stages.<sup>98</sup> Here, CXCL1, CXCL2, IFN- $\gamma$ , IL-1 $\beta$ , IL-6, TNF- $\alpha$  and inducible nitric oxide synthase were regulated normally in the colons of IL-17A<sup>-/-</sup>, IL-17F<sup>-/-</sup> and IL-17A/F<sup>-/-</sup> mice following C. rodentium infection. Similarly, IL-17A<sup>-/-</sup> and IL-17F<sup>-/-</sup> mice showed increased expression of BD2, Lcn2, S100A8, S100A9, RegIII and RegIIIy compared with wild-type mice, yet these mice still had a higher bacterial burden. However, BD1, BD3 and BD4 were impaired in the knockout strains, implying that in this setting they may be the major IL-17 gene targets.

## Porphyromonas gingivalis (oral mucosa)

The oral cavity is another important but often overlooked site of mucosal infection. Studies of a major human periodontal pathogen revealed a strongly protective role for IL-17 receptor signalling, at least in acute oral infections. Porphyromonas gingivalis is a Gram-negative anaerobic microbe that is one of the three major pathogens associated with periodontal disease, characterized by gingival tissue destruction, chronic infection and bone loss in the alveolar bone crest of the jaw. Despite the potential of IL-17 to promote bone destruction as it does in the context of RA, IL-17RA<sup>-/-</sup> mice are more susceptible to infection than wild-type mice. The major IL-17 gene targets shown to be impaired were CXCL1, CXCL2 and CXCL5,<sup>99,100</sup> which correlated with reduced neutrophil recruitment to the gingival area. Consistently, CXCR2<sup>-/-</sup> mice (the receptor for CXCL1, CXCL2 and CXCL5) are exquisitely susceptible to P. gingivalis-induced periodontal disease. Interestingly, these studies also revealed a gender bias for IL-17R activity. Regardless of genetic background, female IL-17RA<sup>-/-</sup> mice were more susceptible than males to P. gingivalis-induced bone loss. Although the female mice showed a more dramatic reduction in CXC chemokine expression, the male mice were mainly impaired in G-CSF. Consequently, although both genders had reduced neutrophil activity, this appeared to be directed through different mechanisms.99

# Intracellular bacterial infections

It is commonly considered that IL-17 is primarily important for protection against extracellular pathogens rather than intracellular bacteria. Indeed, in many cases, but not all, IL-17 is dispensable for host defence against intracellular bacteria.<sup>13</sup> For example, Listeria monocytogenes induces IL-17 in liver  $\gamma \delta$ -T cells, which is associated with enhanced neutrophil recruitment;<sup>101</sup> however, IL-17RA<sup>-/-</sup> mice have no defect in survival, arguing against a role for IL-17 in this disease.<sup>101</sup> Mycobacterium tuberculosis infection studies also revealed critical roles of IFN-y rather than IL-17 in primary infections, although IL-17 is induced during infection.<sup>102,103</sup> Nonetheless, the picture is more complex than this because IL-17 is essential for an effective vaccine-induced response to M. tuberculosis.<sup>104</sup> Although the detailed mechanisms need to be defined more carefully, IL-17 may contribute more to the development of memory responses than to initial infections for some intracellular organisms.<sup>104</sup> In contrast, infection with Francisella tularensis, another intracellular organism, shows a requirement for IL-17.<sup>105</sup> Mechanistically, this is regulated via IL-17-mediated induction of IFN-y and IL-12 in macrophages, linking Th17 and Th1 responses in vivo. Similarly, responses to Chlamydia infection also involve IL-17.<sup>106</sup> Therefore, the concept that IL-17 activity is exclusively linked to protection from extracellular pathogens is overly simplistic.

## Fungal infections

In the setting of fungal infections, IL-17 plays both protective and destructive roles depending on route of infection and perhaps the morphological form of the organism. The primary organism where this has been examined is Candida albicans, a common commensal that colonizes human mucosal surfaces. Candida albicans causes both systemic and mucosal infections, but the immune responses at these sites are quite different.<sup>107</sup> The most severe form of candidiasis is a disseminated disease associated with hospital settings, which is modelled by intravenous injection of C. albicans into mice. The IL-17RA<sup>-/-</sup> mice are highly susceptible to this form of disease. Although detailed analysis of relevant target genes was not performed, neutrophil recruitment was severely defective.108 Consistent with this, two reports using mouse models of oropharyngeal candidiasis ('thrush') showed that IL-17 and IL-23 play a strongly protective role against mucosal C. albicans.<sup>109,110</sup> Microarray analysis showed that immunocompetent wild-type mice infected with C. albicans up-regulate numerous classic IL-17 target genes in the oral mucosa, including BD3, S100A8/9, MMP-8, G-CSF, CCL20, IL-6, CXCL1 and CXCL5.109 BD3 and S100A8/9, in particular, were strongly impaired in IL-17RA<sup>-/-</sup> tongue tissue. In contrast, few IFN- $\nu$  signature genes were induced in this setting, arguing against a role for Th1 cells.<sup>109</sup> Studies of the pattern recognition receptors involved in early activation events indicate that C-type lectin receptors such as Dectin 1, Dectin 2 and the Mannose receptor as well as the NLRP3 inflammasome induce IL-23 from macrophages and DC, hence promoting Th17 cell differentiation (reviewed in ref.<sup>111</sup>). These findings are supported by studies in humans with mutations in STAT3 (who are selectively Th17-deficient), who are exquisitely susceptible to mucosal candidiasis and staphylococcal infections.<sup>112</sup>

In contrast, in a gastrointestinal model of *C. albicans* and *Aspergillus fumigatus* mucosal infection induced by injection of fungi into the gut, Romani and colleagues<sup>113</sup> reported that IFN- $\gamma$  plays a primary role in protection, whereas IL-23 and IL-17 exacerbate inflammation. However, when IFN- $\gamma$  is absent, IL-23 plays a protective role via mechanisms involving cross-regulation of IL-12 and IL-23.<sup>113</sup> A caveat of this model is that it does not have a true parallel human disease state, but may nonetheless reflect the complexity of immune responses that differ by anatomic location.

# Viral infections

Viral host defence depends heavily on Type I IFNs that modulate viral replication, and so IL-17 is considered to be relatively less important. However, there is emerging evidence that IL-17 may participate in viral immune responses, which can be beneficial or detrimental to the host. It is also intriguing that a homologue of IL-17 is encoded in a Herpesvirus saimiri, a T-cell tropic  $\gamma$  herpesvirus.<sup>114</sup> The significance of this is unknown, although viral IL-17 promotes positive signalling through IL-17RA.<sup>114</sup> Nonetheless, this homologue presumably benefits the virus in some aspect of pathogenesis, by some as yet unknown mechanism.

In poxvirus infections, genetically engineered recombinant vaccinia viruses (VV) encoding IL-12, IL-23 or IL-17 were created to test the roles of these cytokines in viral host defence. Surprisingly, VV-IL-23 and VV-IL-17, but not VV-IL-12, caused reduced virulence in mice.<sup>115</sup> Although the mechanism was not well defined, protection was not mediated by enhanced cytotoxic lymphocyte activity. Surprisingly, IL-23-induced viral resistance was also not primarily the result of IL-17, as IL-17<sup>-/-</sup> mice infected with VV-IL-23 were not significantly compromised.<sup>115</sup> Interleukin-22 and other Th17-hallmark cytokines were not evaluated in this study. In a contrasting report, an IL-17-expressing VV was found to be more virulent than its parental virus in mice, associated with altered immunoglobulin G isotype generation.<sup>116</sup> The distinctions between these models is unclear, but may reflect the fine line between host defence and immunopathology mediated by IL-17.

Interleukin-17 signalling may be counterproductive in certain viral settings, by contributing to the 'cytokine

storm' that characterizes disease pathology. In an influenza infection model, IL-17 and IL-17F were induced as soon as 2 days post-infection. The survival rate of IL-17RA<sup>-/-</sup> mice was higher than WT, associated with reduction in neutroophil chemokines and inflammatory cytokines (G-CSF, CXCL1, IL-6, TNF- $\alpha$ , IL-1 $\beta$  and IFN- $\gamma$ ). Lung injury in this setting may be partly the result of IL-17-mediated oxidization of phospholipids by recruited neutrophils.<sup>117</sup>

Another report identified a function of IL-17 in maintaining virus persistence in a model using Theiler's murine encephalomyelitis virus (TMEV),<sup>118</sup> which leads to a demyelinating disease. In a susceptible mouse strain, Th17 development was elevated during TMEV infection, and Th17 cells infiltrated into the central nervous system. Viral persistence in brain and spinal cord and production of IL-6, CXCL1 and MCP-1 correlated with the appearance of Th17 cells.<sup>118</sup> Importantly, the mechanism by which the virus persists was because of the resistance to apoptosis of infected astrocytes. Interestingly, IL-17F did not mediate an anti-apoptotic effect. Collectively, these findings indicate that infection by viruses often induces Th17 differentiation and so its signature target genes, yet this is often ineffective in terms of host defence, and may even promote immunopathology.

# IL-17 signalling: synergy and mechanisms

Mechanisms of IL-17 signalling are poorly described compared with other cytokine receptor subfamilies. Although reviewed in detail elsewhere,<sup>2,27,51</sup> IL-17 binds to both the IL-17RA and IL-17RC subunits to mediate signalling.<sup>119</sup> Both receptors encode a conserved signalling motif known as the SEFIR (SEF/IL17R) domain,<sup>120</sup> which engages the Act1 adaptor/ubiquitin ligase enzyme through its own SE-FIR motif.<sup>121–123</sup> Act1 in turn recruits TRAF6, which leads to activation of the nuclear factor- $\kappa$ B (NF- $\kappa$ B) pathway. Act1 is also upstream of the CCAAT/Enhancer Binding Protein (C/EBP)- $\beta$  and C/EBP- $\delta$  and mitogen-activated protein kinase pathways, all of which act in concert to control target gene expression. Most IL-17 downstream genes have NF- $\kappa$ B and C/EBP binding sites, and in many cases both are necessary for IL-17-mediated promoter activity.<sup>95,124</sup>

As mentioned previously, a notable feature of IL-17 is its strong co-operative effect with other cytokines in regulating down-stream gene/protein expression. Interleukin-17 has been shown to synergize with IL-1 $\beta$ , IL-22, IFN- $\gamma$ , TNF- $\alpha$ , Oncostatin M, CD40, BAFF and Vitamin D3 (1,25-dihydroxyvitamin D3), and this list may grow<sup>29,73,75,125,126</sup> (Table 2). This synergy is reflected in the fact that IL-17 alone is not a potent inducer of inflammatory pathways such as NF- $\kappa$ B, despite the potent *in vivo* effects of an IL-17 deficiency (reviewed in ref.<sup>2</sup>). The molecular mechanisms underlying synergistic signalling are not fully elucidated, although various pathways are implicated. Many IL-17 target genes are controlled post-transcriptionally by messenger RNA stabilization. In particular, CXCL1, CXCL2, IL-6, I-KBζ and CXCL5 messenger RNAs are induced by IL-17 and/or TNF-a somewhat weakly and are subject to rapid degradation, but in the presence of both IL-17 and TNF- $\alpha$  their stability is significantly enhanced.<sup>127</sup> Mechanistically this is mediated by the mitogen-activated protein kinase pathway and Act1, but surprisingly not by TRAF6. At the promoter level, co-operative induction of C/EBP proteins but not NF- $\kappa$ B by the combination of IL-17 and TNF- $\alpha$  contributes to co-operative induction of the IL-6 promoter.<sup>124</sup> The synergy between IL-17 and BAFF, however, is mediated by the NF- $\kappa$ B pathway, where IL-17RA out-competes the BAFFR for Act1, leading to positive rather than suppressive signalling.<sup>46</sup> Interleukin-17 has also been shown to regulate the TNF receptor, which may account for enhanced TNF- $\alpha$  signalling capacity in the presence of IL-17.<sup>128</sup> Similarly, the synergy between IL-17 and IL-19, IL-20 and IL-24 was dependent on the expression of IL-22 receptor.<sup>125</sup> Finally, the mechanisms by which IL-17 and IL-22 synergize to regulate antimicrobial peptides such as S100A8/9 and IL-20 family members still need to be determined. In summary, although IL-17 is generally a weak inducer of target genes, this cytokine has a major impact in vivo. Therefore, the synergistic effects of IL-17 almost certainly play a significant role in dictating its physiological activities.

## **Conclusion and perspectives**

As suggested by ubiquitous IL-17R expression,<sup>14</sup> IL-17 plays a role in multiple cell types and conditions. Initial studies revealed the IL-17 target genes from mesenchymal and epithelial cells, but recent work has shown important IL-17 target genes in lymphocytes and other immune cells as well. Defining the complete range of IL-17 effects *in vivo* will probably never be fully achieved, but it is clear that this cytokine is a central player in numerous disease states.

#### Acknowledgement

S.L.G. is supported by the National Institutes of Health (AR054389), the Alliance for Lupus Research and Amgen.

#### Disclosures

SLG has received a research grant and honoraria from Amgen.

## References

- Aggarwal S, Gurney AL. IL-17: a prototype member of an emerging family. J Leukoc Biol 2002; 71:1–8.
- 2 Gaffen SL. Structure and signalling in the IL-17 receptor family. *Nat Rev Immunol* 2009; **9**:556–67.

- 3 Fossiez F, Djossou O, Chomarat P et al. T cell interleukin-17 induces stromal cells to produce proinflammatory and hematopoietic cytokines. J Exp Med 1996; 183:2593– 603.
- 4 Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 cells. Annu Rev Immunol 2009; 27:485–517.
- 5 Park H, Li Z, Yang X et al. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. Nat Immunol 2005; 6:1133–41.
- 6 Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, Weaver CT. Interleukin 17-producing CD4<sup>+</sup> effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* 2005; 6:1123–32.
- 7 Steinman L. A brief history of T(H)17, the first major revision in the T(H)1/T(H)2 hypothesis of T cell-mediated tissue damage. *Nat Med* 2007; 13:139–45.
- 8 Stockinger B, Veldhoen M. Differentiation and function of Th17 T cells. Curr Opin Immunol 2007; 19:281–6.
- 9 Weaver CT, Hatton RD, Mangan PR, Harrington LE. IL-17 family cytokines and the expanding diversity of effector T cell lineages. Annu Rev Immunol 2007; 25:821–52.
- 10 McGeachy MJ, Cua DJ. Th17 cell differentiation: the long and winding road. Immunity 2008; 28:445–53.
- 11 Bluestone JA, Mackay CR, O'shea JJ, Stockinger B. The functional plasticity of T cell subsets. Nat Rev Immunol 2009; 9:811–6.
- 12 Dong C. TH17 cells in development: an updated view of their molecular identity and genetic programming. Nat Rev Immunol 2008; 8:337–48.
- 13 O'Quinn D, Palmer M, Lee Y, Weaver C. Emergence of the Th17 pathway and its role in host defense. Adv Immunol 2008; 99:115–63.
- 14 Yao Z, Spriggs M, Derry J et al. Molecular characterization of the human interleukin (IL)-17 receptor. Cytokine 1997; 9:794–800.
- 15 Yu J, Gaffen SL. Interleukin-17: a novel inflammatory cytokine that bridges innate and adaptive immunity. *Front Biosci* 2008; 13:170–7.
- 16 Firestein GS. Evolving concepts of rheumatoid arthritis. Nature 2003; 423:356-61.
- 17 Brand S. Crohn's disease: Th1, Th17 or both? The change of a paradigm: new immunological and genetic insights implicate Th17 cells in the pathogenesis of Crohn's disease Gut 2009; 58:1152–67.
- 18 Albanesi C, Cavani A, Girolomoni G. IL-17 is produced by nickel-specific T lymphocytes and regulates ICAM-1 expression and chemokine production in human keratinocytes: synergistic or antagonistic effects with IFN-γ and TNF-α. J Immunol 1999; 162:494–502.
- 19 Aarvak T, Chabaud M, Miossee P, Natvig JB. IL-17 is produced by some proinflammatory Th1/Th0 cells but not by Th2 cells. J Immunol 1999; 162:1246–51.
- 20 Gaffen SL. The role of interleukin-17 in the pathogenesis of rheumatoid arthritis. Curr Rheumatol Rep 2009; 11:365–70.
- 21 Brennan FM, McInnes IB. Evidence that cytokines play a role in rheumatoid arthritis. J Clin Invest 2008; 118:3537–45.
- 22 Lubberts E. IL-17/Th17 targeting: on the road to prevent chronic destructive arthritis? Cytokine 2008; 41:84–91.
- 23 Kotake S, Udagawa N, Takahashi N et al. IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. J Clin Invest 1999; 103:1345–52.
- 24 Lubberts E, Koenders MI, Oppers-Walgreen B, van den Bersselaar L, Coenen-de Roo CJ, Joosten LA, van den Berg WB. Treatment with a neutralizing anti-murine interleukin-17 antibody after the onset of collagen-induced arthritis reduces joint inflammation, cartilage destruction, and bone erosion. Arthritis Rheum 2004; 50:650–9
- 25 Nakae S, Nambu A, Sudo K, Iwakura Y. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. J Immunol 2003; 171:6173–7.
- 26 Lubberts E, Joosten LA, van de Loo FA, Schwarzenberger P, Kolls J, van den Berg WB. Overexpression of IL-17 in the knee joint of collagen type II immunized mice promotes collagen arthritis and aggravates joint destruction. *Inflamm Res* 2002; 51:102–4.
- 27 Shen F, Gaffen S. Structure-function relationships in the IL-17 receptor: implications for signal transduction and therapy. *Cytokine* 2008; **41**:92–104.
- 28 Ogura H, Murakami M, Okuyama Y et al. Interleukin-17 promotes autoimmunity by triggering a positive-feedback loop via interleukin-6 induction. *Immunity* 2008; 29:628–36.
- 29 Shen F, Ruddy M, Plamondon P, Gaffen S. Cytokines link osteoblasts and inflammation: microarray analysis of interleukin-17- and TNF-alpha-induced genes in bone cells. J Leukoc Biol 2005; 77:388–99.
- 30 Takemura S, Braun A, Crowson C, Kurtin PJ, Cofield RH, O'Fallon WM, Goronzy JJ, Weyand CM. Lymphoid neogenesis in rheumatoid synovitis. J Immunol 2001; 167:1072–80.
- 31 Timmer TCG, Baltus B, Vondenhoff M, Huizinga TWJ, Tak PP, Verweij CL, Mebius RE, van der Pouw Kraan TCTM. Inflammation and ectopic lymphoid structures in rheumatoid arthritis synovial tissues dissected by genomics technology: identification

of the interleukin-7 signaling pathway in tissues with lymphoid neogenesis. Arthritis Rheum 2007; 56:2492-502.

- 32 Weyand CM, Goronzy JJ. Ectopic germinal center formation in rheumatoid synovitis. *Ann N Y Acad Sci* 2003; **987**:140–9.
- 33 Chabaud M, Garnero P, Dayer JM, Guerne PA, Fossiez F, Miossec P. Contribution of interleukin 17 to synovium matrix destruction in rheumatoid arthritis. *Cytokine* 2000; 12:1092–9.
- 34 Koenders MI, Kolls JK, Oppers-Walgreen B et al. Interleukin-17 receptor deficiency results in impaired synovial expression of interleukin-1 and matrix metalloproteinases 3, 9, and 13 and prevents cartilage destruction during chronic reactivated streptococcal cell wall-induced arthritis. Arthritis Rheum 2005; 52:3239–47.
- 35 Koenders MI, Lubberts E, Oppers-Walgreen B et al. Blocking of interleukin-17 during reactivation of experimental arthritis prevents joint inflammation and bone erosion by decreasing RANKL and interleukin-1. Am J Pathol 2005; 167:141–9.
- 36 Sylvester J, Liacini A, Li WQ, Zafarullah M. Interleukin-17 signal transduction pathways implicated in inducing matrix metalloproteinase-3, -13 and aggrecanase-1 genes in articular chondrocytes. *Cell Signal* 2004; 16:469–76.
- 37 Rifas L, Arackal S. T cells regulate the expression of matrix metalloproteinase in human osteoblasts via a dual mitogen-activated protein kinase mechanism. Arthritis Rheum 2003; 48:993–1001.

- 87 Andoh A, Fujino S, Bamba S, Araki Y, Okuno T, Bamba T, Fujiyama Y. IL-17 selectively down-regulates TNF-alpha-induced RANTES gene expression in human colonic subepithelial myofibroblasts. J Immunol 2002; 169:1683–7.
- 88 Yang D, Chen Q, Hoover D, Staley P, Tucker K, Lubkowski J, Oppenheim J. Many chemokines including CCL20/MIP-3alpha display antimicrobial activity. J Leukoc Biol 2003; 74:448–55.
- 89 Kolls J, McCray P, Chan Y. Cytokine-mediated regulation of antimicrobial proteins. Nat Rev Immunol 2008; 8:829–35.
- 90 Ye P, Rodriguez FH, Kanaly S et al. Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense. J Exp Med 2001; 194:519–27.
- 91 Happel K, Dubin P, Zheng M et al. Divergent roles of IL-23 and IL-12 in host defense against Klebsiella pneumoniae. J Exp Med 2005; 202:761–9.
- 92 Aujla S, Chan Y, Zheng M et al. IL-22 mediates mucosal host defense against Gramnegative bacterial pneumonia. Nat Med 2008; 14:275–81.
- 93 Berger T, Togawa A, Duncan G et al. Lipocalin 2-deficient mice exhibit increased sensitivity to Escherichia coli infection but not to ischemia-reperfusion injury. Proc Natl Acad Sci U S A 2006; 103:1834–9.
- 94 Flo T, Smith K, Sato S, Rodriguez D, Holmes M, Strong R, Akira S, Aderem A. Lipocalin 2 mediates an innate immune response to bacterial infection by sequestrating iron. *Nature* 2004; 432:917–21.
- 95 Shen F, Hu Z, Goswami J, Gaffen S. Identification of common transcriptional regulatory elements in interleukin-17 target genes. J Biol Chem 2006; 281:24138–48.
- 96 Chan YR, Liu JS, Pociask DA et al. Lipocalin 2 is required for pulmonary host defense against Klebsiella infection. J Immunol 2009; 182:4947–56.
- 97 Zheng Y, Valdez P, Danilenko D et al. Interleukin-22 mediates early host defense against attaching and effacing bacterial pathogens. Nat Med 2008; 14:282–9.
- 98 Ishigame H, Kakuta S, Nagai T *et al.* Differential roles of interleukin-17A and -17F in host defense against mucoepithelial bacterial infection and allergic responses. *Immu*nity 2009; 30:108–19.
- 99 Yu JJ, Ruddy MJ, Conti HR, Boonanantanasarn K, Gaffen SL. The interleukin-17 receptor plays a gender-dependent role in host protection against *Porphyromonas* gingivalis-induced periodontal bone loss. *Infect Immun* 2008; 76:4206–13.
- 100 Yu J, Ruddy M, Wong G, Sfintescu C, Baker P, Smith J, Evans R, Gaffen S. An essential role for IL-17 in preventing pathogen-initiated bone destruction: recruitment of neutrophils to inflamed bone requires IL-17 receptor-dependent signals. *Blood* 2007; 109:3794–802.
- 101 Hamada S, Umemura M, Shiono T et al. IL-17A produced by gammadelta T cells plays a critical role in innate immunity against *Listeria monocytogenes* infection in the liver. J Immunol 2008; 181:3456–63.
- 102 Lockhart E, Green AM, Flynn JL. IL-17 production is dominated by gammadelta T cells rather than CD4 T cells during *Mycobacterium tuberculosis* infection. J Immunol 2006; 177:4662–9.
- 103 Khader SA, Pearl JE, Sakamoto K et al. IL-23 compensates for the absence of IL-12p70 and is essential for the IL-17 response during tuberculosis but is dispensable for protection and antigen-specific IFN-gamma responses if IL-12p70 is available. *J Immunol* 2005; 175:788–95.
- 104 Khader S, Bell G, Pearl J et al. IL-23 and IL-17 in the establishment of protective pulmonary CD4<sup>+</sup> T cell responses after vaccination and during *Mycobacterium tuber*culosis challenge. Nat Immunol 2007; 8:369–77.
- 105 Lin Y, Ritchea S, Logar A et al. IL-17 is required for Th1 immunity and host resistance to the intracellular pathogen Francisella tularensis LVS. Immunity 2009; 5:799– 810.
- 106 Zhang X, Gao L, Lei L et al. A MyD88-dependent early IL-17 production protects mice against airway infection with the obligate intracellular pathogen Chlamydia muridarum. J Immunol 2009; 183:1291–300.

- 107 Dongari-Bagtoglou A, Fidel P. The host cytokine responses and protective immunity in oropharyngeal candidiasis. J Dent Res 2005; 84:966–77.
- 108 Huang W, Na L, Fidel P, Schwarzenberger P. Requirement of interleukin-17A for systemic anti-Candida albicans host defense in mice. J Infect Dis 2004; 190:624–31.
- 109 Conti H, Shen F, Nayyar N et al. Th17 cells and IL-17 receptor signaling are essential for mucosal host defense against oral candidiasis. J Exp Med 2009; 206:299–311.
- 110 Hise AG, Tomalka J, Ganesan S, Patel K, Hall BA, Brown GD, Fitzgerald KA. An essential role for the NLRP3 inflammasome in host defense against the human fungal pathogen *Candida albicans. Cell Host Microbe* 2009; 5:487–97.
- 111 Netea MG, Brown GD, Kullberg BJ, Gow NA. An integrated model of the recognition of *Candida albicans* by the innate immune system. *Nat Rev Microbiol* 2008; 6:67–78.
- 112 Tangye SG, Cook MC, Fulcher DA. Insights into the role of STAT3 in human lymphocyte differentiation as revealed by the hyper-IgE syndrome. J Immunol 2009; 182:21–8.
- 113 Zelante T, De Luca A, Bonifazi P et al. IL-23 and the Th17 pathway promote inflammation and impair antifungal immune resistance. Eur J Immunol 2007; 37:2695–706.
- 114 Yao Z, Fanslow WC, Seldin MF, Rousseau A-M, Painter SL, Comeau MR, Cohen JI, Spriggs MK. Herpesvirus Saimiri encodes a new cytokine, IL-17, which binds to a novel cytokine receptor. *Immunity* 1995; 3:811–21.
- 115 Kohyama S, Ohno S, Isoda A et al. IL-23 enhances host defense against vaccinia virus infection via a mechanism partly involving IL-17. J Immunol 2007; 179:3917–25.
- 116 Patera AC, Pesnicak L, Bertin J, Cohen JI. Interleukin 17 modulates the immune response to vaccinia virus infection. Virology 2002; 299:56–63.
- 117 Crowe CR, Chen K, Pociask DA et al. Critical role of IL-17RA in immunopathology of influenza infection. J Immunol 2009; 183:5301–10.
- 118 Hou W, Kang HS, Kim BS. Th17 cells enhance viral persistence and inhibit T cell cytotoxicity in a model of chronic virus infection. J Exp Med 2009; 206:313–28.
- 119 Toy D, Kugler D, Wolfson M, Vanden Bos T, Gurgel J, Derry J, Tocker J, Peschon J. Cutting edge: interleukin 17 signals through a heteromeric receptor complex. J Immunol 2006; 177:36–9.
- 120 Novatchkova M, Leibbrandt A, Werzowa J, Neubuser A, Eisenhaber F. The STIRdomain superfamily in signal transduction, development and immunity. *Trends Biochem Sci* 2003; 28:226–9.
- 121 Qian Y, Liu C, Hartupee J et al. The adaptor Act1 is required for interleukin 17dependent signaling associated with autoimmune and inflammatory disease. Nat Immunol 2007; 8:247–56.
- 122 Chang S, Park H, Dong C. Act1 adaptor protein is an immediate and essential signaling component of interleukin-17 receptor. J Biol Chem 2006; 281:35603–7.
- 123 Liu C, Qian W, Qian Y et al. Act1, a U-box E3 ubiquitin ligase for IL-17 signaling. Sci Signal 2009; 2:ra63.
- 124 Ruddy M, Wong G, Liu X, Yamamoto H, Kasayama S, Kirkwood K, Gaffen S. Functional cooperation between interleukin-17 and tumor necrosis factor-alpha is mediated by CCAAT/enhancer-binding protein family members. J Biol Chem 2004; 279:2559–67.
- 125 Tohyama M, Hanakawa Y, Shirakata Y et al. IL-17 and IL-22 mediate IL-20 subfamily cytokine production in cultured keratinocytes via increased IL-22 receptor expression. *Eur J Immunol* 2009; **39**:2779–88.
- 126 Woltman AM, de Haij S, Boonstra JG, Gobin SJ, Daha MR, van Kooten C. Interleukin-17 and CD40-ligand synergistically enhance cytokine and chemokine production by renal epithelial cells. J Am Soc Nephrol 2000; 11:2044–55.
- 127 Hartupee J, Liu C, Novotny M, Li X, Hamilton T. IL-17 enhances chemokine gene expression through mRNA stabilization. J Immunol 2007; 179:4135–41.
- 128 Zrioual S, Ecochard R, Tournadre A, Lenief V, Cazalis M-A, Miossec P. Genome-wide comparison between IL-17A- and IL-17F-induced effects in human rheumatoid arthritis synoviocytes. J Immunol 2009; 182:3112–20.