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## Effects of temperature gradient on functional fruit traits: an elevation-for-temperature approach



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### **Abstract**

Fruit traits mediate animal-plant interactions and have to a large degree evolved to match the sensory capacities and morphology of their respective dispersers. At the same time, fruit traits are affected by local environmental factors, which may affect frugivore-plant trait match. Temperature has been identified as a major factor with a strong effect on the development of fruits, which is of serious concern because of the rising threat of global warming. Nonetheless, this primarily originates from studies on domesticated cultivars in often controlled environments. Little is known on the effect of rising temperatures on fruit traits of wild species and the implications this could have to seed dispersal networks, including downstream consequences to biodiversity and ecosystem functioning. In a case study of five plant species from eastern Madagascar, we addressed this using the elevationfor-temperature approach and examined whether a temperature gradient is systematically associated with variation in fruit traits relevant for animal foraging and fruit selection. We sampled across a gradient representing a temperature gradient of 1.5–2.6 °C, corresponding to IPCC projections. The results showed that in most cases there was no significant effect of temperature on the traits evaluated, although some species showed different effects, particularly fruit chemical profiles. This suggests that in these species warming within this range alone is not likely to drive substantial changes in dispersal networks. While no systemic effects were found, the results also indicate that the effect of temperature on fruit traits differs across species and may lead to mismatches in specific animalplant interactions.

**Keywords** Climate change, Functional traits, Temperature, Frugivory, Global warming, Seed dispersal

in this article, unless otherwise stated in a credit line to the data.

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## **Introduction**

Many tropical plants rely on animals for seed dispersal  $[1, 2]$  $[1, 2]$  $[1, 2]$  $[1, 2]$  in a mutualistic interaction in which frugivorous animals obtain nutritious food and plants disperse their seeds [[3,](#page-9-2) [4](#page-9-3)]. Frugivores differ in their senses, fruit handling capacities, foraging behavior, and preferences [[5](#page-10-0), [6](#page-10-1)] and fruit traits have to a large degree evolved to match the different sensory capacities and morphologies of their respective frugivores [\[7](#page-10-2), [8\]](#page-10-3). Therefore, trait matching has been recognized as an important mechanism mediating the structure of seed dispersal networks [[9](#page-10-4), [10\]](#page-10-5). Specifically, traits such as scent  $[6, 11]$  $[6, 11]$  $[6, 11]$  color  $[7, 12]$  $[7, 12]$  $[7, 12]$  $[7, 12]$ , size  $[13]$  $[13]$ ,

hardness [\[14](#page-10-9)] and nutritional content [[15](#page-10-10)] have been shown to play a major role in frugivore foraging and food selection.

At the same time, fruit traits are liable to environmental factors [\[4](#page-9-3), [16,](#page-10-11) [17\]](#page-10-12). Work primarily on cultivated species like mangos, avocados, strawberries, bananas and grapes, revealed that temperature in particular is a key factor affecting fruit development, together with humidity, UV-light intensity, day/night temperature variance, precipitation and soil type, although to a lesser degree [[16,](#page-10-11) [18](#page-10-13)]. During fruit development higher temperatures are associated with changes in fruits at the morphological, physiological and biochemical level [\[16\]](#page-10-11), but the responses of specific fruit traits to temperature are mixed [[17,](#page-10-12) [18](#page-10-13)]. In general, it has been observed that higher temperatures decrease fruit sugar content, size and hardness of fruits [[18](#page-10-13)[–20](#page-10-14)], increase the total emission of volatile organic compounds (VOCs) [[21](#page-10-15)[–23](#page-10-16)] and lead to a poor fruit color development [[24\]](#page-10-17).

The Intergovernmental Panel on Climate Change (IPCC) currently estimates that global temperature has increased by 1 °C compared to pre-industrial levels, and in order to limit temperature rising to 1.5 °C greenhouse gas emissions have to reach net zero by 2050 (>50% probability) [[25](#page-10-18)], being this the most optimistic scenario (SSP1-1.9). A more plausible scenario (SSP1-2.6) indicates the world can limit temperature rise to 2 °C, while stabilizing it in 1.8  $°C$  by the end of the century (>67% probability), assuming net zero by 2070 [[25](#page-10-18), [26](#page-10-19)]. As such, a temperature rise under the current IPCC scenarios that might affect fruit traits is not just possible, but highly likely. Therefore, because of the importance of traits shaping the interactions between fruits and frugivores [[27\]](#page-10-20), trait alterations that affect the way frugivores recognize ripe fruits might also have an impact on seed dispersal networks [\[28\]](#page-10-21). The dispersion of seeds by animals plays an important role on the structure, biodiversity, and maintenance of natural ecosystems dynamics [[29](#page-10-22)[–33](#page-10-23)], hence dramatic changes on it could threaten ecosystem functioning, particularly in tropical regions where seed dispersal by animals is more common [[34\]](#page-10-24).

For example, warming may drive higher rates of scent emission from fruits [[23\]](#page-10-16), which could translate into a stronger chemical signal if relevant secondary metabolites are not a limiting factor [\[35\]](#page-10-25), or to a reduction of the signal if it leads to faster depletion of synthesized chemicals. Such a scenario would be particularly relevant for systems like the one at focus – Madagascar – where fruit scent has been shown to drive animal fruit selection [\[11](#page-10-6), [36\]](#page-10-26). Color, is also strongly associated with fruit selection [[12,](#page-10-7) [37\]](#page-10-27); and as well as scent, it has been linked to nutrient content  $[11, 36-39]$  $[11, 36-39]$  $[11, 36-39]$  $[11, 36-39]$ . Finally, fruit morphological traits like size and hardness [\[40](#page-10-29)] are recognized as limitations for frugivores when selecting fruits, (i.e. birds cannot manipulate nor eat fruits larger than their gape width) [[41–](#page-10-30)[43\]](#page-10-31). All these, while hard to predict, can cause major disruptions to existing dispersal networks by (a) directly altering signals like scent and color or their link with fruit quality, thus reducing animal foraging efficiency; (b) differentially changing the preference of fruit assemblages, driving an increase in dispersal of some species at the expense of others [[13\]](#page-10-8); (c) reducing seed dispersal effectiveness [[44,](#page-10-32) [45\]](#page-10-33) by causing animals to consume more immature fruits, or by (d) changing interactions (e.g. rendering fruits smaller and more available to birds), thus causing a shift in the dispersal kernel of some species [[42](#page-10-34), [43\]](#page-10-31).

As such, quantitatively, this effect can manifest in different degrees and ways in different systems where ecological networks (e.g. specialized vs. generalized) and sensory redundancy differ. But whether or not this is at all a risk hinges on two assumptions: that fruit traits drive fruit selection, and that they are affected by temperature. While the first is well established (see above), despite its importance, the question of how global warming might affect wild fruit traits has been widely overlooked. Regarding wild populations, previous research has focused on other plant traits (height, leaf area, seedmass) and how they are affected by different environmental drivers (nitrogen deposition, precipitation) [[46](#page-10-35), [47\]](#page-10-36) and on the geographic distribution of traits based on environmental variables [\[48](#page-10-37)]. Whereas studies on fruit traits have had solely an anthropogenic interest, in the sense they have only looked into cultivated species to evaluate quality parameters according to consumer and market preferences [\[17,](#page-10-12) [49](#page-10-38), [50](#page-10-39)]. Regarding species interactions significant more attention has been given to the question of how temperature rise might affect flower traits and downstream flower-pollinators networks, indicating worrying trends which may or may not be similar to seed-dispersal networks [\[51,](#page-10-40) [52](#page-10-41)].

Under this framework, the goal of the current study was to evaluate, as a first proof of concept on a case study of five Malagasy plant species, the potential effect of temperature change on fruit functional traits of wild species. We used the elevation-for-temperature approach, in which elevational gradients provide a steady gradual temperature change of approximately −0.55 °C for every 100 m upslope [[53\]](#page-11-0), while allowing the comparison of individuals belonging to a genetically continuous population [[54](#page-11-1)], thus avoiding conflating genetically-determined phenotypic differences with variance driven by edaphic factors. This method has shown to be useful to predict how plants and other organisms might respond to global warming [[55–](#page-11-2)[58](#page-11-3)]. We analyzed fruit traits across a single slope in the montane primary forest of Mangevo, Madagascar, which compromises a continuous gradient of 520 m, providing an estimated temperature difference of 2.6 °C, from the lowest to the highest point, corresponding to end of the century IPCC projections and in line with projections for tropical regions [\[59](#page-11-4)]. Madagascar is a biodiversity hotspot where many local (often endemic) plants rely on the dispersal services of often endemic animals [\[60](#page-11-5), [61\]](#page-11-6) and it has already been used as a model system on animal-plant interactions in several studies [\[11](#page-10-6), [12,](#page-10-7) [14](#page-10-9), [38](#page-10-42), [62](#page-11-7)]. We quantified five functional traits and focused on those that have been shown to be strongly associated with fruit selection and preference among frugivores in this or comparable systems: scent, color, size, hardness and sugar content. In short, fruit scent is increasingly recognized as a driver of fruit selection [\[63](#page-11-8)], specifically in this system it was observed to be associated with frugivore behavior [\[11\]](#page-10-6) and be predictive of fruit quality (sugar content) [[11,](#page-10-6) [36](#page-10-26), [64\]](#page-11-9). Fruit color functions as a detection and selection signal to many frugivores [\[37,](#page-10-27) [65](#page-11-10), [66\]](#page-11-11), and particularly in this system has been observed to have evolved in response to frugivore visual systems [\[12](#page-10-7)]. Fruit size is a major driver of fruit selection in similar systems and beyond  $[14, 67]$  $[14, 67]$  $[14, 67]$ , particularly birds who tend not to feed on fruits/seeds larger than their gape width [[41,](#page-10-30) [43\]](#page-10-31). Fruit hardness has shown to be under an independent selection of mechanical constraints [[14](#page-10-9)] and be associated with fruit selection in similar systems [[40\]](#page-10-29). Finally, sugar is a major macronutrient sought after by frugivores and variation among fruits in sugar content has shown to affect fruit selection [[68](#page-11-13)[–70](#page-11-14)].

#### **Methods**

#### **Model system and sample analyzes**

Fruit traits were analyzed directly on the field between May and June 2022, in the montane rainforest of Mangevo, a protected area that has never been logged, in Ranomafana National Park, eastern Madagascar (Fig. [1\)](#page-2-0). We covered a single continuous forested slope from 580 to 1100 m above sea level (masl), which provided an estimated temperature difference of 2.6 °C. Across the elevational gradient, forest canopy closure was not significantly different [\[71](#page-11-15)] and precipitation was homogeneous.

Five species were selected based on the availability of ripe fruits across the gradient and their being at a reachable distance (max 4 m): *Ficus botryoides*, species with a lemur-specialist dispersion syndrome [[11\]](#page-10-6) and *Ficus politoria*, *Psychotria* sp1, *Psychotria* sp2, and *Pittosporum verticillatum*, all primarily dispersed by birds with some dispersal by lemurs [[36\]](#page-10-26). The species sampled belonged to a single functional population, in which pollinators and frugivores can travel freely across the gradient [\[72–](#page-11-16)[75\]](#page-11-17). All species are insect pollinated. Given that samples were all from a continuous community along a single slope, where seed dispersers and pollinators populations are also continuous on a relatively short distance that is very minor compared to disperser movement range, there is no reason to assume that genetic factors alone would generate a trait value gradient. Two to eight fruits were obtained from an individual plant and pooled together as a single sample (Table [1](#page-3-0)). Plants were identified on the field by a local expert: Pela August (technician at Ranomafana National Park), and no plant material was exported. Fruits recognized as ripe by their softening when mature, presence of mature seeds and their specific change in color. Fruits were brought within 3 h to the field processing station to measure scent, color, size, hardness and sugar content, in this order. The fruits are

<span id="page-2-0"></span>

**Fig. 1** Map of Madagascar, showing Mangevo coordinates and elevational gradient

<b>Species</b>	<b>Number of individuals</b>	Number of samples	Gradient (masl)	Temperature variance approximation (°C)
Ficus botryoides			627-903	
Ficus politoria		38	627-1069	2.4
Psychotria sp 1		70	597-1072	2.6
Psychotria sp 2		54	628-918	- 6
Pittosporum verticillatum			680-993	

<span id="page-3-0"></span>**Table 1** Description of number of samples analyzed for each species

not suspected to systematically differ in their time on the maturation curve, which could in theory lead to a trait gradient if e.g. fruits mature earlier in lower elevations. This is because (a) all species had relatively low synchrony and hence crop of mature fruits at any given moment, which leads to a relatively quick removal by animals; and (b) we found very low correlation between fruit traits within species (mean  $r=0.12$ ), negating the possibility that some fruits were significantly more ripe than others.

#### **Analysis fruit traits**

#### *Scent*

Fruit scent was sampled following a similar methodology to  $[11]$  $[11]$ . Fruits were placed in sampling bags of 40 cm (Toppits oven bags, Toppits). The bags were tightly closed with a zip tie at one end and on the other a Teflon tube with a chromatoprobe was mounted and carefully tighten. The chromatoprobes contained trapped in layers of glass wool 1.5 mg of Tenax, 1.5 mg of Carbotrap and 1.5 mg of Carbosieve III (all Sigma Aldrich). The samples were left to rest in the chamber for 20 min, afterwards the air was pumped for 10 min at 200mL/min. The probes were stored in 2 mL glass vials sealed with a Teflon cap and placed in at -32 °C upon return to the field station and until analysis. Controls to identify ambient contaminants were collected following the same procedure with empty bags.

**Chemical analyses** Samples were analyzed using Shimadzu GCMS-QP2020 NX equipped with an TD30-R thermo desorption unit and a Shimadzu SH-RXI-5MS low polarity phase capillary column (SH-RXI-5MS, 30 m, 0.25 mm diameter). Samples were introduced to the thermal desorption unit and the tube heating started at 35 °C until it reached 300 °C. a Tenax liner was cooled to -20 °C. After the transfer to the liner, it was heated up until 230 °C. All samples were introduced into the system spitless, with the exception of *Pi.verticillatum*, for which split was set to 1:60 due to a high VOC signature that otherwise overloaded the column and resulted in broad unquantifiable peaks. As such, quantitative measures of fruit scent in this study are comparable within but not among all species. Initial oven temperature was 40 °C. This temperature was maintained for 1 min, then it was increased by 10 °C/min until it reached 280 °C, where it was held for 8 min. The MS transfer line temperature was set to 250 °C and MS source temperature was set to 230 °C. The MS operated at electron ionization mode and scanned between 30 and  $400 \frac{m}{z}$ 

Chromatograms obtained were analyzed in AMDIS 2.7. VOCs were identified on the basis of their mass spectra by comparing them with the NIST11 mass spectra library and their retention index, which was calculated using a *n-*alkane reference mixture. Known contaminants (e.g. siloxanes) were fully removed from the dataset, whereas for genuine plant VOCs which were identified in both the controls and samples, we calculated the mean amount in the controls as a proxy of the baseline contamination level, and subtracted this amount from all samples. Peak area of all compounds was then divided by the respective number of fruits in each sample to obtain a standardized measure per functional unit (fruit).

VOCs were further classified to seven chemical classes based on functional or biosynthetic groups: alcohols, terpenes (including all monoterpenes, sesquiterpenes and derivates such as linalool), aldehydes, esters, aromatic (compounds with at least one aromatic ring), alkanes, and nitrogen containing compounds. The total amount of each class in each sample was calculated as the sum of all peaks belonging to the class.

#### *Color*

Fruits were photographed using a stand-mounted digital camera (Canon EOS rebel t3i, Canon Inc., Tokyo Japan) with a 18–55 mm lens (Canon EF-S 18–55 mm), at a focal length of 55 mm with no flash. The camera was positioned 40 cm directly in front of the fruits. The shots were intentionally underexposed to avoid the loss of data by overexposing [[76](#page-11-18), [77](#page-11-19)]. To calibrate and standardize the photos we used a color chart (ColorChecker Classic, X-Rite, Grand Rapids, MI, USA) [[78\]](#page-11-20). The fruits were placed in front of the chart. Because the chart and the fruits could not be focused on at the same time, a first shot focusing the fruits was taken and immediately after a second shot was taken focusing the chart, this assured the pictures had the same light conditions and camera settings [\[77](#page-11-19), [79](#page-11-21)]. All files were exported in the RAW format, because it is linear and displays a wider variety of colors [[77,](#page-11-19) [79](#page-11-21)] .

The photographs were processed in ImageJ v1.8.0\_172 [[78\]](#page-11-20) using the micaToolbox plugin  $[80]$ . Each photo was

converted into a multispectral image, using the 5% and 95% reflectance standards and the regions of interest were selected [\[77](#page-11-19), [78\]](#page-11-20). Because the camera was not UV sensitive the analyses were restricted to visible spectrum (400–700 nm) and the percentage values of the red, green and blue (RGB) color channels were extracted from each photograph.

#### *Size*

The size of each fruit was determined by measuring the dimensions along cardinal directions (length, width and depth) using sliding calipers. Afterwards, the fruit volume was stablished using the ellipsoid formula:  $V = \frac{4}{3}$  *∗* πabc [\[14](#page-10-9)].

#### *Hardness*

Hardness was defined as skin puncture resistance in *kg/mm*<sup>2</sup> and measured using a hand-held durometer (Shimpo MX)  $[14, 40, 81]$  $[14, 40, 81]$  $[14, 40, 81]$  $[14, 40, 81]$  $[14, 40, 81]$  $[14, 40, 81]$ . Fruit hardness was not measured for the species *P. verticillatum*, because of the dehiscent nature of this species when ripen.

#### *Sugar content*

Consistently for each species, based on the size of the fruits, between 0.5 and 2 gr were macerated with 1–2 mL of water (*F.botryoides* 2gr with 2mL, *F. politoria* 1gr with 1mL, *Psychotria* sp 1 0.5gr with 1mL and *Psychotria* sp 2 and *P. verticillatum* 1gr with 2 mL), until a homogenous solution was obtained. Four drops of the solution were used to measure the sugar content with a refractometer (MASTER-AGRI, Atago). Refractometers have demonstrated to be good instruments to measure the total content of sugar in fruits [[82,](#page-11-24) [83](#page-11-25)]. Results are expressed in Brix/TSS (total soluble solids) and represent the percentage of sugar present in a fruit. Given that the amount of fruit flesh and water added was standardized within species, these results allow a reliable comparison of the relative concentration of sugar *within species and only within our dataset* but are not valid for across-species comparisons or for estimation of absolute amounts.

#### **Statistical analyses**

Multiple samples from the same individual tree were pooled by averaging values, making individual tree the basic datapoint in all analyses (Table [1](#page-3-0)). To analyze the effect of temperature on each fruit trait, we applied two similar but complementary approaches. The first was generalized linear models (GLMs) conducted for each species separately in which a trait was set as the response variable: total amount of VOCs, total amount of each VOC class, percentage RGB color channels, hardness  $\left(kg/mm^2\right)$ , size  $\left(mm^3\right)$  and sugar content (Bx°). Elevation in m was the sole predictor (fixed) factor. We then applied a Hommel correction for multiple testing. For all models, we verified the normal distribution of the random effect as well as the normality and homogeneity of the residuals using histograms, quantile-quantile plots, and plotting the fitted vs. the residuals. In the cases where assumptions were not met, the data was log-transformed before running the model. These analyses were conducted because we did not have a clear directional hypothesis (e.g. that higher elevations are associated with redder fruits). In addition, we also ran generalized linear mixed models (GLMMs) with a gaussian error structure in which each of the predictors mentioned above was a sole predictor, elevation was a fixed factor, and species as a random intercept factor. This analysis was set to test if trends across species are similar or not, although it should be noted that the use of species as a random factor violates the assumption of full randomness. *P*-values were calculated by comparing the full models with their respective null models, which did not include elevation in a likelihood ratio test. All analyses were performed in *R* studio 4.1.2 with the package lme4 [[84](#page-11-26)].

#### **Results**

The linear models showed that after correction for multiple testing, most traits did not change across the gradient within species ( $p > 0.05$ ), with some exceptions: hardness and esters in *Psychotria* sp1 had a positive correlation with elevation ( $p = 0.03$ ;  $p = 0.039$ , respectively), whereas the total amount of VOCs in *P. verticillatum* and total amount of volatile alcohols in *Psychotria* sp2 had a negative correlation with elevation  $(p=0.036;$  $p=0.029$  respectively) (Table S1). All mixed models examining fruit traits as a function of elevation were not significantly different from their respective null models  $[X^2 = 0(1), P = 1]$ , in all models), indicating no consistent effect of elevation on any trait across the gradient (sugar content, size and hardness: Fig. [2](#page-5-0); RGB color channels: Fig. [3;](#page-6-0) scent: Fig. [4](#page-7-0)**)**.

#### **Discussion**

The objective of our study was to use the elevation-fortemperature approach to estimate, for the first time to our knowledge, whether temperature variance may affect a set of wild fruit functional traits that are known to be relevant for seed-dispersal networks. We did so on a case study of five Malagasy plant species sampled along a continuous forested slope between 597 and 1072 masl, corresponding to a temperature variance between 1.5 and 2.6 °C [[53](#page-11-0)] – in line with the most likely IPCC scenarios to the end of the century [\[25](#page-10-18), [26\]](#page-10-19) and in line with predicted temperature increase in the tropics [[59\]](#page-11-4).

#### **Scent**

Scent is possibly the trait most likely to be affected by temperature change as it is comprised of dozens to

<span id="page-5-0"></span>

**Fig. 2** Relationship between elevation (masl) and fruits sugar content; size; hardness, green slope with *Psychotria* sp1 (*p=0.03*), in all five species. Sugar content was measured in °Brix with a refractometer by macerating 0.5-2gr with 1-2mL of water (depending on the species). Size was calculated based on fruit dimensions assuming ellipsoid shape in *mm*<sup>3</sup>. Hardness was defined as skin puncture resistance in *kg/mm*<sup>2</sup> . All data log transformed to comply with the statistical test assumptions

hundreds of different VOCs and each can be affected differently by ambient temperature. In contrast to other traits, it can be affected directly by changing enzymatic efficiency or VOC volatilization from the fruit or indirectly (temperature changes growth conditions which lead to differential VOC synthesis). In our study *P. verticillatum* was the only species which showed a higher amount of total amount of VOCs at higher temperatures, whereas the total amount of VOCs of the other species did not change. This increase of VOCs at higher temperatures agrees with previous studies that argue the world will become a more scented place with global warming, as high temperatures enhance enzymatic activity of VOCs biosynthesis, decrease diffusion pathway resistance and increase volatility [[21](#page-10-15), [22,](#page-10-43) [35](#page-10-25)].

Nonetheless, existing data on mostly cultivated species also does not reveal a systematic pattern of increase in VOC emission with temperature [\[23](#page-10-16), [50,](#page-10-39) [85](#page-11-27)]. This indicates that any effect of temperature on the total emission of VOCs may be subtle and possibly limited to the responses each compound class; therefore, the changes on the chemical profile of species probably depend on the specific compounds that constitute it. In our results, different classes of compounds showed different responses

<span id="page-6-0"></span>

Fig. 3 Relationship between elevation (masl) and red%; green%; blue% in all five species. The RGB percentage values were extracted in ImageJ software using the MicaToolbox plugin. All photographs were standardized with a color chart (ColorChecker Classic, X-Rite, Grand Rapids, MI, USA)

to the temperature gradient: while volatile alcohols and terpenes decreased at higher elevations in *Psychotria* sp 2 and *P verticillatum* respectively, esters of *Psychotria* sp 1 increased at higher elevations as well as aromatic compounds in *F. botryoides* (see Table S1). Although sample size of esters in *Psychotria* sp 1 was very small, previous results by [[85](#page-11-27)] also showed an increase of this class of compound at higher elevations in cultivated bananas. We recognize the correlation of terpenes in *P. verticillatum* and aromatic compounds in *F. botryoides* were not statistically significant after correction for multiple testing. However, previous studies in flowers suggest these results could be genuine  $[21, 86]$  $[21, 86]$  $[21, 86]$  $[21, 86]$  $[21, 86]$ , still further examination is required.

Future studies should aim to elucidate the effect of temperature on: (1) different species, especially those with a mammalian dispersal syndrome, as is in these species where scent plays a major role in fruit recognition [[11,](#page-10-6) [87\]](#page-11-29) and consequently in the ones that changes could cause major disturbances in ecosystems [\[88\]](#page-11-30); and (2) different compounds and classes of compounds, focusing attention on those that have shown to play a key role in

<span id="page-7-0"></span>

**Fig. 4** Relationship between elevation (masl) and total amount (TA) of volatile organic compounds, light blue slope with *P. verticillatum* (*p*=0.036); volatile esters, green slope with *Psychotria* sp1 (*p=*0.039), and volatile alcohols, yellow slope with *Psychotria* sp2 (*p=*0.029), in all five species. Total amount of VOCs was defined as the summatory of the area of all peaks in all samples and total amount of each class of compound was defined as the summatory of the area of all peaks corresponding to each class in all samples. All data log transformed to comply with the statistical test assumptions

fruit recognition by seed dispersers, as are esters or terpenoids [[36,](#page-10-26) [63\]](#page-11-8). Changes in the VOCs profile of flowers have already shown to be detrimental in pollination networks, as they have altered fidelity and probability of pollinators recognition [\[21](#page-10-15)], highlighting the importance of scent in mutualistic networks.

#### **Color**

Fruit color is sensible to temperature changes, because temperature can influence on the synthesis of pigments (e.g.: anthocyanins and carotenoids) [\[24](#page-10-17)] and for most species increases in temperature have shown to downregulate the genes involved in these pathways [\[24](#page-10-17), [89](#page-11-31)]. Accordingly, global warming has the potential to lead to an impairment in fruit coloration that can negatively alter the way frugivores identify ripe fruits. In our study,

none of the fruits evaluated presented a change in their color channels in response to the temperature gradient. Previous studies on cultivated species have shown that significant changes in fruit coloration usually occur at temperatures above 27ºC [[90–](#page-11-32)[92](#page-11-33)], therefore the species evaluated here most probably did not change because temperatures did not surpass 22ºC. Nonetheless, because other regions most likely will exceed 27ºC with global warming [[93](#page-11-34)], it is important that future studies foresee the implications this could have for seed dispersal networks. Future works on color should ideally include fruit reflectance at the UV part of the spectrum. This is particularly relevant for birds and while requiring special instrumentation, it has been successfully demonstrated in this and other model systems [[94](#page-11-35)].

Another environmental variable that regulates fruit coloration is solar UV radiation and contrary to temperature, it has been observed that UV exposure increases the synthesis of pigments [\[18](#page-10-13), [81](#page-11-23), [82\]](#page-11-24). UV-radiation increases with altitude, up to 10% for every 1000 m, and consistently several studies on cultivated species show that fruits at higher elevations have a darker as well as a more intense color [[18,](#page-10-13) [82,](#page-11-24) [95](#page-11-36)]. However, this induced accumulation of pigments by UV-radiation is limited by increased temperatures [\[91,](#page-11-37) [96](#page-11-38)]. The species evaluated here, were not affected by UV-radiation because forest canopy was homogenous across the elevational gradient [[71\]](#page-11-15).

#### **Size**

Temperature affects fruit size by accelerating (high temperatures) or slowing down (low temperatures) the ripening process [[18\]](#page-10-13). A slower ripening leads to a prolonged accumulation of photo-assimilates that ultimately leads to fruits being bigger [[81](#page-11-23), [97\]](#page-11-39). Fruit size did not change for any of the species evaluated here. This is most probably because the temperature variance was not enough to cause changes in the photo-assimilates accumulation rate of the species examined.

#### **Hardness**

Previous studies on cultivated species indicate temperature affects fruit hardness; however, the directionality (i.e., whether fruits become softer or harder) by an increase in temperature varies across species [[97](#page-11-39)[–99](#page-11-40)]. In this study, only one species *Psychotria* sp1 increased its hardness at lower temperatures (higher elevations), whereas the other species did not change.

To explain the mixed responses of fruit hardness towards temperature, authors have proposed that this trait strongly depends on cell wall composition as well as calcium and starch concentration, intrinsic physiological characteristics related to fruit softening  $[18]$  $[18]$ . Therefore, more research is needed to better elucidate the ways in which global warming might affect fruit hardness, more importantly because further environmental changes are expected with global warming. For instance, changes in rainfall regimes, which can subsequently alter UV-radiation and relative humidity through changes in cloud patterns [\[100,](#page-11-41) [101](#page-12-0)]; factors that have also shown to influence in fruit hardness: while UV has shown to increase it, relative humidity to decrease it [\[16,](#page-10-11) [18](#page-10-13), [81\]](#page-11-23).

#### **Sugar content**

Temperature at which a fruit develops influences the amount of sugar content  $[18]$  $[18]$ . According to previous studies, a mild increase in temperature can promote the translocation of sugars to fruits, hence increasing sugar content [\[97](#page-11-39)]. Nonetheless, when the increase of temperature surpasses the normal range of species, it causes a loss of carbohydrates by an acceleration in respiration [\[18,](#page-10-13) [81](#page-11-23), [99\]](#page-11-40). It is therefore, of high concern that fruits become less sugary as a consequence of global warming, as it could cause a deficit in the energy frugivores obtain when foraging. The fruits studied here did not change their fruit sugar content in response to the temperature gradient, probably because it was inside their normal range. However, other regions might experience increases in temperature that are outside the tolerant ranges of populations [[102\]](#page-12-1), it is then important that future studies bring attention to changes in fruit sugar content in response to global warming.

#### **Synthesis, caveats, and conclusions**

Under the temperature gradient examined, which corresponds to IPCC predictions for the end of the century, most of the traits in the five species included did not show significant changes. This indicates prima facia that fruit functional traits, measured in the model system studied here, may not be strongly, or at least systematically, affected by anticipated temperature rise by the end of the century and that most likely the frugivores present will be able to keep on relying on these traits to successfully identify ripe fruits. However, this pattern may also be explained by other factors. A likely explanation is that the current projected change is inside the tolerant temperature range of the species examined, hence suggesting species are likely to maintain a similar expression of their functional traits. Nonetheless, while the projected temperature increase will not push the species studied here into an environment that substantially change their fruit phenotypes, further temperature rise may still lead to change, and individuals growing in higher or lower elevations will find themselves outside their optimal temperature range. This can lead to a shift in fruit traits, altering fidelity in which frugivores successfully identify ripe fruits and potentially affecting forest regeneration and structure.

Another important caveat is that the model system used here was limited to only five species. While significant in the sense that data on this question from the wild are all but absent, it is imperative for future studies to build on our results – expanding model species, traits, season variation, and elevation (or temperature directly) gradients – to assess whether these patterns are an exception or the rule.

Further, an advantage of the elevation-for-temperature approach is that it can assess the effect of temperature independent of many other factors such as rainfall. Yet global warming is likely to bring about many other changes in addition to mere temperature change: precipitation patterns, wind regimes, soil degradation and other factors which together may drive more significant and disruptive changes to fruit traits [\[103–](#page-12-2)[105\]](#page-12-3). Along these lines, global warming may directly and indirectly change animal populations and behavior, with potential downstream effects for seed dispersal dynamics. Finally, global warming may affect fruit traits indirectly by altering pollination patterns.

It is important to note that abiotic conditions along an elevational gradient are not always restricted to temperature, and may include variation in rainfall patterns, soil quality, etc., yet given the relatively high homogeneity along the gradient studied here [[71\]](#page-11-15), these factors are unlikely to have affected the results. Therefore, we can infer that nor temperature, which did change across the elevational gradient, nor other environmental factors strongly affected the fruit traits in this system.

As such, our results should be taken only as an indication that global warming is not likely to dramatically and immediately change fruit phenotypes across all traits and in all fleshy fruits. By no means do they indicate that global warming will not change fruit traits and downstream animal-plant interactions and seed dispersal networks. Indeed, the rich body of literature on cultivated species indicates that effects are not uniform and not always linear. Rather, they indicate that the effects are complex and will differ across species and systems. Therefore, we highlight the need for more nuanced analyses on wild species across systems, that contribute to test predictions on the possible downstream effects that trait alterations could cause on the ecosystem functions sustained by seed dispersal interactions. We hope that our work will serve as a starting point in this endeavor so that species and systems which are vulnerable to a rapidly changing world are identified sooner rather than later.

#### **Abbreviations**

- IPCC Intergovernmental Panel on Climate Change
- RGB Red, green, blue
- VOCs Volatile organic compounds

#### **Supplementary Information**

The online version contains supplementary material available at [https://doi.](https://doi.org/10.1186/s12862-024-02271-w) [org/10.1186/s12862-024-02271-w.](https://doi.org/10.1186/s12862-024-02271-w)

Supplementary Material 1

Supplementary Material 2

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#### **Author contributions**

ON formulated the idea. ON and LGD obtained funding. LGD developed the methodology, conducted fieldwork and performed the statistical analyses. ON and LGD wrote the manuscript.

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#### **Data availability**

All raw data is available as online supplementary material.

#### **Declarations**

#### **Ethics approval and consent to participate**

Permission to perform this research in Mangevo forest located in Ranomafana National Park was obtained from the proper authority. Madagascar Institute pour la Conservation des Ecosystèmes Tropicaux (MICET) was the body in charge that provided us with the corresponding research permit: 079/22/ MEDD/SG/DGGE/'DAPRNE/SCBE. Plant species were identified on the field and no plant material was exported; therefore, no specimens were vouchered in a publicly herbarium. Fruits traits were directly measured on the field and no fruit samples were exported. Fruit scent (volatile organic compounds) were collected on the field and further on exported under the export permit number: 159/22/MEDD/SG/DGF/DREDD\_y\_7\_y, also obtained trough MICET.

#### **Consent for publication**

Not applicable.

#### **Competing interests**

The authors declare no relevant competing interests.

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#### **References**

- <span id="page-9-0"></span>1. Fleming TH, Breitwisch R, Whitesides GH. Patterns of tropical vertebrate frugivore diversity. Annu Rev Ecol Syst. 1987;18:91–109.
- <span id="page-9-1"></span>2. Howe F, Smallwood J. Ecology of seed dispersal. Annu Rev Ecol Syst. 1982;13:201–28.
- <span id="page-9-2"></span>3. Jordano P. (2000). Fruits and frugivory. In Gallagher, R. S, editor, *Seeds: The Ecology of Regeneration in Plant Communities*, (pp. 125–165). CABI, 2013.
- <span id="page-9-3"></span>4. Ruxton GD, Schaefer MH. The conservation physiology of seed dispersal. Philosophical Trans Royal Soc B: Biol Sci. 2012;367:1708–18.
- <span id="page-10-0"></span>5. Lomáscolo SB, Levey DJ, Kimball RT, Bolker BM, Alborn HT. Dispersers shape fruit diversity in Ficus (Moraceae). Proc Natl Acad Sci USA. 2010;107:14668–72.
- <span id="page-10-1"></span>6. Hodgkison R, Ayasse M, Schulz S, Zubaid A, Aida Mustapha WW, Kunz TH,v, Kalko EK. Fruit bats and bat fruits: the evolution of fruit scent in relation to the foraging behaviour of bats in the New and Old World tropics. Funct Ecol. 2013;27:1075–84.
- <span id="page-10-2"></span>7. Sinnott-Armstrong MA, Donoghue MJ, Jetz W. Dispersers and environment drive global variation in fruit colour syndromes. Ecol Lett. 2021;24:1387–99.
- <span id="page-10-3"></span>8. Valenta K, Nevo O. The dispersal syndrome hypothesis: how animals shaped fruit traits, and how they did not. Funct Ecol. 2020;34:1158–69.
- <span id="page-10-4"></span>9. McFadden IR, Fritz SA, Zimmermann NE, Pellissier L, Kissling WD, Tobias JA, Schleuning M, Graham CH. Global plant-frugivore trait matching is shaped by climate and biogeographic history. Ecol Lett. 2022;25(3):686–96.
- <span id="page-10-5"></span>10. Bender IMA, Kissling WD, Blendinger PG, Böhning-Gaese K, Hensen I, Kühn I, Muñoz MC, Neuschulz EL, Nowak L, Quitián M, Saavedra F, Santillán V, Töpfer T, Wiegand T, Dehling DM, Schleuning M. Morphological trait matching shapes plant-frugivore networks across the Andes. Ecography. 2018;41(11):1910–9.
- <span id="page-10-6"></span>11. Nevo O, Razafimandimby D, Antonio J, Jeffrey J, Schulz S, Ayasse M. Fruit scent as an evolved signal to primate seed dispersal. Sci Adv. 2018;4:1–7.
- <span id="page-10-7"></span>12. Nevo O, Valenta K, Razafimandimby D, Melin AD, Ayasse M, Chapman CA. Frugivores and the evolution of fruit colour. Biol Lett. 2018;14:1–4.
- <span id="page-10-8"></span>13. Brodie JF. Evolutionary cascades induced by large frugivores. PNAS. 2017;114:11998–2002.
- <span id="page-10-9"></span>14. Valenta K, Bhramdat HD, Calhoun GV, Daegling DJ, Nevo O. Variation in ripe fruit hardness: a mechanical constraint? Oikos. 2022;2:1–7.
- <span id="page-10-10"></span>15. Lei B, Cui J, Newman C, Buesching CD, Xie Z, MacDonald DW, Zhou Y. (2021). Seed dispersers shape the pulp nutrients of fleshy-fruited plants. *Proceedings of the Royal Society B: Biological Sciences*, *288*, 1–10.
- <span id="page-10-11"></span>16. Moretti C, Mattos L, Calbo A, Sargent S. Climate changes and potential impacts on postharvest quality of fruit and vegetable crops: a review. Food Res Int. 2010;43:1824–32.
- <span id="page-10-12"></span>17. Stewart AL, Ahmed S. Effects of climate change on fruit nutrition. In: Srivastava AK, Hu C, editors. Fruit crops: diagnosis and management of nutrient constraints. Amsterdam, Netherlands: Elsevier; 2020. pp. 77–93.
- <span id="page-10-13"></span>18. Fischer G, Parra-Coronado A, Balaguera-López HE. Altitude as a determinant of fruit quality with emphasis on the Andean tropics of Colombia. A review. Agron Colombiana. 2022;40:70–85.
- 19. Nerd A, Gutman F, Mizrahi Y. Ripening and postharvest behaviour of fruits of two Hylocereus species (Cactaceae). Postharvest Biol Technol. 1999;17:39–45.
- <span id="page-10-14"></span>20. Welles GWH, Buitelaar K. Factors affecting soluble solids content of muskmelon (*Cucumis melo* L). Neth J Agric Sci. 1988;36:239–46.
- <span id="page-10-15"></span>21. Farré-Armengol G, Filella I, Llusià J, Niinemets Ü, Peñuelas J. Changes in floral bouquets from compound-specific responses to increasing temperatures. Glob Change Biol. 2014;20:3660–9.
- <span id="page-10-43"></span>22. Peñuelas J, Staudt M. BVOCs and global change. Trends Plant Sci. 2010;15:133–44.
- <span id="page-10-16"></span>23. Silvanini A, Dall'Asta C, Morrone L, Cirlini M, Beghè D, Fabbri A, Ganino T. Altitude effects on fruit morphology and flour composition of two chestnut cultivars. Sci Hort. 2014;176:311–8.
- <span id="page-10-17"></span>24. Koshita Y. Effect of temperature on fruit color development. In: Kanayama Y, Kochetov A, editors. Abiotic Stress Biology in Horticultural plants. Tokyo: Springer; 2015. pp. 47–58.
- <span id="page-10-18"></span>25. IPCC. In: Pörtner DC, Roberts M, Tignor ES, Poloczanska K, Mintenbeck A, Alegría M, Craig S, Langsdorf S, Löschke V, Möller A, Okem B, Rama, editors. Climate change 2022: impacts, adaptation, and vulnerability. Contribution of working group II to the sixth assessment report of the Intergovernmental Panel on Climate Change [H.-O. Cambridge University Press, Cambridge, UK and New York, NY, USA: Cambridge University Press; 2022.
- <span id="page-10-19"></span>26. IPCC. (2021): Climate Change 2021: The physical science basis. contribution of working group I to the sixth assessment report of the Intergovernmental Panel on Climate Change [Masson-Delmotte, V., P. Zhai, A. Pirani, S.L. Connors, C. Péan, S. Berger, N. Caud, Y. Chen, L. Goldfarb, M.I. Gomis, M. Huang, K. Leitzell, E. Lonnoy, J.B.R. Matthews, T.K. Maycock, T. Waterfield, O. Yelekçi, R. Yu, and B. Zhou, editors]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- <span id="page-10-20"></span>27. Jordano P, Bascompte J, Olesen JM. Invariant properties in coevolutionary networks of plant-animal interactions. Ecol Lett. 2003;6:69–81.
- <span id="page-10-21"></span>28. Nowak L, Schleuning M, Bender IMA, Böhning-Gaese K, Dehling DM, Fritz SA, Kissling WD, Mueller T, Neuschulz EL, Pigot AL, Sorensen MC, Donoso I. Avian seed dispersal may be insufficient for plants to track future temperature change on tropical mountains. Glob Ecol Biogeogr. 2022;31:848–60.
- <span id="page-10-22"></span>29. Raoelinjanakolona NN, Ramananjato V, Andrianarimisa A, Andrianiaina AF, Nantenaina RH, Razafindratsima OH. Fragile plant-frugivore interaction networks in tropical forest edges. Biol Conserv. 2023;277:1–11.
- 30. Acevedo-Quintero JF, Saldaña-Vázquez RA, Mendoza E, Zamora-Abrego JG. Sampling bias affects the relationship between structural importance and species body mass in frugivore-plant interaction networks. Ecol Complex. 2020;44:1–8.
- 31. Donoso I, Schleuning M, García D, Fründ J. Defaunation effects on plant recruitment depend on size matching and size trade-offs in seed-dispersal networks. Proc Royal Soc B: Biol Sci. 2017;284(1855):20162664.
- 32. Bello C, Galetti M, Pizo MA, Magnago LFS, Rocha MF, De Lima RaF, Peres CA, Ovaskainen O, Jordano P. (2015). Defaunation affects carbon storage in tropical forests. *Science Advances*, *1*.
- <span id="page-10-23"></span>33. Traveset R, Heleno M, Nogales. The ecology of seed dispersalR.S. In: Gallagher, editor. Seeds: the Ecology of Regeneration in Plant communities. Wallingford, Oxfordshire: CAB International; 2014. pp. 62–93.
- <span id="page-10-24"></span>34. Herrera CM. In: Herrera CM, Pellmyr O, editors. Seed dispersal by vertebrates. Blackwell Scientific; 2002. pp. 185–208.
- <span id="page-10-25"></span>35. Peñuelas J. An increasingly scented world. New Phytol. 2008;180:735–8.
- <span id="page-10-26"></span>36. Nevo O, Valenta K, Helman A, Ganzhorn JU, Ayasse M. Fruit scent as an honest signal for fruit quality. BMC Ecol Evol. 2022;22:1–10.
- <span id="page-10-27"></span>37. Schaefer HM, Valido A, Jordano P. (2014). Birds see the true colours of fruits to live off the fat of the land. Proceedings of the Royal Society B: Biological Sciences, 281(1777).
- <span id="page-10-42"></span>38. Albert-Daviaud A, Perillo S, Stuppy W. Seed dispersal syndromes in the madagascan flora: the unusual importance of primates. Oryx. 2018;52:418–26.
- <span id="page-10-28"></span>39. Barnett JR, Tieman DM, Caicedo AL. (2023). Variation in ripe fruit volatiles across the tomato clade: an evolutionary framework for studying fruit scent diversity in a crop wild relative. Am J Bot, *110*(9).
- <span id="page-10-29"></span>40. Valenta K, Miller CN, Monckton SK, Melin AD, Lehman SM, Styler SA, Jackson DA, Chapman CA, Lawes MJ. Fruit ripening signals and cues in a madagascan dry forest: haptic indicators reliably indicate fruit ripeness to dichromatic Lemurs. Evol Biol. 2016;43:344–55.
- <span id="page-10-30"></span>41. Lord JM. Frugivore gape size and the evolution of fruit size and shape in southern hemisphere floras. Austral Ecol. 2004;29(4):430–6.
- <span id="page-10-34"></span>42. Bufalo FS, Galetti M, Culot L. Seed dispersal by primates and implications for the conservation of a biodiversity hotspot, the Atlantic Forest of South America. Int J Primatol. 2016;37:333–49.
- <span id="page-10-31"></span>43. Galetti M, Guevara R, Côrtes MC, Fadini R, Von Matter S, Leite AB, Labecca F, Ribeiro T, Carvalho CS, Collevatti RG, Pires MM, Guimarães PR, Brancalion PH, Ribeiro MC, Jordano P. Functional extinction of birds drives rapid evolutionary changes in seed size. Science. 2013;340:1086–90.
- <span id="page-10-32"></span>44. Schupp EW, Jordano P, Gómez JM. Seed dispersal effectiveness revisited: a conceptual review. New Phytol. 2010;188(2):333–53.
- <span id="page-10-33"></span>45. Nevo O, Filla C, Valenta K, Schupp EW. (2023). What drives seed dispersal effectiveness? Ecol Evol, *13*(9).
- <span id="page-10-35"></span>46. Qi W, Guo S, Chen X, Cornelissen JHC, Bu H, Du G, Cui X, Li W, Liu K. Disentangling ecological, allometric and evolutionary determinants of the relationship between seed mass and elevation: insights from multiple analyses of 1355 angiosperm species on the eastern tibetan Plateau. Oikos. 2013;123:23–32.
- <span id="page-10-36"></span>47. Maes SL, Perring MP, Depauw L, Bernhardt-Römermann M, Blondeel H, Brūmelis G, Brunet J, Decocq G, Ouden JD, Govaert S, Härdtle W, Hédl R, Heinken T, Heinrichs S, Hertzog LR, Jaroszewicz B, Kirby KJ, Kopecký M, Landuyt D, Verheyen K. Plant functional trait response to environmental drivers across European temperate forest understorey communities. Plant Biol. 2020;22:410–24.
- <span id="page-10-37"></span>48. Trethowan LA, Jennings L, Bramley GLC, Clark RP, Dawson S, Moore A, Pearce L, Puglisi C, Riwu-Kaho M, Rosaria, Trias‐Blasi A, Utteridge TMA, Rustiami H. (2022). Environment‐dependent influence of fruit size upon the distribution of the malesian archipelagic flora. Plants People Planet, 712–21.
- <span id="page-10-38"></span>49. Keller M. Managing grapevines to optimise fruit development in a challenging environment: a climate change primer for viticulturists. Aust J Grape Wine Res. 2010;16:56–69.
- <span id="page-10-39"></span>50. Zhang C, Yi H, Gao X, Bai T, Ni Z, Chen Y, Wang M, Zhang Y, Pan J, Yu W, Xie D. Effect of different altitudes on morpho-physiological attributes associated with mango quality. Diversity. 2022;14:1–11.
- <span id="page-10-40"></span>51. Maluf RP, Alzate-Marin AL, Silva CC, Pansarin LM, Bonifácio-Anacleto F, Schuster I, de Mello Prado R, Martinez CA. Warming and soil water availability affect plant–flower visitor interactions for Stylosanthes capitata, a tropical forage legume. Sci Total Environ. 2022;817:1–10.
- <span id="page-10-41"></span>52. Descamps C, Jambrek A, Quinet M, Jacquemart AL. Warm temperatures reduce flower attractiveness and bumblebee foraging. Insects. 2021;12:1–13.
- <span id="page-11-0"></span>53. Nowak L, Kissling WD, Bender IMA, Dehling DM, Töpfer T, Böhning-Gaese K, Schleuning M. Projecting consequences of global warming for the functional diversity of fleshy-fruited plants and frugivorous birds along a tropical elevational gradient. Divers Distrib. 2019;25:1362–74.
- <span id="page-11-1"></span>54. Bründl AC, Sallé L, Lejeune LA, Sorato E, Thiney AC, Chaine AS, Russell AF. Elevational gradients as a model for understanding associations among temperature, breeding phenology and success. Front Ecol Evol. 2020;8:1–14.
- <span id="page-11-2"></span>55. Sundqvist MK, Sanders NJ, Wardle DA. Community and ecosystem responses to elevational gradients: processes, mechanisms, and insights for global change. Annu Rev Ecol Evol Syst. 2013;44(1):261–80.
- 56. Spitzer CM, Sundqvist MK, Wardle DA, Gundale MJ, Kardol P. (2022). Root trait variation along a sub-arctic tundra elevational gradient. Oikos, *2023*(1).
- 57. Lasmar CJ, Rosa D, De Queiroz CA, Nunes ACM, Imata CA, Alves MM, Nascimento GP, De Ázara GB, Vieira LN, Louzada L, Feitosa J, Brescovit RM, Passamani AD, M., Ribas CR. Temperature and productivity distinctly affect the species richness of ectothermic and endothermic multitrophic guilds along a tropical elevational gradient. Oecologia. 2021;197(1):243–57.
- <span id="page-11-3"></span>58. Corcos D, Cerretti P, Mei M, Taglianti AV, Paniccia D, Santoiemma G, De Biase A, Marini L. Predator and parasitoid insects along elevational gradients: role of temperature and habitat diversity. Oecologia. 2018;188(1):193–202.
- <span id="page-11-4"></span>59. Corlett RT. (2014): The impacts of climate change in the tropics. State Tropics. 155–61.
- <span id="page-11-5"></span>60. Torppa KA, Wirta H, Hanski I. Unexpectedly diverse forest dung beetle communities in degraded rain forest landscapes in Madagascar. Biotropica. 2020;52:351–65.
- <span id="page-11-6"></span>61. Ganzhorn JU, Lowry PP, Schatz GE, Sommer S. The biodiversity of Madagascar: one of the world's hottest hotspots on its way out. Oryx. 2001;35:346–8.
- <span id="page-11-7"></span>62. Bollen A, Van Elsacker L, Ganzhorn JU. Tree dispersal strategies in the littoral forest of Sainte Luce (SE-Madagascar). Oecologia. 2004;139:604–16.
- <span id="page-11-8"></span>63. Nevo O, Ayasse M. Fruit Scent: Biochemistry, ecological function, and evolution. In: Mérillon JM, Ramawat K, editors. Co-evolution of secondary metabolites. Reference Series in Phytochemistry. Cham: Springer; 2020.
- <span id="page-11-9"></span>64. Nevo O, Razafimandimby D, Valenta K, Jeffrey JAJ, Reisdorff C, Chapman CA, Ganzhorn JU, Ayasse M. Signal and reward in wild fleshy fruits: does fruit scent predict nutrient content? Ecol Evol. 2019;9:10534–43.
- <span id="page-11-10"></span>65. Valenta K, Burke RJ, Styler SA, Jackson DA, Melin AD, Lehman SM. (2013). Colour and odour drive fruit selection and seed dispersal by mouse Lemurs. Sci Rep, *3*.
- <span id="page-11-11"></span>66. Wright PC, Razafindratsita VR, Pochron ST, Jernvall J. The key to Madagascar frugivores. Tropical fruits and frugivores: the search for strong interactors. Springer Netherlands; 2005. pp. 121–38.
- <span id="page-11-12"></span>67. Valenta K, Daegling DJ, Nevo O, Ledogar J, Sarkar D, Kalbitzer U, Bortolamiol S, Omeja P, Chapman CA, Ayasse M, Kay R, Williams B. Fruit Selectivity in Anthropoid Primates: size matters. Int J Primatol. 2020;41:525–37.
- <span id="page-11-13"></span>68. Dudley R. Ethanol, fruit ripening, and the historical origins of human alcoholism in primate frugivory. Integr Comp Biol. 2004;44(4):315–23.
- 69. Nevo O, Schmitt MH, Ayasse M, Valenta K. Sweet tooth: elephants detect fruit sugar levels based on scent alone. Ecol Evol. 2020;10(20):11399–407.
- <span id="page-11-14"></span>70. Sánchez F, Korine C, Steeghs M, Laarhoven LJ, Cristescu SM, Harren FJM, Dudley R, Pinshow B. Ethanol and methanol as possible odor cues for Egyptian fruit bats (Rousettus aegyptiacus). J Chem Ecol. 2006;32(6):1289–300.
- <span id="page-11-15"></span>71. Wright P, Johnson. S. *Unpublished data*
- <span id="page-11-16"></span>72. Batist CH, Razafindraibe MN, Randriamanantena F, Baden AL. Factors affecting call usage in wild black-and-white ruffed Lemurs (*Varecia variegata*) at Mangevo, Ranomafana National Park. Primates. 2022;63:79–91.
- 73. Campera M, Santini L, Balestri M, Nekaris KAI, Donati G. (2020). Elevation gradients of lemur abundance emphasise the importance of Madagascar's lowland rainforest for the conservation of endemic taxa. In Mammal Review (Vol. 50, Issue 1, pp. 25–37). Blackwell Publishing Ltd.
- 74. Lees DC. A null model for species richness gradients: bounded range overlap of butterflies and other rainforest endemics in Madagascar LANTO ANDRIA-MAMPIANINA. Biol J Linn Soc. 1999;67:529–84.
- <span id="page-11-17"></span>Pauly A, Brooks RW, Nilsson A, Apesenko Y, Eardley CD, Terzo M, Griswold T, Schwarz M, Patiny S, Munzinger J, Barbier Y. (2001). Hymenoptera Apoidea de Madagascar et des Iles Voisines, *286*. Musee Royal de l'Afrique Centrale Tervuren, Belgique.
- <span id="page-11-18"></span>76. Bergman TJ, Beehner JC. A simple method for measuring colour in wild animals: validation and use on chest patch colour in geladas (*Theropithecus gelada)*. Biol J Linn Soc. 2008;94:231–40.
- <span id="page-11-19"></span>77. Potash AD, Greene DU, Foursa GA, Mathis VL, Conner M, L., Mccleery RA. A comparison of animal color measurements using a commercially available digital color sensor and photograph analysis. Curr Zool. 2021;66:601–6.
- <span id="page-11-20"></span>78. Troscianko J, Stevens M. Image calibration and analysis toolbox - a free software suite for objectively measuring reflectance, colour and pattern. Methods Ecol Evol. 2015;6:1320–31.
- <span id="page-11-21"></span>79. Boratyński Z, Brito JC, Campos JC, Karala M, Mappes T. Large spatial scale of the phenotype-environment color matching in two cryptic species of African desert jerboas (Dipodidae: Jaculus). PLoS ONE. 2014;9:1–9.
- <span id="page-11-22"></span>80. Schneider CA, Rasband WS, Eliceiri KW. NIH image to ImageJ: 25 years of image analysis HHS public access. Nat Methods. 2012;9:671–5.
- <span id="page-11-23"></span>81. De Pérez M, Ojeda M, Giménez A, González M, Hernández A. Atributos De calidad en frutos de fresa 'Capitola' cosechados en diferentes condiciones climáticas en Venezuela. Bioagro. 2017;29:163–74.
- <span id="page-11-24"></span>82. Karagiannis E, Michailidis M, Tanou G, Scossa F, Sarrou E, Stamatakis G, Samiotaki M, Martens S, Fernie AR, Molassiotis A. Decoding altitude-activated regulatory mechanisms occurring during apple peel ripening. Hortic Res. 2020;7:1–12.
- <span id="page-11-25"></span>83. Li Y, Sun H, Li J, Qin S, Yang W, Ma X, Qiao X, Yang B. Effects of genetic background and altitude on sugars, malic acid and ascorbic acid in fruits of wild and cultivated apples (Malus Sp). Foods. 2021;10:1–20.
- <span id="page-11-26"></span>84. Bates D, Mächler M, Bolker BM, Walker SC. Fitting linear mixed-effects models using lme4. J Stat Softw. 2015;67:3–48.
- <span id="page-11-27"></span>85. Brat P, Yahia A, Chillet M, Bugaud C, Bakry F, Reynesa M, Brilloueta JM. Influence Du Cltivar, de l'altitude de la bananeraie et du stade de maturité sur la composition en composés volatils de la banane. Fruits. 2004;59:75–82.
- <span id="page-11-28"></span>86. Cheng S, Fu X, Mei X, Zhou Y, Du B, Watanabe N, Yang Z. Regulation of biosynthesis and emission of volatile phenylpropanoids/benzenoids in petunia× hybrida flowers by multi-factors of circadian clock, light, and temperature. Plant Physiol Biochem. 2016;107:1–8.
- <span id="page-11-29"></span>87. Nevo O, Heymann EW, Schulz S, Ayasse M. Fruit odor as a ripeness signal for seed-dispersing primates? A case study on four neotropical plant species. J Chem Ecol. 2016;42:323–8.
- <span id="page-11-30"></span>88. Schaefer HM, Schmidt V, Winkler H. Testing the defence trade-off hypothesis: how contents of nutrients and secondary compounds affect fruit removal. Oikos. 2003;102:318–28.
- <span id="page-11-31"></span>89. Kato M, Ikoma Y, Matsumoto H, Sugiura M, Hyodo H, Yano M. Accumulation of carotenoids and expression of carotenoid biosynthetic genes during maturation in citrus fruit. Plant Physiol. 2004;134:824–37.
- <span id="page-11-32"></span>90. He F, Mu L, Yan G, Liang N, Pan Q, Wang J, Reeves MJ, Duan C. Biosynthesis of anthocyanins and their regulation in Colored grapes. Molecules. 2010;15(12):9057–91.
- <span id="page-11-37"></span>91. Xie X, Li S, Zhang R, Zhao J, Chen Y, Zhao Q, Yao Y, You C, Zhang X, Hao Y. The bHLH transcription factor MdbHLH3 promotes anthocyanin accumulation and fruit colouration in response to low temperature in apples. Plant Cell Environ. 2012;35(11):1884–97.
- <span id="page-11-33"></span>92. Zoratti L, Jaakola L, Häggman H, Giongo L. Anthocyanin Profile in berries of Wild and Cultivated *Vaccinium* spp. along Altitudinal gradients in the Alps. J Agric Food Chem. 2015;63:8641–50.
- <span id="page-11-34"></span>93. IFRC OCHA, Climate Centre. &. (2022). Extreme heat, preparing for the heatwaves of the future [OCHA, editor].
- <span id="page-11-35"></span>94. Valenta K, Kalbitzer U, Razafimandimby D, Omeja P, Ayasse M, Chapman CA, Nevo O. The evolution of fruit colour: phylogeny, abiotic factors and the role of mutualists. Sci Rep. 2018;8:14302.
- <span id="page-11-36"></span>95. Mansour G, Ghanem C, Mercenaro L, Nassif N, Hassoun G, del Caro A. Effects of altitude on the chemical composition of grapes and wine: a review. Oeno One. 2022;56:227–39.
- <span id="page-11-38"></span>96. Zoratti L, Jaakola L, Häggman H, Giongo L. Modification of sunlight Radiation through Colored Photo-Selective nets affects Anthocyanin Profile in Vaccinium spp. Berries. PLoS ONE. 2015;10:1–17.
- <span id="page-11-39"></span>97. Mayorga M, Fischer G, Melgarejo. L, Parra-Coronado M., A. Growth, development and quality of *Passifora Tripartita* var. *Mollissima* fruits under two environmental tropical conditions. J Appl Bot Food Qual. 2020;93:66–75.
- 98. Kumar P, Sethi S, Sharma RR, Singh S, Saha S, Sharma VK, Sharma SK, Varghese E. Influence of altitudinal variation on the physical and biochemical characteristics of apple (Malus domestica). Indian J Agric Sci. 2019;89:145–52.
- <span id="page-11-40"></span>99. Parra-Coronado A, Fischer G, Camacho-Tamayo JH. Post-harvest quality of pineapple guava [*Acca sellowiana* (O. Berg) Burret] fruits produced in two locations of Cundinamarca, Colombia, at different altitudes. Agronomía Colombiana. 2018;36:68–78.
- <span id="page-11-41"></span>100. Luppichini M, Bini M, Barsanti M, Giannecchini R, Zanchetta G. Seasonal rainfall trends of a key Mediterranean area in relation to large-scale atmospheric circulation: how does current global change affect the rainfall regime? J Hydrol. 2022;612:128233.
- <span id="page-12-0"></span>101. Bony S, Stevens B, Frierson DMW, Jakob C, Kageyama M, Pincus R, Shepherd TG, Sherwood SC, Siebesma AP, Sobel AH, Watanabe M, Webb MJ. Clouds, circulation and climate sensitivity. Nat Geosci. 2015;8(4):261–8.
- <span id="page-12-1"></span>102. Stillman JH. Heat waves, the new normal: Summertime temperature extremes will impact animals, ecosystems, and human communities. Physiology. 2019;34:86–100.
- <span id="page-12-2"></span>103. Almagro A, Oliveira PTS, Nearing MA, Hagemann S. Projected climate change impacts in rainfall erosivity over Brazil. Sci Rep. 2017;7:1–12.
- 104. Sohoulande Djebou DC, Singh VP. Impact of climate change on precipitation patterns: a comparative approach. Int J Climatol. 2017;36:3588–606.
- <span id="page-12-3"></span>105. Pryor SC, Barthelmie RJ. Climate change impacts on wind energy: a review. Renewable and Sustainable Energy; 2010.

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