

WNT enhancing signals in pancreatic cancer are transmitted by LGR6

Jing Wang^{1,2}, Dominik T. Koch¹, Felix O. Hofmann¹, Daniel Härtwig¹, Iris Beirith¹, Klaus Peter Janssen³, Alexandr V. Bazhin^{1,4,5}, Hanno Niess¹, Jens Werner^{1,4,5}, Bernhard W. Renz^{1,4,5*}, Matthias Ilmer^{1,4,5*}

¹Department of General, Visceral and Transplantation Surgery, Hospital of the University of Munich, Ludwig-Maximilians-University (LMU), Munich, Bavaria, Germany

²Department of General Surgery, The First Affiliated Hospital of USTC, Division of Life Sciences and Medicine, University of Science and Technology of China, Hefei, Anhui, P.R. China

³Department of Surgery, School of Medicine, Klinikum Rechts der Isar, Technical University of Munich, Munich, Bavaria, Germany

⁴German Cancer Consortium (DKTK), Partner Site Munich, Munich, Bavaria, Germany

⁵Bavarian Cancer Research Center (BZKF), LMU Munich, Munich, Bavaria, Germany

*Equal contribution

Correspondence to: Matthias Ilmer; email: mailmer@med.lmu.de

Keywords: pancreatic ductal adenocarcinoma, WNT signaling, epithelial-mesenchymal transition, LGR6, cancer stem cells

Received: April 3, 2023

Accepted: July 17, 2023

Published: September 27, 2023

Copyright: © 2023 Wang et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/3.0/) (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

The G-protein-coupled receptor LGR6 associates with ligands of the R-Spondin (RSPO) family to potentiate preexisting signals of the canonical WNT pathway. However, its importance in pancreatic ductal adenocarcinoma (PDAC) remains unclear. Here, we show that LGR6 is differentially expressed in various PDAC cell lines of mesenchymal and epithelial phenotype, respectively, siding with the latter subsets. LGR6 expression is altered based upon the cells' WNT activation status. Furthermore, extrinsic enhancement of WNT pathway signaling increased LGR6 expression suggestive of a reinforcing self-regulatory loop in highly WNT susceptible cells. Downregulation of LGR6 on the other hand, seemed to tamper those effects. Last, downregulation of LGR6 reduced cancer stemness as determined by functional *in vitro* assays. These findings shed new insights into regulatory mechanisms for the canonical WNT pathway in pancreatic cancer cells. It may also have potential value for treatment stratification of PDAC.

INTRODUCTION

Pancreatic ductal adenocarcinoma (PDAC) is one of the most aggressive malignancies with an overall 5-year survival rate of 10% for all stages combined [1, 2]. By 2030, current predictions see the disease as the second most common cause of cancer-related death [3]. Regulation of PDAC on a cellular level during early and later pancreatic tumorigenesis remains incompletely

understood and hence, presents challenges for better treatment options. Precise regulation of the canonical WNT signaling pathway is fundamental for normal development on the one hand as well as tissue regeneration of various origins on the other hand [4, 5]. WNT signaling has also been shown to conduct critical functions in physiologic pancreatic development [6, 7]. Although aberrant WNT signaling has been linked to tumorigenesis in multiple organs [8–11], typical

activating mutations, such as in APC or β -catenin, are not commonly seen in PDAC cells with the exception of RNF43 [12, 13]. However, it was shown in some pancreatic tumors that an elevated nuclear accumulation of β -catenin indicative of activated WNT signaling correlated with progression of PDAC [14, 15].

The WNT pathway gets activated upon the engagement of canonical WNT ligands to their co-receptors low-density lipoprotein receptor-related protein (LRP) 5 or 6 and frizzled. Intracellularly, the transcription factor β -catenin escapes from its destruction complex in the cytosol comprised of APC, AXIN2, and GSK3 β . Protein kinase A (PKA) phosphorylates β -catenin at Ser⁶⁷⁵ and thereby induces its subsequent accumulation and translocation into the nucleus, where it associates with TCF/LEF binding sites (T-cell factor / lymphoid enhancer factor) regulating canonical WNT target gene expression [16, 17]. The R-spondin (RSPO) family is a group of secreted factors which enhance a previously activated WNT signal by binding to Leucine-rich repeat containing G-protein coupled Receptors 4/5/6 (LGR4/5/6). This mechanism normally implicates the interaction of the LGR/RSPO-complex with transmembrane E3 ubiquitin ligases ZNRF3/RNF43 [18]. These ligases execute negative feedback on canonical WNT activity by promoting the ubiquitination and hence, the inactivation of the WNT co-receptors frizzled and LRP6 [18, 19]. Upon the association of RSPO with LGR, ZNRF3/RNF43 is removed from the cell surface and subsequently, the brakes on WNT signaling are released [18].

One process by which epithelial cells disconnect from each other and transdifferentiate into mesenchymal cells is the so-called epithelial–mesenchymal transition (EMT) [20–22]. EMT and canonical WNT signaling are intertwined through the key mediator β -catenin, which is also part of the cell-cell adhesion complex that consists of different catenin proteins (α , β) and E-cadherin [23, 24]. In this regard, E-cadherin acts as a negative regulator of WNT signaling through its recruitment of β -catenin to adherence junctions. Upon induction of EMT, loss of E-cadherin leads to release of β -catenin and its accumulation in the nucleus, which subsequently activates WNT signaling [25–27]. Meanwhile, high WNT activity has been shown to activate the EMT transcription factors SNAIL2 directly or ZEB1 indirectly to induce EMT, suggesting a feed-forward loop of when cancer cells undergo processes of dedifferentiation [28].

LGR5 has been serially described as a WNT target gene as well as a marker for (cancer) stem cells in multiple neoplasms [29–31]. Whether its homologue LGR6 incorporates similar functional aspects, still remains to be answered. In this work, we aimed to decipher the

functions of LGR6 in WNT signaling of PDAC, apart from its assumed assignment as a receptor to RSPO. Taken into account the connections between WNT signaling and EMT, we further hypothesized a likely interplay of LGR6 and EMT.

MATERIALS AND METHODS

Cell culture

Human pancreatic cancer cell lines (BxPC3, Capan2, MiaPaCa2, Panc1) were obtained from American Type Culture Collection (ATCC). Cells were cultured in RPMI1640 (Lonza, Basel, Switzerland). Media were supplemented with 10% Fetal Bovine Serum (FBS; Biochrom, Cambridge, UK) and 1% penicillin/streptomycin (Lonza). All cell cultures were kept at 5% CO₂ at 37° C. Following our internal Standard Operating Procedures (SOP), routine cell testing for mycoplasma was performed every four months. Cell authentication was conducted by IDEXX BioResearch once a year (Ludwigsburg, Germany).

Reagents

Recombinant human proteins WNT3a and RSPO2 were purchased from R&D systems (Wiesbaden, Germany) and PeproTech® (Hamburg, Germany), respectively. TGF β 1 and U0126 were from ImmunoTools (Friesoythe, Germany) and Cell Signaling Technology® (Frankfurt am Main, Germany), respectively. IWP2 was purchased from Selleck Chemicals (Munich, Germany).

GSEA

Gene set enrichment analysis (GSEA) of public datasets was performed with the GSEAPreRanked tool of the GSEA 4.1.0 Desktop Application and following the guidelines published by the Broad Institute (<http://www.broadinstitute.org/gsea/index.jsp>). Hallmark gene sets were collected from the Molecular Signature Database (MSigDB). Data derived from pancreatic cancer patients were selected from the TCGA database (Project ID: TCGA-PAAD) and normalized enrichment scores (NES) were calculated according to the ranked-ordered gene list. When NES>1 or <-1, it was considered as “enriched”, showing a positive or negative correlation of selected gene sets in PAAD LGR6corr patients.

Sphere-formation assay (SFA)

Adherent cells were trypsinized and subsequently resuspended as single cell suspensions. 1,000 cells were seeded in 100 μ l/well into low attachment 96-well plates (Corning, Krailling, Germany) in CSC medium. The FBS-depleted CSC medium was supplemented with

1XB27 (Thermo Fisher Scientific GmbH, Dreieich, Germany), 1% penicillin/streptomycin, recombinant human epidermal growth factor 10ng/ml and recombinant human fibroblast growth factor 20ng/ml (ImmunoTools, Friesoythe, Germany) in DMEM/F12 (Thermo Fisher Scientific GmbH, Dreieich, Germany). To prevent cell-cell aggregation, 1% methylcellulose (Sigma-Aldrich Chemie GmbH, Munich, Germany) was added to the medium. Medium changes occurred every 3-4 days. Spheres were counted after an incubation period of 12-15 days at 37° C.

Colony-formation assay (CFA)

After thorough counting, 1,000 cells were seeded into 6-well plates in regular complete culture medium as described above. The colonies were incubated at 37° C with 5% CO₂ for 7-10 days. Colonies were then fixed with 4% PFA and stained with crystal violet (Sigma GmbH, Rödermark, Germany). Cell clusters with approximately 50 cells or more were considered and counted as a colony.

siRNA transfection

For siRNA transfection, PDAC cells were seeded at 60% confluence into 24-well plates in RPMI1640 with 10% FBS only. After overnight incubation, the old medium was removed and new RPMI1640 without penicillin/streptomycin and FBS was added. According to the supplier's instructions, LGR6 siPOOL (siTOOLS Biotech, Martinsried, Germany) and Lipofectamine RNAiMAX Transfection Reagent (Thermo Fisher Scientific) were mixed at a ratio of 1:1 and added into the medium. The control group was prepared with negative control (nonsense) siRNA and Lipofectamine RNAiMAX Transfection Reagent. RNA was then isolated 48h after transfection. For SFA and CFA, cells were seeded 24h after transfection as described above.

Super TOP/FOP assay

Transient transfections for SuperTOP/FOP vector assays were carried out as detailed previously [32]. Normalization was carried out with Renilla luciferase vector pRLTK (Promega, Madison, WA, USA). Passive lysis buffer was added and firefly / Renilla luciferase activity was determined with the Dual Luciferase Reporter Assay System (Promega) in triplicates.

RNA extraction, reverse transcription, and qPCR

Total RNA from cultured cells was extracted using the total RNA isolation kit peqGOLD (VWR™/Avantor™) according to the manufacturer's instructions. Concentration and purity of extracted RNAs were verified by a

NanoDrop™ 2000 spectrophotometer (Thermo Fisher Scientific). cDNA transcription was accomplished using the cDNA synthesis kit (Bio-Rad Laboratories GmbH, Feldkirchen, Germany). qPCR was performed with the Sso Fast™ EvaGreen® kit (Bio-Rad); for normalization of gene of interest expression, we used the house-keeping gene GAPDH.

Western blot

Treated cells were lysed in RIPA buffer including complete protease and phosphatase inhibitor cocktails (Roche). Gel electrophoresis of cell lysates was carried out on 12% SDS-PAGE gels and later transferred onto PVDF membranes. Membranes were incubated with primary antibodies overnight at 4° C and HRP-conjugated secondary antibodies for 1h at room temperature. Exposure was performed using ECL™ Western Blotting Detection Reagent (GE Healthcare, Munich, Germany). All antibodies are listed in Supplementary Table 1.

Immunofluorescence staining

Cells were seeded onto coverslips, fixed with 4% PFA for 20 mins, and followed by blocking steps with endogenous peroxidase and permeabilization with 0.1% Triton X-100/PBS. After blocking with 5% goat serum, the slides were incubated with primary antibodies at 4° C overnight, followed by secondary antibody incubation at room temperature for 1h in the dark. Slides were counterstained with DAPI (labeling and detection) and then covered with Fluorescence Mounting Medium (VECTASHIELD® Antifade Mounting Medium H-1000, Vector Labs, Eching, Germany). All primary antibodies used here were itemized in Supplementary Table 1.

Flow cytometry

After culturing in regular medium for 24h, cells were trypsinized, washed, and collected. For analysis of membrane-bound LGR6, cells were incubated with human LGR6 APC-conjugated antibody (10µL/10⁶cells) (R&D Systems®, FAB8458A-025) for 30 mins at RT. In contrast, cytoplasmic LGR6 detection included fixation of the cells with IC fixation buffer and washing steps with permeabilization buffer (both from eBioscience, Waltham, MA, USA). Then, cells were incubated with anti-LGR6 antibody solution for 30 mins at RT. After washing, cells were analyzed by a BD LSRFortessa™ cell analyzer (BD Biosciences, Franklin lakes, NJ, USA). Negative controls were prepared likewise.

Statistical analysis

Statistical analysis was performed by GraphPad Prism 9 (GraphPad Software, San Diego, USA). Data are

represented as mean $\pm$ SEM. Significance levels were calculated by t-test. $P < 0.05$ was considered as statistically significant.

RESULTS

LGR6 expression correlates with epithelial phenotype in PDAC cell lines

PDAC cell lines were categorized into epithelial and mesenchymal phenotypes according to their morphology (Supplementary Figure 1) and expression patterns of the epithelial marker E-cadherin [33] or mesenchymal transcription factor ZEB1 [34].

According to this, BxPC3 and Capan2 phenotypically categorized as epithelial due to their cobblestone-like cell-cell connections and higher expression of *CDH1* (E-cadherin). In contrast, MiaPaCa2 and Panc1 showed loose cell-cell connectivity with spindle-shaped morphology, lower expression of *CDH1* (E-cadherin), high expression of *ZEB1* and high sphere-formation potential (SFA) classifying them as mesenchymal (Figure 1A, left). To understand LGR6 expression patterns in the above-mentioned categorized cell lines, *LGR6* gene expression was determined and sided with the epithelial markers (Figure 1A). As shown before, phenotypical differences were also detectable in 3D with epithelial spheres appearing very compact and

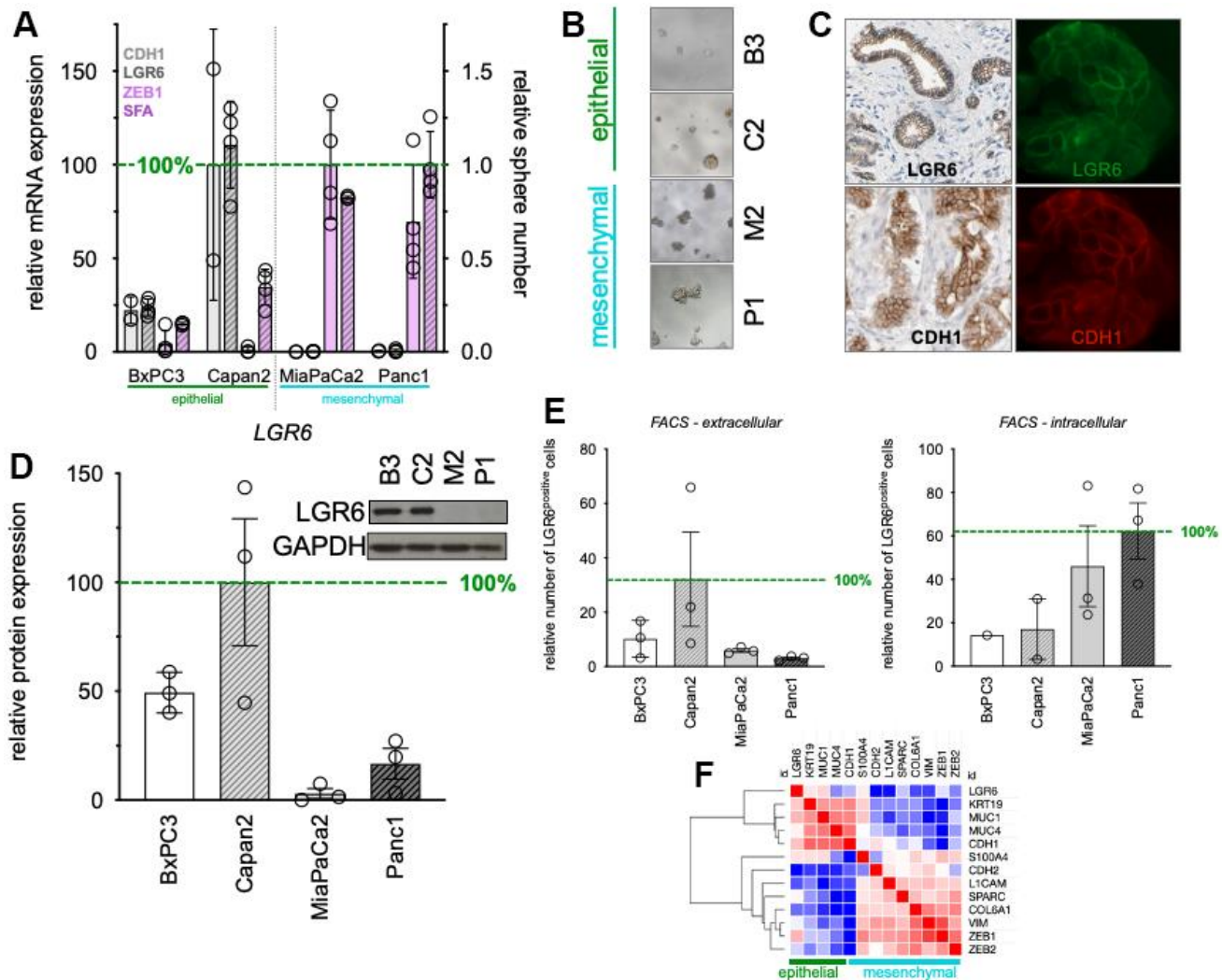


Figure 1. Baseline expression of LGR6 in PDAC cell lines. (A) qPCR of epithelial (*CDH1*), mesenchymal (*ZEB1*) markers, and *LGR6*. In addition, sphere-formation capacity (SFA) was evaluated. (B) Morphology of defined epithelial and mesenchymal PDAC cell lines; (C) Immunohistochemistry (left panels, Protein atlas) and immunofluorescence staining LGR6 (green) and E-cadherin (red) in Capan2; (D) Western blot analysis of LGR6 in pancreatic cancer cell lines. Quantification is shown as normalized to Capan2; (E) Flow cytometric analysis of extra- and intra-cellular LGR6 in pancreatic cancer cell lines; (F) Clustering analysis of the TCGA databank on PDAC identified *LGR6* with epithelial marker group *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$; ****, $P < 0.0001$.

mesenchymal spheres rather grape-like (Figure 1B) [32]. Immuno-fluorescence staining detected strong LGR6 signals on the cell membrane in epithelial cells (Figure 1C right panels) suggesting colocalization with E-Cadherin (CDH1) which is in line with immunohistochemical stains of public databases (Figure 1C left panels). Western blotting for LGR6 protein corroborated these findings and demonstrated up to 33-fold stronger expression (Capan2 vs. MiaPaCa2) in epithelial cells (Figure 1D).

In line with this, flowcytometric intra- (Figure 1E, left) and extra-cellular (Figure 1E, right) LGR6 expression patterns were analyzed. Epithelial subsets showed markedly higher membrane bound LGR6 (up to 10-fold, Capan2 vs. Panc1) and lower cytoplasmic LGR6 expression compared to the mesenchymal cell lines, where proportions were reversed (Figure 1E).

Last, database analysis of the TCGA databank on PDAC revealed strong hierarchical clustering of *LGR6* with epithelial markers *KRT19*, *MUC1*, *MUC4*, and

CDH1, whereas mesenchymal markers robustly clustered separately (Figure 1F). GSEA analysis of typical EMT signatures corroborated these data (Supplementary Figure 2). Moreover, overlap analysis of TCGA and CCLE data, where we compared LGR6 high to LGR6 low expression, revealed 30 genes in common of both databases (Supplementary Figure 3). Several of those genes were related to cell-cell adhesion, such as cytokeratins or cell adhesion molecules (Supplementary Table 2).

Heterogeneous WNT activity associates with LGR6 expression and epithelial signatures in PDAC

As we could show before, WNT activity is very heterogenous in between different cell types of PDAC as well as different cells of the same cell line [32]. In this work, we show in Super TOP/FOP assays, that BxPC3 and Capan2 (epithelial category) tended to contain higher baseline WNT activity levels (Figure 2A). Clustering of mRNA results revealed an affiliation of *LGR6* with WNT target genes *AXIN2*

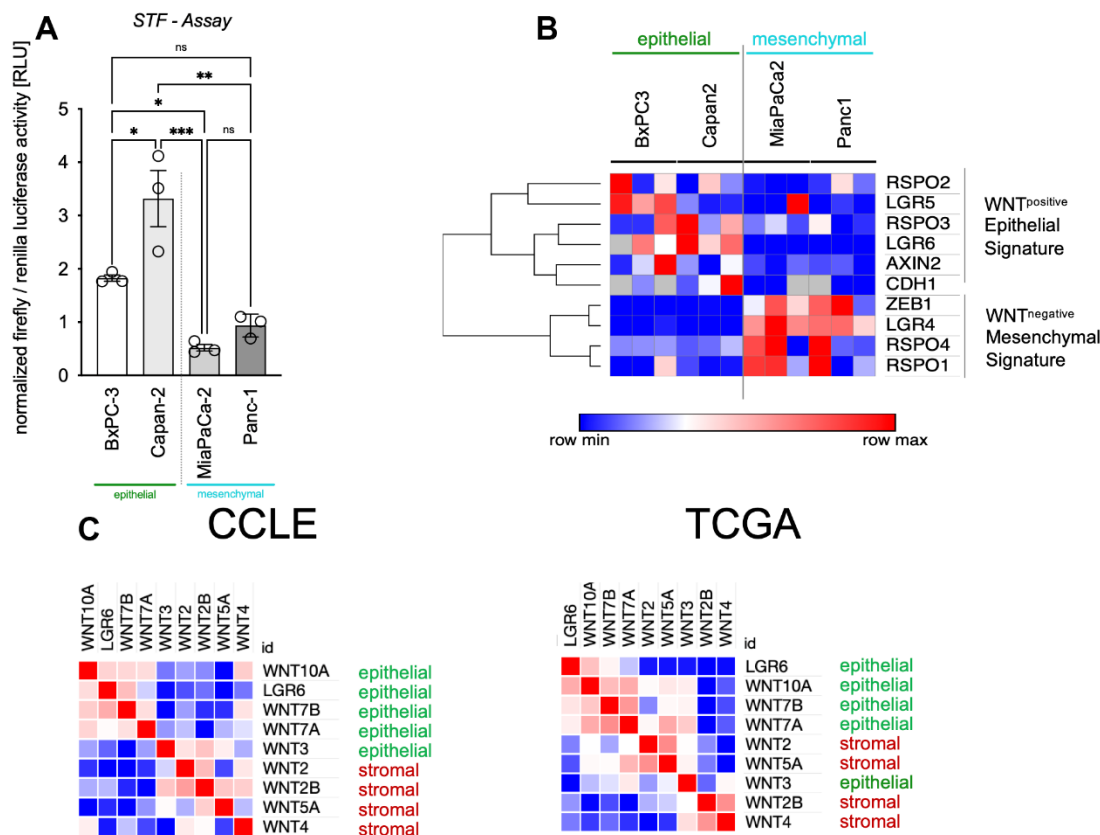


Figure 2. LGR6 expression correlates with epithelial signatures. (A) Super TOP/FOP assays (STF assay) revealed higher baseline WNT activity levels in epithelial (green) vs. mesenchymal (blue) pancreatic cancer cells. (B) Clustering of gene expression in the same cell lines affiliates *LGR6* with WNT^{positive} epithelial signature, whereas the WNT^{negative} mesenchymal signature was highly expressed in mesenchymal PDAC cell lines (blue). (C) Epithelial WNT niche subtypes described by Seino and colleagues clustered with *LGR6* in CCLE and TCGA datasets. *, P<0.05; **, P<0.01; ***, P<0.001.

and *LGR5* as well as *RSPO2*, *RSPO3*, and *CDH1* what we describe as WNT^{positive} epithelial signature. In contrast, *RSPO1*, *RSPO4*, *LGR4*, and *ZEB1* were highly expressed in mesenchymal cells as part of the WNT^{negative} mesenchymal signature (Figure 2B). According with this, we analyzed WNT niche subtypes described by Seino and colleagues in CCLE and TCGA datasets. Here, we also found that epithelial WNTs (3, 7A and B, and 10A) clustered strongly with *LGR6*. This might give hints towards a robust role of *LGR6* creating WNT independence in those cell lines (Figure 2C) [35].

Exogenous canonical WNT activation mediates *LGR6* expression

As described before, exogenous WNT3a and *RSPO2* enhance WNT signaling in highly responsive cells, whereas *RSPO2* alone seems insufficient in most cell lines [32]. Immunofluorescent assays confirmed this notion and showed beta-Catenin translocation into the nucleus after co-stimulation in Panc1 (Figure 3A). Western blotting for *LGR6* demonstrated upregulation upon exogenous WNT stimulation with stronger effects with both *RSPO2* and WNT3a (Figure 3B).

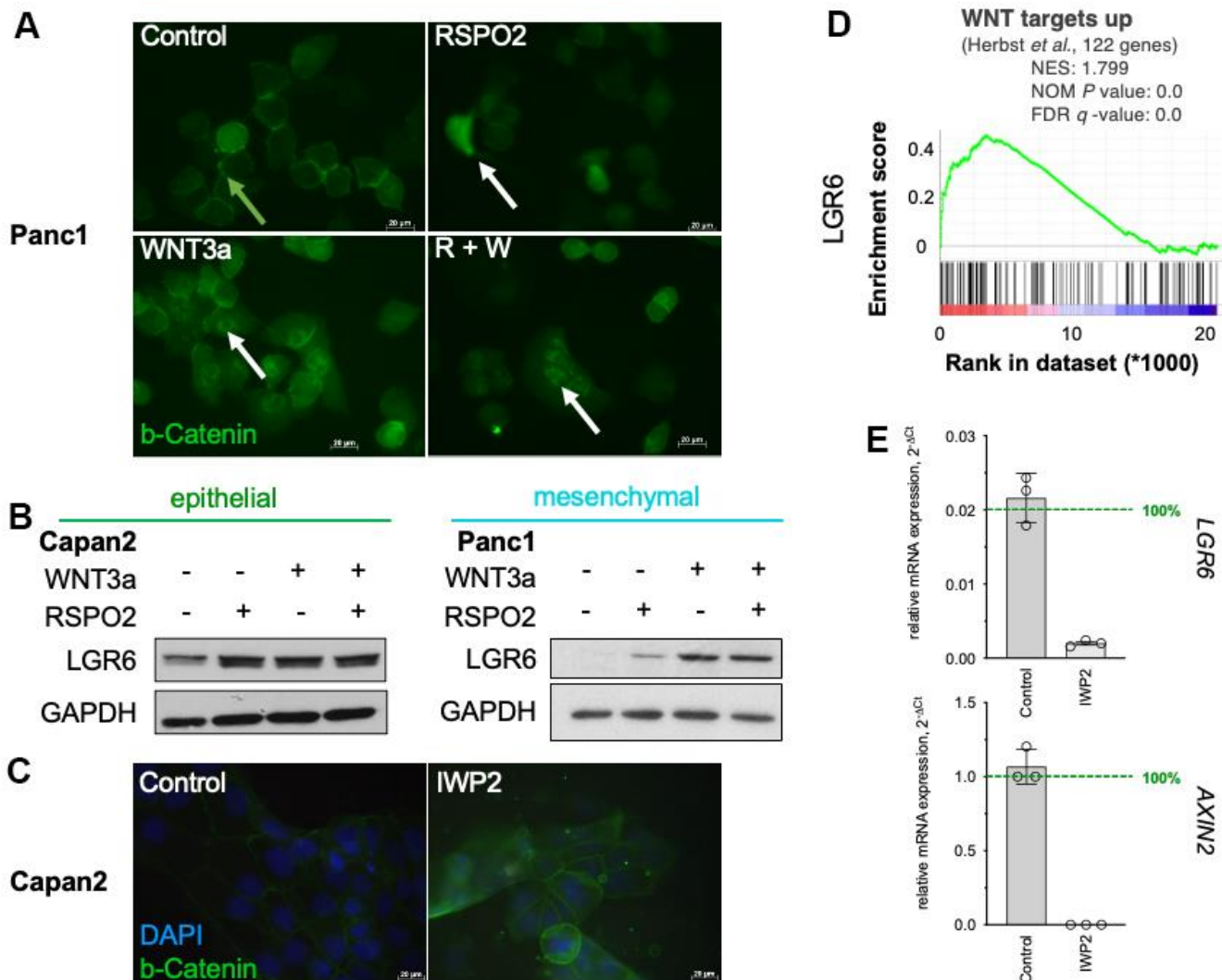


Figure 3. *LGR6* as a WNT target gene. (A) Immunofluorescence staining of β -catenin (green) in Panc1 after stimulation with *RSPO2*, WNT3a or its combination (R+W) showed more nuclear staining upon WNT activation, in particular with WNT3a or R+W. White arrows indicate nuclear β -catenin, green arrows indicate membrane-bound β -catenin. (B) Western blot of *LGR6* after similar stimulation showed increase in both Capan2 and Panc1. (C) GSEA of TCGA data revealed positive correlation of *LGR6* with the WNT signature “WNT targets up” by Herbst et al. (D) Immunofluorescence staining of β -catenin in Capan2 reveals increase in membrane-bound and decrease of nuclear β -catenin upon IWP2 inhibition. The cells were cultured with 10uM IWP2 for 72h, medium was changed every other day. Bar, 20 μ m; (E) Effect of IWP2 on *AXIN2* and *LGR6* mRNA expression in Capan2. Significant decreases were detected in presence of IWP2. ***, $P < 0.001$; ****, $P < 0.0001$.

Gene Set Enrichment Analysis (GSEA) also revealed a strong positive correlation of WNT targets and LGR6 in PDAC supporting our hypothesis that *LGR6* might be a novel WNT target gene in PDAC (Figure 3C). To further investigate the regulatory effects of WNT signaling on LGR6 expression, we reduced WNT signaling activity using the porcupine inhibitor IWP2. IWP-2 selectively targets porcupine, a membrane-bound acyltransferase (MBOAT), which is essential to produce WNT proteins. Administration of IWP-2 leads to inhibition of LRP6 and Dvl2 phosphorylation and decreased beta-Catenin accumulation [32]. These effects are transient and reversible [36].

As shown in Figure 3D, we detected decreased β -catenin signal upon IWP2 inhibition in Capan2. Moreover, down-regulation of the WNT target gene *AXIN2* in Capan2 corroborated this finding on RNA level (Figure 3E). A reduced *LGR6* expression was detected upon the inhibition of WNT activity (Figure 3E). These data suggest that downregulation of WNT signaling might be followed by a reduction of LGR6 expression.

LGR6 knock-down reduces cancer stemness and responsiveness to exogenous WNT stimulation in PDAC

To fully explore functional roles of LGR6 in cancer cell stemness, LGR6 was knocked down by siRNA transfection. Efficient knock down resulted in a 90% decrease in *LGR6* mRNA level in comparison to the vector control (Supplementary Figure 4A). Next, we evaluated the effect of LGR6 silencing in PDAC cell lines on their colony formation ability. As shown in Supplementary Figure 4B, colonies were markedly smaller in the siLGR6-transfected group (siLGR6) compared to the vector control (VC). Consistent with this result, SFAs revealed that siLGR6 led to a decrease in spheroid size and number compared with VC (Supplementary Figure 4C). To investigate the role of LGR6 in exogenous mediation of WNT, we also evaluated SFA, *LGR6*, and WNT target *AXIN2* expression after stimulation with RSPO2 and WNT3a after VC (Figure 4A) or LGR6-KD (Figure 4B) in both Capan2 and Panc1. As expected, SFA,

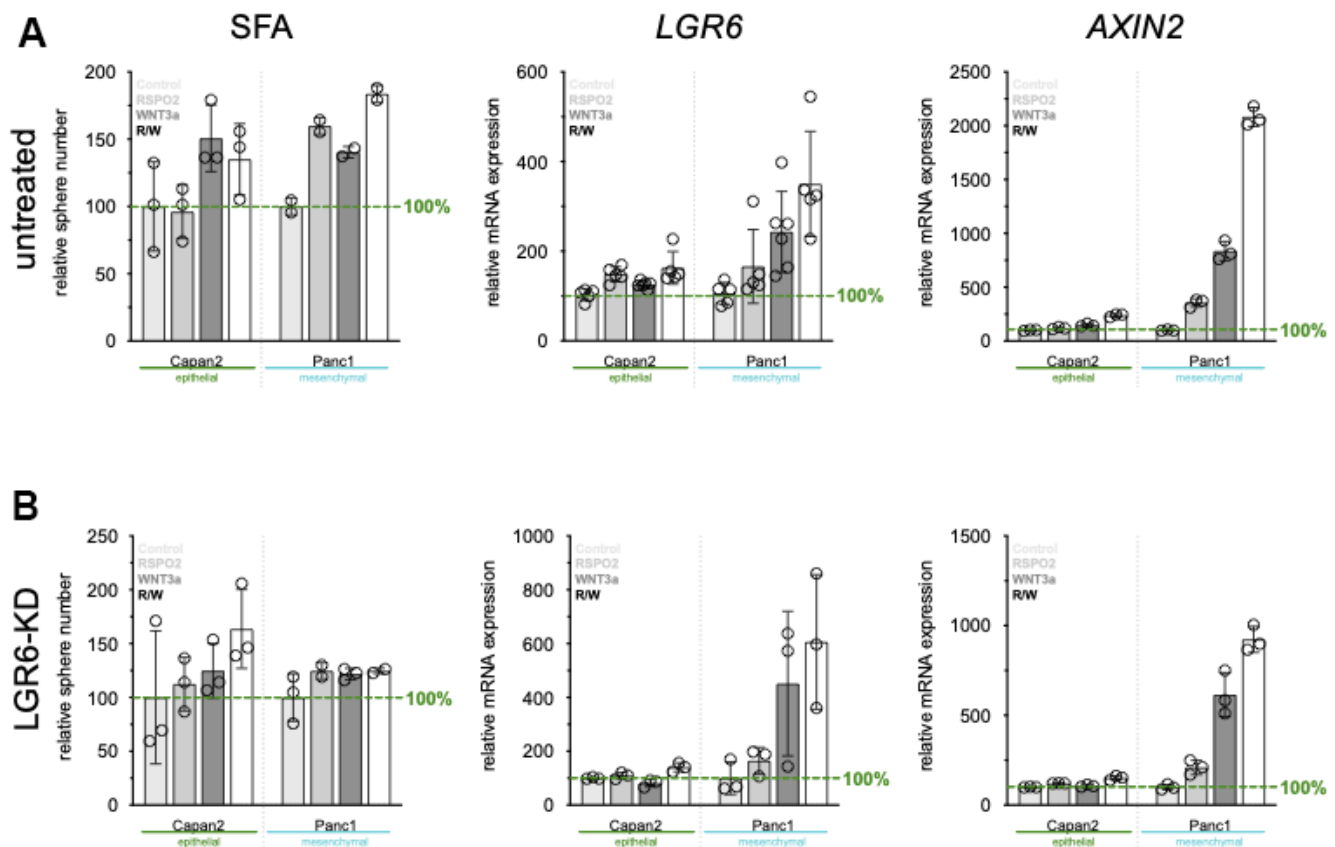


Figure 4. LGR6 depletion correlates with reduced cancer stemness. (A) sphere formation ability (SFA) and qPCR of *LGR6* and WNT target gene *AXIN2* in vector control treated cells. Stimulation with RSPO2, WNT3a or its combination leads to increased SFA, and *LGR6*/*AXIN2*. Knock-down of *LGR6* (B) reduces SFA and influences *LGR6* and *AXIN2* gene expression, suggesting partial regulation of canonical WNT through LGR6.

LGR6, and the WNT target gene *AXIN2* significantly increased upon RSPO2 and/or WNT3a stimulation, indicating a successful WNT axis activation (Figure 4A). *LGR6*-KD impeded sphere-formation even after exogenous WNT stimulation, especially in the more WNT-sensitive cell line Panc1 (Figure 4B). *LGR6* and *AXIN2* expression were lower, also after exogenous WNT stimulation; however, both were not completely inhibited suggesting an important, but not all-encompassing role of *LGR6* in regulating the canonical WNT axis in PDAC (Figure 4B).

***LGR6* mRNA expression is associated with malignancy in PDAC and a trend towards worse survival**

Finally, we tested for clinical correlations of *LGR6* mRNA levels in different publicly available data sets with a total of 299 PDAC and 75 normal pancreas (NP) cases. *LGR6* expression was significantly higher in PDAC as compared to NP in GSE62165 and GSE71729 with a similar trend in GSE16515 (Figure 5A). Using ROC curve analyses and Youden's index, we identified

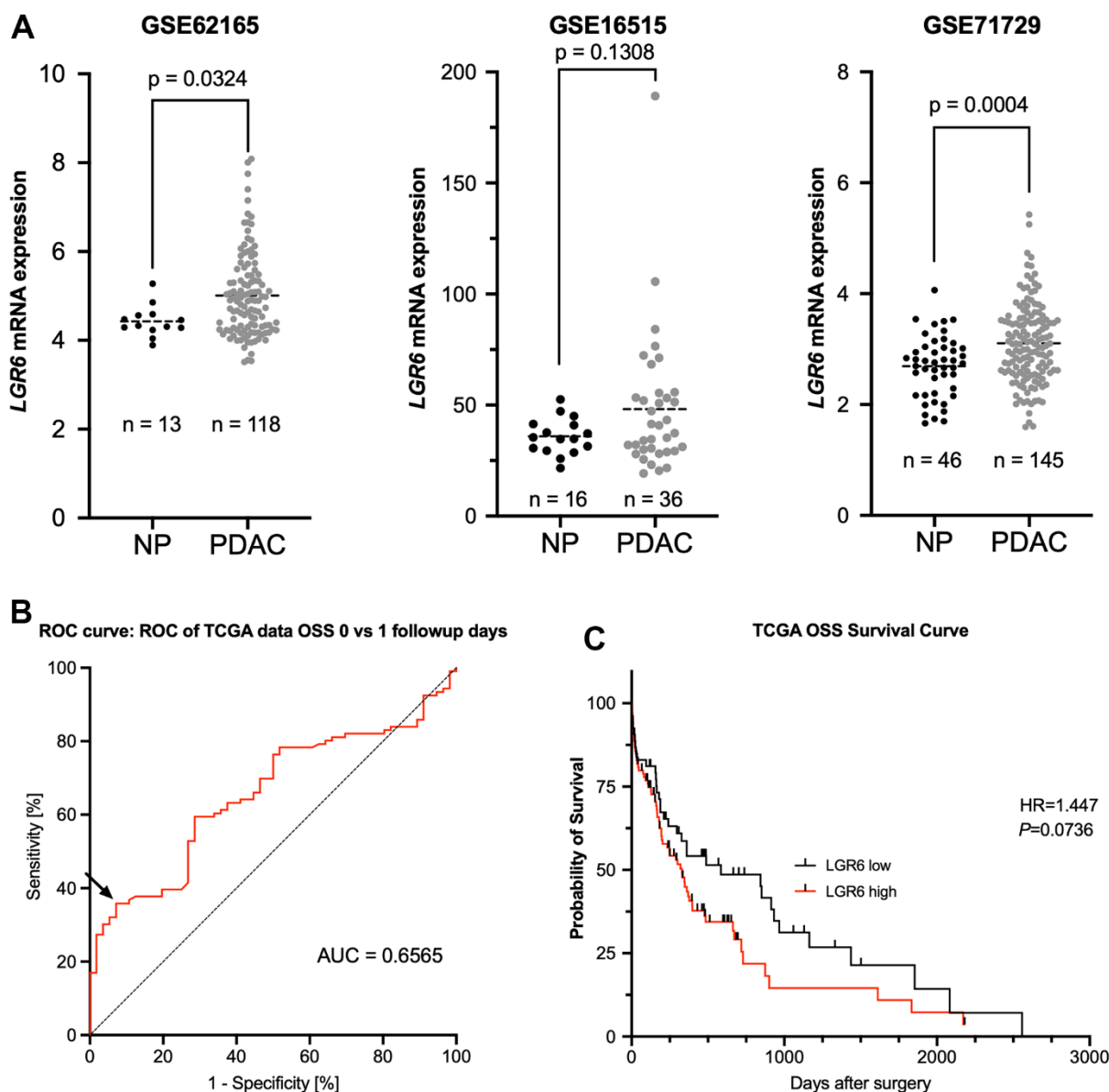


Figure 5. *LGR6* is higher expressed in PDAC compared to normal tissue and trends towards worse overall survival. (A) Analysis of publicly available datasets (GSE62165, GSE16515, and GSE71729) shows significantly more *LGR6* in tumor vs. normal pancreatic tissue (NP). (B) ROC curve analysis of normalized *LGR6* mRNA expression for best discrimination threshold with an area under the curve (AUC) of 0.6565. (C) Dichotomal classification revealed a trend of high *LGR6* expression and poor overall survival; survival of PDAC patients of *LGR6*^{high} versus *LGR6*^{low} is decreased without statistical significance.

an ideal cutoff at a normalized expression intensity of 0.6565 (natural scale) of *LGR6* mRNA (Figure 5B). Dichotomal classification revealed a trend of high *LGR6* expression and poor overall survival (Figure 5C). Expressed in numbers, median survival of the *LGR6* high subgroup was 329 days, whereas *LGR6* low led to an increased survival with a median of 584 days. However, the remaining clinical data of the TCGA databank revealed no statistically significant difference (Supplementary Table 3).

DISCUSSION

The canonical WNT pathway is reportedly an essential protagonist in organ development as well as oncogenesis in multiple cancers. *LGR5* was initially identified as a WNT target gene in human colon cancer and especially in those that harbored WNT-activating mutations [37]. Moreover, it has been shown to act as a receptor of RSPO and as such, *LGR5* enhances the canonical WNT pathway [29, 31]. The RSPOs are a family of secreted factors that augment already activated WNT signaling through binding to *LGR4-6*. Our previous study indicated that RSPO2 could enhance WNT activity of certain PDAC cells and endow highly responsive subgroups with CSCs characteristics [32]. As a close homolog of *LGR5*, we hypothesized that *LGR6* might be an essential requirement in PDAC for reasons other than its assumed role as a receptor of RSPOs and enhancer of WNT signaling. In the present study, we were able to demonstrate that *LGR6* expression was altered with differential WNT activity. This led to the idea that the *LGR6* gene itself may be a direct target of the WNT signaling pathway in PDAC cells as we were able to show that activated WNT led to enhanced *LGR6* expression; vice versa, WNT inhibition was associated with decreased expression patterns of *LGR6*. *LGR6* regulation in PDAC may therefore constitute a crucial role within a positive feedback loop of WNT signaling activity.

Previous research showed that overexpression or knockout of *LGR5* resulted in pronounced changes of the cytoskeleton and cell adhesion complexes in some cancer cell lines lacking endogenous or exogenous RSPO stimulation [38, 39]. *LGR5* silenced colorectal cancer cells tended to be more mesenchymal, while overexpression of *LGR5* was linked to a more epithelial phenotype [38]. *LGR5* overexpression in hepatocellular carcinoma cells resulted in changes from a mesenchymal phenotype to a more aggregated phenotype typical for the epithelial subtype. Knockdown of *LGR5* shifted cells from an aggregated phenotype to a spindle-shaped one [39]. On the other hand, overexpression of *LGR6* in HeLa cells increased cell movement after treatment with RSPO1 and WNT3a. Moreover, overactivation of WNT signaling correlates with

increased cell migration [40]. To investigate these – in part - conflicting findings, we submerged into more detail in the present study. We used PDAC cell lines of different morphological appearances to study the association of *LGR6* in EMT subgroups of PDAC. In this regard, Capan2 and BxPC3 represent an epithelial phenotype with intensive cell-cell contacts, while MiaPaCa2 and Panc1 exemplify the mesenchymal phenotype with spindle-shaped morphology and lose cell-cell contacts [32, 41]. Using multiple approaches, we found that *LGR6* is preferentially expressed in more epithelial cell lines, whereas mesenchymal cell lines harbored much lower *LGR6* expression levels. Furthermore, expression pattern analysis of *LGR6* showed that its distribution was mostly located on the cell membrane in epithelial cell lines, while less or no expression was detected in mesenchymal cell lines in this position.

Cancer stem cells (CSCs) are believed to initiate and maintain malignancies of different types and engage in chemo-resistance mechanisms as well as metastatic activities. *LGR5* was identified as an adult stem cell marker in various tissues, including intestine, liver, skin, stomach, and ovarian epithelia [42, 43]. Both *LGR5*⁺ and *LGR6*⁺ stem cell compartments contribute to epidermal repair in response to acute wounds [44, 45]. In the present study, we investigated stemness PDAC cell lines by typical *in vitro* assays, such as colony formation capacity in 2D or sphere formation capacity in 3D. The deletion of *LGR6* negatively impacted on the growth of colonies and spheres, indicating that *LGR6* might have a potential role in maintaining PDAC stemness. In part, this could also be explained by reduced canonical WNT activity after *LGR6* knock down.

There are several limitations to the present study. First, we did not generate any *LGR6*-overexpressing PDAC cell lines for further functional analysis. Second, comparison of *LGR6*⁺ with *LGR6*⁻ cells after flowcytometric cell sorting might be another elegant way to explore the roles of *LGR6* in WNT signaling, EMT, and cancer stemness. Third, mouse models have not been implemented to translate our *in vitro* results into *in vivo* conditions where essential mechanisms, such as metastasis or chemo-resistance could be further evaluated. In a clinical translational approach, we believe that correlating gene or protein expression with different genomic as well as metabolic subtypes [46–48] could further enlighten the role of *LGR6*. Stratification of PDAC after primary resection into *LGR6*^{high} versus *LGR6*^{low} tumors and/or organoids could further help to decide whether enforced chemotherapeutic regimens such as FOLFIRINOX should be applied.

Taken together, we present new evidence in PDAC that LGR6 might be a novel WNT target gene in this tumor. LGR6 seems to be involved in EMT and cancer stemness. This knowledge could be applicable for detection and treatment of special subsets of pancreatic cancer cells. Further research is still needed to dissect the exact mechanisms under physiological as well as pathological conditions of benign and cancerous pancreatic cells.

Abbreviations

DFS: Disease-free survival; OS: Overall survival; PDAC: Pancreatic ductal adenocarcinoma; RSPO: R-Spondin; PKA: Protein kinase A; TCF/LEF binding sites: T-cell factor/lymphoid enhancer factor; LGR4/5/6: Leucine-rich repeat containing G-protein coupled receptors 4/5/6; EMT: Epithelial–mesenchymal transition; GSEA: Gene set enrichment analysis; MsigDB: Molecular Signature Database; ES: Enrichment scores; SFA: sphere-formation assay; CSCs: Cancer stem cells; WB: Western blot.

AUTHOR CONTRIBUTIONS

Jing Wang and Dominik T. Koch conceived the study, designed and carried out experiments, analyzed and interpreted data, generated the figures and tables and wrote the manuscript; Felix O. Hofmann, Daniel Härtwig, Iris Beirith, and Klaus Peter Janssen carried out experiments, analyzed and interpreted data; Alexandr V. Bazhin and Hanno Niess, performed statistical data analysis and interpreted data; Jens Werner, Bernhard W. Renz and Matthias Ilmer analyzed and interpreted data. BWR and MI designed and supervised the study and MI provided funding. All authors reviewed, edited, and approved the final manuscript.

ACKNOWLEDGMENTS

We thank Karin Enderle and Sevdije Issar Amerchel for their excellent technical assistance.

CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

FUNDING

Research of M.I. received funding through Else Kröner-Fresenius Stiftung (2019_A130), Wilhelm Sander-Stiftung (2019.022.1), Bayerisches Zentrum für Krebsforschung (BZKF) as well as intramural support through the Förderprogramm für Forschung und Lehre (FöFeLe) of the LMU.

REFERENCES

1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015; 136:E359–86.
<https://doi.org/10.1002/ijc.29210>
PMID:25220842
2. Hidalgo M, Cascinu S, Kleeff J, Labianca R, Löhner JM, Neoptolemos J, Real FX, Van Laethem JL, Heinemann V. Addressing the challenges of pancreatic cancer: future directions for improving outcomes. *Pancreatology*. 2015; 15:8–18.
<https://doi.org/10.1016/j.pan.2014.10.001>
PMID:25547205
3. Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrisian LM. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. *Cancer Res*. 2014; 74:2913–21.
<https://doi.org/10.1158/0008-5472.CAN-14-0155>
PMID:24840647
4. Cadigan KM, Nusse R. Wnt signaling: a common theme in animal development. *Genes Dev*. 1997; 11:3286–305.
<https://doi.org/10.1101/gad.11.24.3286>
PMID:9407023
5. Haegel H, Larue L, Ohsugi M, Fedorov L, Herrenknecht K, Kemler R. Lack of beta-catenin affects mouse development at gastrulation. *Development*. 1995; 121:3529–37.
<https://doi.org/10.1242/dev.121.11.3529>
PMID:8582267
6. Wells JM, Esni F, Boivin GP, Aronow BJ, Stuart W, Combs C, Sklenka A, Leach SD, Lowy AM. Wnt/beta-catenin signaling is required for development of the exocrine pancreas. *BMC Dev Biol*. 2007; 7:4.
<https://doi.org/10.1186/1471-213X-7-4>
PMID:17222338
7. Papadopoulou S, Edlund H. Attenuated Wnt signaling perturbs pancreatic growth but not pancreatic function. *Diabetes*. 2005; 54:2844–51.
<https://doi.org/10.2337/diabetes.54.10.2844>
PMID:16186384
8. de La Coste A, Romagnolo B, Billuart P, Renard CA, Buendia MA, Soubrane O, Fabre M, Chelly J, Beldjord C, Kahn A, Perret C. Somatic mutations of the beta-catenin gene are frequent in mouse and human hepatocellular carcinomas. *Proc Natl Acad Sci USA*. 1998; 95:8847–51.
<https://doi.org/10.1073/pnas.95.15.8847>
PMID:9671767

9. Hugh TJ, Dillon SA, O'Dowd G, Getty B, Pignatelli M, Poston GJ, Kinsella AR. beta-catenin expression in primary and metastatic colorectal carcinoma. *Int J Cancer*. 1999; 82:504–11.
[https://doi.org/10.1002/\(sici\)1097-0215\(19990812\)82:4<504::aid-ijc6>3.0.co;2-6](https://doi.org/10.1002/(sici)1097-0215(19990812)82:4<504::aid-ijc6>3.0.co;2-6) PMID:10404062
10. Turashvili G, Bouchal J, Burkadze G, Kolar Z. Wnt signaling pathway in mammary gland development and carcinogenesis. *Pathobiology*. 2006; 73:213–23.
<https://doi.org/10.1159/000098207> PMID:17314492
11. Rimm DL, Caca K, Hu G, Harrison FB, Fearon ER. Frequent nuclear/cytoplasmic localization of beta-catenin without exon 3 mutations in malignant melanoma. *Am J Pathol*. 1999; 154:325–9.
[https://doi.org/10.1016/s0002-9440\(10\)65278-9](https://doi.org/10.1016/s0002-9440(10)65278-9) PMID:10027390
12. Abraham SC, Klimstra DS, Wilentz RE, Yeo CJ, Conlon K, Brennan M, Cameron JL, Wu TT, Hruban RH. Solid-pseudopapillary tumors of the pancreas are genetically distinct from pancreatic ductal adenocarcinomas and almost always harbor beta-catenin mutations. *Am J Pathol*. 2002; 160:1361–9.
[https://doi.org/10.1016/s0002-9440\(10\)62563-1](https://doi.org/10.1016/s0002-9440(10)62563-1) PMID:11943721
13. Gerdes B, Ramaswamy A, Simon B, Pietsch T, Bastian D, Kersting M, Moll R, Bartsch D. Analysis of beta-catenin gene mutations in pancreatic tumors. *Digestion*. 1999; 60:544–8.
<https://doi.org/10.1159/000007704> PMID:10545724
14. Zeng G, Germinaro M, Micsenyi A, Monga NK, Bell A, Sood A, Malhotra V, Sood N, Midda V, Monga DK, Kokkinakis DM, Monga SP. Aberrant Wnt/beta-catenin signaling in pancreatic adenocarcinoma. *Neoplasia*. 2006; 8:279–89.
<https://doi.org/10.1593/neo.05607> PMID:16756720
15. Pasca di Magliano M, Biankin AV, Heiser PW, Cano DA, Gutierrez PJ, Deramautd T, Segara D, Dawson AC, Kench JG, Henshall SM, Sutherland RL, Dlugosz A, Rustgi AK, Hebrok M. Common activation of canonical Wnt signaling in pancreatic adenocarcinoma. *PLoS One*. 2007; 2:e1155.
<https://doi.org/10.1371/journal.pone.0001155> PMID:17982507
16. Clevers H, Nusse R. Wnt/ β -catenin signaling and disease. *Cell*. 2012; 149:1192–205.
<https://doi.org/10.1016/j.cell.2012.05.012> PMID:22682243
17. Taurin S, Sandbo N, Qin Y, Browning D, Dulin NO. Phosphorylation of beta-catenin by cyclic AMP-dependent protein kinase. *J Biol Chem*. 2006; 281:9971–6.
<https://doi.org/10.1074/jbc.M508778200> PMID:16476742
18. Hao HX, Xie Y, Zhang Y, Charlat O, Oster E, Avello M, Lei H, Mickanin C, Liu D, Ruffner H, Mao X, Ma Q, Zamponi R, et al. ZNRF3 promotes Wnt receptor turnover in an R-spondin-sensitive manner. *Nature*. 2012; 485:195–200.
<https://doi.org/10.1038/nature11019> PMID:22575959
19. Koo BK, Spit M, Jordens I, Low TY, Stange DE, van de Wetering M, van Es JH, Mohammed S, Heck AJ, Maurice MM, Clevers H. Tumour suppressor RNF43 is a stem-cell E3 ligase that induces endocytosis of Wnt receptors. *Nature*. 2012; 488:665–9.
<https://doi.org/10.1038/nature11308> PMID:22895187
20. Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. *Cell*. 2009; 139:871–90.
<https://doi.org/10.1016/j.cell.2009.11.007> PMID:19945376
21. Thiery JP, Sleeman JP. Complex networks orchestrate epithelial-mesenchymal transitions. *Nat Rev Mol Cell Biol*. 2006; 7:131–42.
<https://doi.org/10.1038/nrm1835> PMID:16493418
22. Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *J Clin Invest*. 2009; 119:1420–8.
<https://doi.org/10.1172/JCI39104> PMID:19487818
23. Perez-Moreno M, Jamora C, Fuchs E. Sticky business: orchestrating cellular signals at adherens junctions. *Cell*. 2003; 112:535–48.
[https://doi.org/10.1016/s0092-8674\(03\)00108-9](https://doi.org/10.1016/s0092-8674(03)00108-9) PMID:12600316
24. Perez-Moreno M, Fuchs E. Catenins: keeping cells from getting their signals crossed. *Dev Cell*. 2006; 11:601–12.
<https://doi.org/10.1016/j.devcel.2006.10.010> PMID:17084354
25. Gottardi CJ, Wong E, Gumbiner BM. E-cadherin suppresses cellular transformation by inhibiting beta-catenin signaling in an adhesion-independent manner. *J Cell Biol*. 2001; 153:1049–60.
<https://doi.org/10.1083/jcb.153.5.1049> PMID:11381089
26. Kuphal F, Behrens J. E-cadherin modulates Wnt-dependent transcription in colorectal cancer cells but does not alter Wnt-independent gene expression in fibroblasts. *Exp Cell Res*. 2006; 312:457–67.
<https://doi.org/10.1016/j.yexcr.2005.11.007> PMID:16368435
27. Onder TT, Gupta PB, Mani SA, Yang J, Lander ES, Weinberg RA. Loss of E-cadherin promotes metastasis via multiple downstream transcriptional pathways. *Cancer Res*. 2008; 68:3645–54.

<https://doi.org/10.1158/0008-5472.CAN-07-2938>
PMID:18483246

28. Ghahhari NM, Babashah S. Interplay between microRNAs and WNT/ β -catenin signalling pathway regulates epithelial-mesenchymal transition in cancer. *Eur J Cancer*. 2015; 51:1638–49.
<https://doi.org/10.1016/j.ejca.2015.04.021>
PMID:26025765
29. de Lau W, Barker N, Low TY, Koo BK, Li VS, Teunissen H, Kujala P, Haegebarth A, Peters PJ, van de Wetering M, Stange DE, van Es JE, Guardavaccaro D, et al. Lgr5 homologues associate with Wnt receptors and mediate R-spondin signalling. *Nature*. 2011; 476:293–7.
<https://doi.org/10.1038/nature10337> PMID:21727895
30. Carmon KS, Lin Q, Gong X, Thomas A, Liu Q. LGR5 interacts and cointernalizes with Wnt receptors to modulate Wnt/ β -catenin signaling. *Mol Cell Biol*. 2012; 32:2054–64.
<https://doi.org/10.1128/MCB.00272-12>
PMID:22473993
31. Barker N, van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, Haegebarth A, Korving J, Begthel H, Peters PJ, Clevers H. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature*. 2007; 449:1003–7.
<https://doi.org/10.1038/nature06196>
PMID:17934449
32. Ilmer M, Boiles AR, Regel I, Yokoi K, Michalski CW, Wistuba II, Rodriguez J, Alt E, Vykoukal J. RSP02 Enhances Canonical Wnt Signaling to Confer Stemness-Associated Traits to Susceptible Pancreatic Cancer Cells. *Cancer Res*. 2015; 75:1883–96.
<https://doi.org/10.1158/0008-5472.CAN-14-1327>
PMID:25769727
33. Arumugam T, Ramachandran V, Fournier KF, Wang H, Marquis L, Abbruzzese JL, Gallick GE, Logsdon CD, McConkey DJ, Choi W. Epithelial to mesenchymal transition contributes to drug resistance in pancreatic cancer. *Cancer Res*. 2009; 69:5820–8.
<https://doi.org/10.1158/0008-5472.CAN-08-2819>
PMID:19584296
34. Wellner U, Schubert J, Burk UC, Schmalhofer O, Zhu F, Sonntag A, Waldvogel B, Vannier C, Darling D, zur Hausen A, Brunton VG, Morton J, Sansom O, et al. The EMT-activator ZEB1 promotes tumorigenicity by repressing stemness-inhibiting microRNAs. *Nat Cell Biol*. 2009; 11:1487–95.
<https://doi.org/10.1038/ncb1998>
PMID:19935649
35. Seino T, Kawasaki S, Shimokawa M, Tamagawa H, Toshimitsu K, Fujii M, Ohta Y, Matano M, Nanki K, Kawasaki K, Takahashi S, Sugimoto S, Iwasaki E, et al. Human Pancreatic Tumor Organoids Reveal Loss of Stem Cell Niche Factor Dependence during Disease Progression. *Cell Stem Cell*. 2018; 22:454–67.e6.
<https://doi.org/10.1016/j.stem.2017.12.009>
PMID:29337182
36. Chen B, Dodge ME, Tang W, Lu J, Ma Z, Fan CW, Wei S, Hao W, Kilgore J, Williams NS, Roth MG, Amatruda JF, Chen C, Lum L. Small molecule-mediated disruption of Wnt-dependent signaling in tissue regeneration and cancer. *Nat Chem Biol*. 2009; 5:100–7.
<https://doi.org/10.1038/nchembio.137>
PMID:19125156
37. van de Wetering M, Sancho E, Verweij C, de Lau W, Oving I, Hurlstone A, van der Horn K, Batlle E, Coudreuse D, Haramis AP, Tjon-Pon-Fong M, Moerer P, van den Born M, et al. The beta-catenin/TCF-4 complex imposes a crypt progenitor phenotype on colorectal cancer cells. *Cell*. 2002; 111:241–50.
[https://doi.org/10.1016/S0092-8674\(02\)01014-0](https://doi.org/10.1016/S0092-8674(02)01014-0)
PMID:12408868
38. Walker F, Zhang HH, Odorizzi A, Burgess AW. LGR5 is a negative regulator of tumorigenicity, antagonizes Wnt signalling and regulates cell adhesion in colorectal cancer cell lines. *PLoS One*. 2011; 6:e22733.
<https://doi.org/10.1371/journal.pone.0022733>
PMID:21829496
39. Fukuma M, Tanese K, Effendi K, Yamazaki K, Masugi Y, Suda M, Sakamoto M. Leucine-rich repeat-containing G protein-coupled receptor 5 regulates epithelial cell phenotype and survival of hepatocellular carcinoma cells. *Exp Cell Res*. 2013; 319:113–21.
<https://doi.org/10.1016/j.yexcr.2012.10.011>
PMID:23127514
40. Gong X, Carmon KS, Lin Q, Thomas A, Yi J, Liu Q. LGR6 is a high affinity receptor of R-spondins and potentially functions as a tumor suppressor. *PLoS One*. 2012; 7:e37137.
<https://doi.org/10.1371/journal.pone.0037137>
PMID:22615920
41. Minami F, Sasaki N, Shichi Y, Gomi F, Michishita M, Ohkusu-Tsukada K, Toyoda M, Takahashi K, Ishiwata T. Morphofunctional analysis of human pancreatic cancer cell lines in 2- and 3-dimensional cultures. *Sci Rep*. 2021; 11:6775.
<https://doi.org/10.1038/s41598-021-86028-1>
PMID:33762591
42. Barker N, Clevers H. Leucine-rich repeat-containing G-protein-coupled receptors as markers of adult stem cells. *Gastroenterology*. 2010; 138:1681–96.
<https://doi.org/10.1053/j.gastro.2010.03.002>
PMID:20417836
43. Koo BK, Clevers H. Stem cells marked by the R-spondin receptor LGR5. *Gastroenterology*. 2014; 147:289–302.

<https://doi.org/10.1053/j.gastro.2014.05.007>

PMID:[24859206](#)

44. Kasper M, Jaks V, Are A, Bergström Å, Schwäger A, Svård J, Teglund S, Barker N, Toftgård R. Wounding enhances epidermal tumorigenesis by recruiting hair follicle keratinocytes. *Proc Natl Acad Sci USA*. 2011; 108:4099–104.

<https://doi.org/10.1073/pnas.1014489108>

PMID:[21321199](#)

45. Snippert HJ, Haegebarth A, Kasper M, Jaks V, van Es JH, Barker N, van de Wetering M, van den Born M, Begthel H, Vries RG, Stange DE, Toftgård R, Clevers H. Lgr6 marks stem cells in the hair follicle that generate all cell lineages of the skin. *Science*. 2010; 327:1385–9.

<https://doi.org/10.1126/science.1184733>

PMID:[20223988](#)

46. Collisson EA, Bailey P, Chang DK, Biankin AV. Molecular subtypes of pancreatic cancer. *Nat Rev Gastroenterol Hepatol*. 2019; 16:207–20.

<https://doi.org/10.1038/s41575-019-0109-y>

PMID:[30718832](#)

47. Waddell N, Pajic M, Patch AM, Chang DK, Kassahn KS, Bailey P, Johns AL, Miller D, Nones K, Quek K, Quinn MC, Robertson AJ, Fadlullah MZ, et al, and Australian Pancreatic Cancer Genome Initiative. Whole genomes redefine the mutational landscape of pancreatic cancer. *Nature*. 2015; 518:495–501.

<https://doi.org/10.1038/nature14169>

PMID:[25719666](#)

48. Daemen A, Peterson D, Sahu N, McCord R, Du X, Liu B, Kowanzetz K, Hong R, Moffat J, Gao M, Boudreau A, Mroue R, Corson L, et al. Metabolite profiling stratifies pancreatic ductal adenocarcinomas into subtypes with distinct sensitivities to metabolic inhibitors. *Proc Natl Acad Sci USA*. 2015; 112:E4410–7.

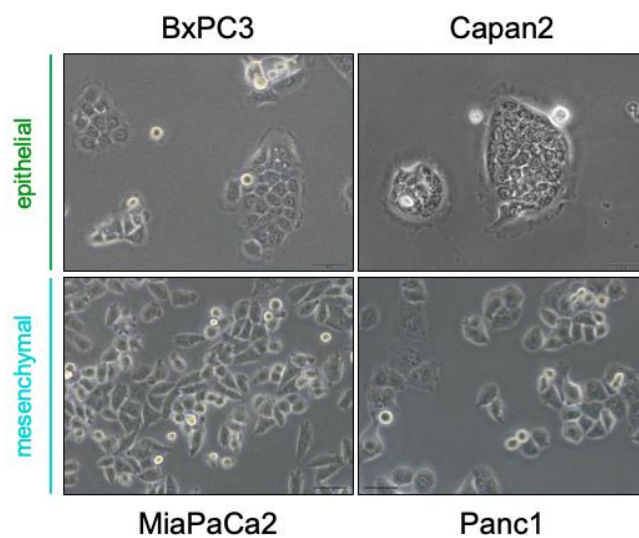
<https://doi.org/10.1073/pnas.1501605112>

PMID:[26216984](#)

SUPPLEMENTARY MATERIALS

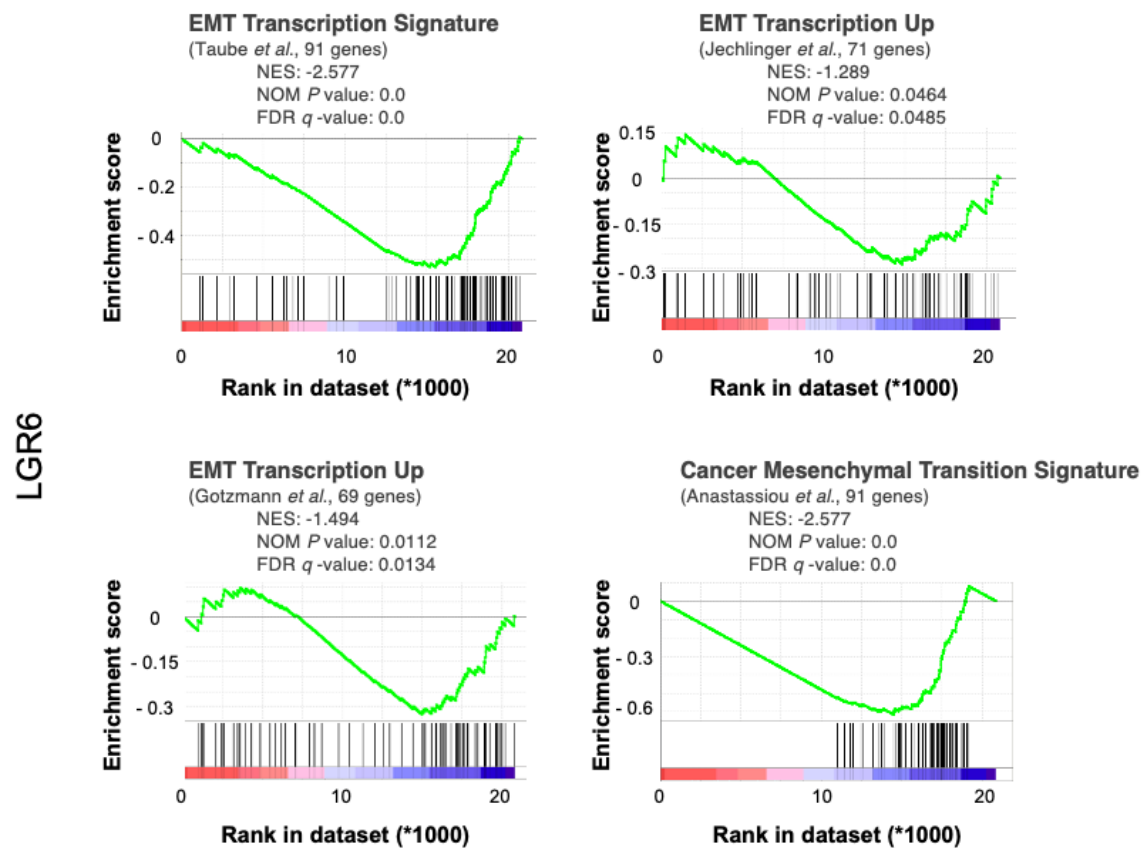
Supplementary Figures

Morphology of all PDAC cell lines used in this study in 2D



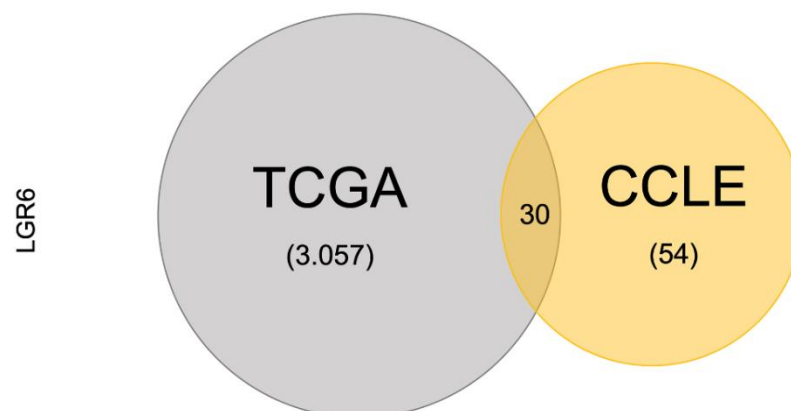
Supplementary Figure 1. 2D-Morphology of PDAC cell lines used in this study. Grouping according to epithelial (green; BxPC3, Capan2) or mesenchymal (blue; MiaPaCa2, Panc1) phenotype.

Prediction of inverse correlation between LGR6 and EMT signatures in PDAC

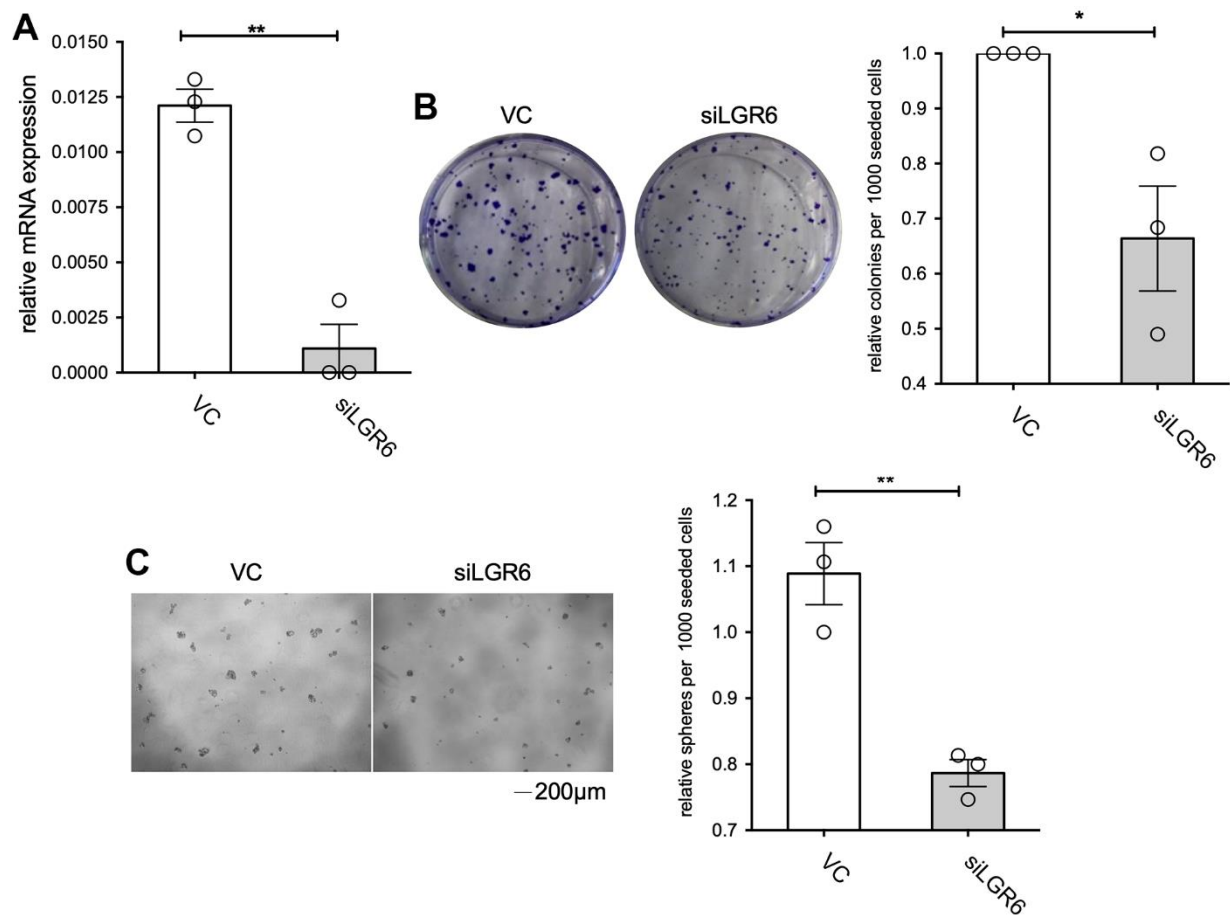


Supplementary Figure 2. GSEA analysis of typical EMT signatures in TCGA data (PDAC).

VENN Diagram LGR6^{high} vs. LGR6^{low} TCGA and CCLE Overlap



Supplementary Figure 3. VENN diagram of overlap analysis (TCGA in grey and CCLE in yellow); comparison of LGR6^{high} to LGR6^{low} expression (overlapping genes are listed in Supplementary Table 2).



Supplementary Figure 4. LGR6 knock-down reduces features of cancer stemness in PDAC. (A) Knock-down (KD) efficiency was tested by PCR for LGR6 mRNA levels; vector control (VC) vs. siLGR6 revealed significant KD. (B) colony-formation units and (C) sphere-formation ability after VC or siLGR6 treatment was carried out and resulted in decreased and smaller colony or sphere numbers.

Supplementary Tables

Supplementary Table 1. Antibodies used in this study.

Antibodies	Brand	Application	Dilution
LGR6	Sigma, HPA008556	WB	1:1000
E-Cadherin	BD Biosciences, 610181	WB	1:1000
beta-Catenin	Cell Signaling Technologies (CST) #8480	WB	1:1000
phospho-beta-Catenin (Ser675)	CST #4176	WB	1:1000
GAPDH	Santa Cruz Biotechnology (SCBT), sc-25778	WB	1:1000
Mouse IgG-HRP	SCBT, sc-2005	WB	1:2500
Rabbit IgG-HRP	CST, #7074	WB	1:2500
LGR6	Sigma, HPA008556	IF	1:50
beta-Catenin	CST, #8480	IF	1:100
Mouse IgG-FITC	SCBT, sc-2010	IF	1:100
Rabbit IgG-FITC	SCBT, sc-2012	IF	1:100
Mouse IgG-TR	SCBT, sc-2781	IF	1:100
LGR6 APC-conjugated	R&D Systems, FAB8458A-025	Flowcytometry	Acc. to manufacturer

Supplementary Table 2. Common genes of overlap analysis (TCGA and CCLE data) with comparison of LGR6^{high} to LGR6^{low} expression (related to VENN diagram in Supplementary Figure 3).

ID	Full name
AKR1B10	aldo-keto reductase family 1 member B10
ALDH1A1	aldehyde dehydrogenase 1 family member A1
ALDH3A1	aldehyde dehydrogenase 3 family member A1
ANO1	anoctamin 1
BCHE	butyrylcholinesterase
BMP4	bone morphogenetic protein 4
BST2	bone marrow stromal cell antigen 2
CEACAM5	CEA cell adhesion molecule 5
CEACAM6	CEA cell adhesion molecule 6
CLIC3	chloride intracellular channel 3
CXCL14	C-X-C motif chemokine ligand 14
DPP4	dipeptidyl peptidase 4
EDAR	ectodysplasin A receptor
FGF19	fibroblast growth factor 19
FGFBP1	fibroblast growth factor binding protein 1
GPX3	glutathione peroxidase 3
KLK10	kallikrein related peptidase 10
KRT14	keratin 14
KRT17	keratin 17
KRT6A	keratin 6A
LY6D	lymphocyte antigen 6 family member D
MSLN	mesothelin
PLA2G10	phospholipase A2 group X
PRR15	proline rich 15
PSCA	prostate stem cell antigen

S100P	S100 calcium binding protein P
SERPINB5	serpin family B member 5
SPESP1	sperm equatorial segment protein 1
SPRR1B	small proline rich protein 1B
SPRR3	small proline rich protein 3

Supplementary Table 3. Clinical data (TCGA) in correlation with LGR6 mRNA expression in pancreatic ductal adenocarcinoma.

Characteristics	Total	<i>LGR6</i>		<i>P</i>
		Low	High	
All patients	178 (100.0)	62 (34.8)	116 (65.2)	
Gender				
Male	95 (56.5)	33 (34.7)	62 (65.3)	0.87
Female	73 (43.5)	24 (32.9)	49 (67.1)	
T-category				
T1	8 (5.0)	1 (12.5)	7 (87.5)	0.5696
T2	19 (11.8)	12 (63.2)	7 (36.8)	
T3	131 (81.4)	42 (32.1)	89 (67.9)	
T4	3 (1.8)	1 (33.3)	2 (66.7)	
Nodal Metastasis				
Negative	42 (26.6)	15 (35.7)	27 (64.3)	0.8504
Positive	116 (73.4)	39 (33.6)	77 (66.4)	
Distant Metastasis				
Negative	74 (45.7)	26 (35.1)	48 (64.9)	>0.9999
Positive	88 (54.3)	31 (35.2)	57 (64.8)	
Diabetes				
Negative	98 (73.1)	35 (35.7)	63 (64.3)	>0.9999
Positive	36 (26.9)	13 (36.1)	23 (63.9)	
Alcohol history				
Negative	57 (37.5)	22 (38.6)	35 (61.4)	0.7277
Positive	95 (62.5)	33 (34.7)	62 (65.3)	
History Chronic Pancreatitis				
Negative	117 (90.7)	43 (36.8)	74 (63.2)	>0.9999
Positive	12 (9.3)	4 (33.3)	8 (66.7)	

Values in parentheses indicate column and row percentage for total and *LGR6* low or high cases, respectively.