Epidemic of multiple *Treponema pallidum* strains in men who have sex with men in Japan: efficient multi-locus sequence typing scheme and indicator biomarkers



Wakana Sato^{1,2}, Ayako Sedohara³, Michiko Koga³, Yu Nakagama⁴, Hiroshi Yotsuyanagi^{2,3}, Yasutoshi Kido⁴ and Eisuke Adachi^{2*}

Abstract

Background The challenges in culturing Treponema pallidum have hindered molecular-biological analysis. This study aims to establish a molecular epidemiological analysis of syphilis among Japanese men who have sex with men (MSM) and to investigate the relationship between bacteremia and associated pathophysiology.

Methods We used whole blood specimens from syphilis-diagnosed individuals in Tokyo, collected between February 2019 and June 2022. All individuals were MSM, and most were people with HIV (97.2%). We used a multi-locus sequence typing (MLST) scheme for epidemiological analysis. Sequences for MLST (*TP0136*, *TP0548*, and *TP0705*) were obtained.

Results Out of 71 whole blood samples, 26 samples (36.6%) were positive for *TP0136*, and we sequenced three loci for MLST in 22 samples (31.0%). The most frequently detected sequence type (ST) was ST3 (n = 9), followed by ST6 (n = 6). Phylogenetic analysis revealed that 12 samples belonged to the SS14-like group (60%), and 8 samples belonged to the Nichols-like group (40%). *Treponema pallidum* subsp. *endemicum* (TEN), the cause of bejel was detected in three samples (12%). There was a signi cant association between *TP0136* detection rate and C- reactive protein (CRP) (77.0% at a cut-o :0.5 mg/dL).

Conclusion Both SS14-like and Nichols-like strains were circulating concurrently, and TEN could have been sexually transmitted among MSM with HIV. Elevated CRP may indicate the presence of the pathogen in the blood.

Keywords Multi-locus sequence typing, *Treponema pallidum subspecies pallidum*, Syphilis, *Treponema pallidum* subsp. *endemicum*, Bejel

*Correspondence: Eisuke Adachi e-adachi@ims.u-tokyo.ac.jp ¹Department of Laboratory Medicine, IMSUT Hospital of The Institute of Medical Science, The University of Tokyo, Tokyo, Japan ²Department of Infectious Diseases and Applied Immunology, IMSUT Hospital of The Institute of Medical Science, The University of Tokyo, 4-6-1 Shirokanedai, Minato-ku, Tokyo 108-8639, Japan
 ³Division of Infectious Diseases, Advanced Clinical Research Center, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan
 ⁴Department of Virology and Parasitology, Graduate School of Medicine, Osaka Metropolitan University, Osaka, Japan



[®] The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the articles Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the articles Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

Introduction

Treponema pallidum (T.pallidum) subsp. pallidum (TPA), the causative agent of syphilis, has long been known to be prevalent among men who have sex with men (MSM). Currently, the number of new cases of syphilis is estimated to be 6 million worldwide [1]. In Japan, in particular, heterosexual transmission among young women has increased signi cantly in recent years [2]. TPA poses a challenge for clinical use due to the difculty in culturing it. Edmondson et al. reported successful long-term culture of TPA using the rabbit epithelial cell co-cultivation system; however, this system has not been readily available in hospital laboratories worldwide [3]. Endemic treponemas (yaws, bejel, and pinta), classi ed as neglected tropical diseases (NTDs) and recognized as non-sexually transmitted syphilis, are caused by microbes T. pallidum subsp. pertenue (TPE), T. pallidum subsp. endemicum (TEN), and T. pallidum subsp. ese are genetically highly similar to TPA carateum. and cannot be di erentiated from it using the serological diagnostic methods. While it is possible that endemic treponemas are sexually transmitted and lead to epidemics, investigating this requires establishing a genetic diagnostic method capable of distinguishing them.

e diagnosis of syphilis involves both treponemal and nontreponemal tests [4]. e nontreponemal test measures antibodies against lipoidal material released by damaged host cells, such as cardiolipin; the nontreponemal test assesses the e ect of treatment [5]. Although clinical diagnostic methods utilizing direct detection targets like *polA* and *tpp47* have been proposed [6, 7], these methods became less common due to the di culty to culture TPA [3]. Moreover, molecular epidemiological analysis faced similar challenges. Samples for molecular epidemiological analysis are limited. Chancre specimens are suitable for molecular epidemiological analysis, but patients with primary syphilis who present with chancres represent only a small fraction of those with syphilis encountered in the typical outpatient setting for MSM. On the other hand, whole blood is less sensitive, and whole-genome sequence (WGS) analysis has been difere is a need for molecular epidemiological cult [8]. analysis methods that can be performed with easily collected blood samples, enabling phylogenetic analysis with su cient discriminatory power. Multi-locus sequence typing (MLST) has been proposed for molecular epidemiological analysis of TPA. Grillová et al. developed a new MLST with high resolution power using three loci: TP0136, TP0548, TP0705, achieving enough discriminatory power to distinguish TPA from other treponemal subspecies and di erentiate between the two TPA clades SS14 and Nichols [9]. Although whole genome sequencing has recently been an alternative option for molecular epidemiology, simpler MLST has more advantages in terms of cost, a subset of samples, and a growing public treponemal MLST database for storing and analyzing typing data.

Ultimately, challenges in molecular epidemiological studies, genetic diagnosis, and the establishment of markers other than antibody titers in syphilis, where the pathogen presents signi cant challenges to culture, arise from the di culty in collecting specimens containing genomic information. To address this, we explored the feasibility of utilizing blood samples, which can be collected from all patients with syphilis, including those with asymptomatic early-stage syphilis. is study overcomes technical challenges by typing multiple genetic polymorphisms of T. pallidum using the MLST method and utilizing blood samples. Our aim is to identify the prevalent T. pallidum species in Japanese MSM with HIV, determine factors responsible for changes in the infected population, and prevent the spread of infection.

Materials and methods

Clinical samples

Clinical samples were collected from the individuals with syphilis diagnosed by clinical symptoms and serological tests in IMSUT hospital, the Institute of Medical Science,

e University of Tokyo between February 2019 to June 2022. All cases were above 10 U/mL of TPLA and 1.0 RU of RPR. We reviewed routinely collected clinical records of the patients, including gender, age, HIV status, and the stage of syphilis, along with the results of serological tests. Ethics approval was granted by the ethics board of the Institute of Medical Science, University of Tokyo (2020-79-0324). Written consent was obtained from all participants.

DNA extraction, puri cation, and nested PCR

DNA was extracted from whole blood samples of participants using QIAamp DNA Mini Kit (Qiagen, Germany) following the manufacture's protocol. e elution (100 μ l) was concentrated to a 20 μ l solution using Ethachinmate (Nippon gene, Japan).

PCR primers as described previously by Grillová et al. [9], were employed. Optimal PCR conditions were veried using synthetic DNA (Azenta). Modi ed PCR procedure was used as below: For the rst PCR, the total volume was 50 μ l, with each reaction consisting of 15 μ l of the concentrated template solution, 25 μ l of KOD one PCR master mix (TOYOBO, Japan), and 1 μ l of each primer (10 μ M). e rst PCR conditions were as follows: 95 °C for 2 min, followed by 5 cycles at 98 °C for 10 s, 56 °C for 10 s, 5 cycles at 98 °C for 10 s, 5 cycles at 98 °C for 10 s, 5 cycles at 98 °C for 10 s, 50 °C for 10 s

KOD one PCR master mix (TOYOBO, Japan), and 0.4 μ l of each primer. e second PCR conditions were: 95 °C for 2 min, followed by 5 cycles at 98 °C for 10 s, 63 °C for 10 s, 63 °C for 10 s, 5 cycles at 98 °C for 10 s, 60 °C for 10 s, and nally, 68 °C for 5 min.

In case of *TP0705* and *TP0548* PCR negative despite *TP0136* PCR positive, direct pellet PCR was performed increase the sensitivity of PCR detection. e elution (50 μ l) was concentrated using Ethachinmate, and the pellet was dissolved directly in PCR ingredients. e solutions were incubated for 10 min at room temperature. PCR products were puri ed using the QIAquick puri cation kit (Qiagen, Germany) following the manufacturer's protocol.

Multi-locus sequence typing and phylogenetic analysis

e puri cation products were used to determine the sequence by Sanger sequencing. MLST was performed for three loci, TP0136, TP0548 and TP0705. e sequences were uploaded to PubMLST BIGSdb of TPA for decision of allele number and Sequence Type e phylogenetic tree was constructed using (ST) [10]. MEGA X with the Maximum Likelihood method and bootstrap test [11]. e evolutionary history was determined employing the Maximum Likelihood method and the Tamura-Nei model [12]. e percentage of trees where the associated taxa clustered together is presented next to the branches. Initial trees for the heuristic search were automatically generated by applying Neighbor-Join and BioNJ algorithms to a matrix of pairwise distances estimated through the Maximum Composite Likelihood (MCL) approach. e topology with the highest log-likelihood value was then selected. is analysis comprised 9 nucleotide sequences, resulting in a total of 2577 positions in the nal dataset. e concatenated sequences were downloaded from PubMLST BIGSdb.

Investigation of biomarkers associated with the *TP0136* detection

We initially tested for *TP0136*, and the PCR-positive samples were subsequently tested for TP0548 and TP0705. Percentage of samples sequenced as *TP0136* (i.e., *TP0136* detection rate) was examined in relation to the following biomarkers: *T. pallidum* latex agglutination (TPLA), rapid plasma regain (RPR), and C-reactive protein (CRP). e tests were performed using RAPIDIA Auto TP (FUJIREBIO Inc., Japan), LASAY Auto RPR (SHIMA Laboratories Co., Ltd, Japan), and N-assay LA CRP-T Nittobo (Nittobo Medical Co., Ltd., Japan) on LABOSPECT 006 (Hitachi Hi-Tech Cp., Ltd., Japan),

Table 1 Clinical characteristics of cases with syphilis

		All cases		Cases that were not accessible for MLST		Fully-typed cases by MLST		<i>P</i> value
Number of cases		71		51		20		
Median age, yr (IQR)		43	(37–49)	41	(34–48)	44	(37–50)	0.37 [†]
HIV infection, n (%)		69	(97.2)	51	(100)	18	(90.0)	0.08 [‡]
Stage of syphilis, n (%)	Primary syphilis	7	(9.9)	3	(7.8)	4	(20)	0.09 [‡]
	Secondary syphilis	23	(32)	15	(29)	8	(40)	0.41 [‡]
	Early Asymptomatic syphilis	35	(49)	28	(54)	7	(35)	0.19 [‡]
	Neurosyphilis [*]	6	(8.5)	3	(5.9)	3	(15)	0.34 [‡]
	Unknown	6	(8.5)	5	(9.8)	1	(5%)	0.67 [‡]
Serum TPLA titer, Median, U/mL (IQR)		6571	(2592– 15857)	5351	(2325– 10774)	14,483	(4054– 24596)	0.018 [†]
Serum RPR titer, Median, R.U. (IQR)		75.7	(34.4 -188.5)	60.9	(27.8 -136.5)	188.7	(59.8 -337.2)	0.0081
CRP, mg/dL (IQR)		0.23	(0.09-0.65)	0.14	(0.065-0.33)	1.04	(0.61–1.36)	< 0.0001

MLST, multi-locus sequence typing; TPLA, Treponema pallidum latex agglutination; RPR, Rapid Plasma Reagin; CRP, C-reactive protein * All symptoms were indicative of secondary syphilis and were categorized as such [†]Mann-Whitney test, [‡]Fisher's exact test

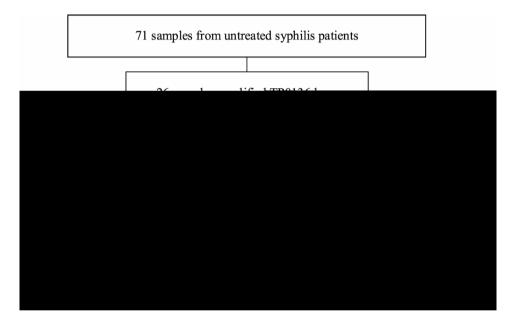


Fig. 1 Flowchart of the molecular epidemiology analysis of *Treponema pallidum*. TPA, *Treponema pallidum* subsp. *pallidum*; TEN, *Treponema pallidum* subsp. *endemicum*; MLST, multi-locus sequence typing

(28.2%), and a partial prole for 4 samples. In this study, ST3 (1.1.8) (n=9) was predominant, followed by ST6 (3.2.3) (n=6). Additionally, we identi ed ST60 (9.2.3), ST26 (9.7.3), ST1 (1.3.1), and ST24 (6.1.8). For sample No. 21,100, a new allele was detected on *TP0136* (*TP0136-37*, BIGSdb_20230921112653_3477711_09755) and we applied for a new ST (ST120 [37.3.1]), which was related to ST1(1.3.1) and di ered by one substitution. We constructed the phylogenetic tree using the concatenated sequence we detected (Fig. 2). Twelve samples belonged to the SS14-like lineage group (60%), and eight samples belonged to the Nichols-like lineage group (40%). ese results indicated that both SS14-like and Nichols-like

strains were circulating concurrently in Japanese MSM-PWH. Partial pro les of TPA were obtained for the remaining three cases in Table 2.

Two of the 22 samples, in which all three alleles could be sequenced, and one in which only two alleles (*TP0136* and *TP0548*) could be sequenced, could not have the allele numbers determined by MLST. ese allelic proles exhibited unique nucleotide sequences at the typing loci and we referred to BLAST (https://blast.ncbi. nlm.nih.gov/Blast.cgi), and there was a possibility that these three samples (12%, 3/26) were TEN. e reference strain reported from Japan (Japan326e, Gen-Bank: CP073518.1:158259–159364) was used for the

 Table 2
 Multi-locus sequence typing pro les of Treponema pallidum subsp. pallidum

Sample No.	. TP0136 TP0548 TP0705		TP0705	ST	clonal complex		
1959	9	2	3	60	Nichols-like		
1974	3	2	3	6	Nichols-like		
2083	9	7	3	26	Nichols-like		
2085	1	3	1	1	SS14-like		
2189	3	2	3	6	Nichols-like		
2192	1	1	8	3	SS14-like		
2193	1	1	8	3	SS14-like		
2194	1	1	8	3	SS14-like		
2198	3	2	3	6	Nichols-like		
21,100	37	3	1	120	SS14-like		
21,108	1	1	8	3	SS14-like		
21,109	1	1	8	3	SS14-like		
21,111	1	1	8	3	SS14-like		
21,117	1	1	8	3	SS14-like		
21,120	3	*	*		Nichols-like [†]		
21,121	3	2	3	6	Nichols-like		
21,122	3	2	3	6	Nichols-like		
22,126	1	1	8	3	SS14-like		
22,129	1	*	*		SS14-like [†]		
22,133	3	2	3	6	Nichols-like		
22,138	3	2	*		Nichols-like [†]		
22,144	6	1	8	24	SS14-like		
22,147	1	1	8	3	SS14-like		

ST, sequence type, *Sequencing of this allele failed. [†] Classification is based on the sequence type for TP0136 only, not the multi-locus sequence typing profile

identi cation of TEN [13]. In one case, where all three alleles were successfully sequenced, both *TP0136* and *TP0548* had 100% query cover and 100% identity, with *TP0705* identi ed as allele number 9. In another case, where all three alleles were sequenced, *TP0136* showed

100% query cover and 100% identity, while TP0548 had 99.91% identity due to a single nucleotide insertion between reference positions 158,578 and 158,579, with *TP0705* again identi ed as allele number 9. is reference strain also identi ed TP0705 as allele number 9. In a third case, where only two alleles were sequenced, both *TP0136* and *TP0548* had 100% query cover and 100% identity. None of the three cases had visited tropical regions where bejel is prevalent. Of the three cases with suspected TEN infection, one was diagnosed with secondary syphilis, and the others with early asymptomatic syphilis.

Investigation of biomarkers and stage of syphilis associated with the *TP0136* detection rate

We examined the factors in uencing the detection rate of TPA-derived DNA in whole blood samples (refer to Additional Table for clinical data on *TP0136*-detected samples). In Table 3, no signi cant association between syphilis stage and *TP0136* detection was observed. However, TP0136 detection rates were 50% (15/30) and 27% (11/41) for symptomatic and asymptomatic syphilis, which includes early asymptomatic syphilis and cases with an unknown stage, respectively (P=0.052).

e RPR, TPLA, and CRP levels were all signi cantly higher in the *TP0136* detected group compared to the *TP0136* not detected group. Meanwhile, the *TP0136* detection rate and TPLA did not show a statistically signi cant correlation with cut-o s of 1000 U/mL (p=0.25) or 5000 U/mL (p=0.070). e Cochran-Armitage test for trend also failed to reveal an increase in *TP0136* detection as TPLA levels increased (p=0.056, Table 3). Similarly, the *TP0136* detection rate did not exhibit a

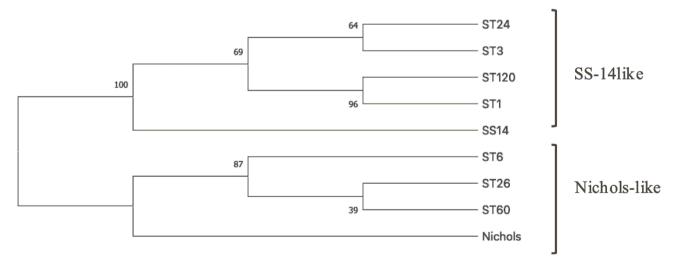


Fig. 2 Phylogenetic tree of the identi ed sequence types from multi-locus sequence typing. The evolutionary history was elucidated using the Maximum Likelihood method. The percentage of trees in which the associated taxa clustered together is presented next to the branches. Initial trees for the heuristic search were automatically generated by applying Neighbor-Join and BioNJ algorithms to a matrix of pairwise distances estimated through the Maximum Composite Likelihood approach

Table 3 Characteristics of cases that TP0136 was detected

		All cases		TP0136	TP0136 not detected		TP0136 detected	
Number of cases		71		45		26		
Stage of syphilis, n (%)	Primary syphilis	7	(9.9)	3	(6.6)	4	(15)	0.41 [‡]
	Secondary syphilis	23	(32)	12	(27)	11	(42)	0.19
	Early asymptomatic syphilis	35	(49)	26	(58)	9	(35)	0.085 [‡]
	Neurosyphilis [*]	6	(8.5)	2	(4.4)	4	(15)	0.18 [‡]
	Unknown	6	(8.5)	4	(8.9)	2	(7.8)	0.99 [‡]
Serum TPLA titer,	Median, U/mL (IQR)	6571	(2582–15857)	5209	(2042–9858)	11,262	(4527–23081)	0.0071 [†]
	< 1000 U/mL, n	7		6		1		0.056 [§]
	1000–5000 U/mL, n	23		16		6		
	> 5000 U/mL, n	41		23		19		
Serum RPR titer	Median, RU (IQR)	75.7	(34.4 -188.5)	66.9	(27.3–136)	115	(52.5–317)	0.016 [†]
	< 10 RU, n	6		5		1		0.12 [§]
	10–100 RU, n	36		25		12		
	> 100 RU, n	29		15		13		
CRP	Median, mg/mL (IQR)	0.23	(0.09-0.65)	0.13	(0.06-0.31)	0.865	(0.575–1.29)	< 0.0001 [†]
	< 0.1 mg/dL, n	19		18		1		< 0.0001 [§]
	0.1–0.5 mg/dL, n	26		21		5		
	> 0.5 mg/dL, n	26		6		20		

TPLA, Treponema pallidum latex agglutination; RPR, Rapid Plasma Reagin; CRP, C-reactive protein * All symptoms were indicative of secondary syphilis and were categorized as such †Mann-Whitney test, ‡Fisher's exact test, §Cochran-Armitage test for trend

signi cant association with the RPR cut-o s of 10 RU (p=0.40) or 100 RU (p=0.21). e trend test did not show an increase in *TP0136* detected as the RPR increased (p=0.12, Table 3).

In contrast, positive CRP levels were signi cantly associated with *TP0136* detection at cut-o s of 0.1 mg/dL (p=0.0007) or 0.5 mg/dL (p<0.0001). For CRP \ge 0.5 mg/ dL, 77.0% were positive for *TP0136*. A trend test further revealed that *TP0136* detection signi cantly increased as CRP levels increased (p<0.0001).

Discussion

Two distinct phylogenetic lineages of TPA strains, namely Nichols-like and SS14-like, have been identied [14]. Upon reviewing previously analyzed data, it was observed that the SS14-like lineage predominated among the samples obtained in Japan (refer to PubMLST BIGSdb). A previous study, utilizing Sequence-Based Molecular Typing (SBMT) other than the MLST method, reported greater genetic diversity of T. pallidum in the MSM population compared to the heterosexual population. Kojima et al. found that in MSM, 63% were SS14like, 37% were Nichols-like, whereas in heterosexuals, 100% were SS14-like [15]. Similarly, Kanai et al. reported that in MSM, 75% were SS14-like, 25% were Nichols-like, and 25% were macrolide-resistant, while 100% of the heterosexuals were SS14-like [16]. Given that the predominant demographic in our study comprised MSM, our MLST analysis generally aligns with ndings from earlier studies that employed di erent typing methods. Furthermore, WGS analysis revealed a close relationship between SS14-lineage strains in Japan and China [17]. In our study, ST3 (1.1.8) was predominant among 20 blood samples, followed by ST6 (3.2.3). is aligns with the prevalence of ST3 in China [18]. We also identi-

ed a new allele belonging to the SS14 lineage, resulting in a new ST that di ered from ST1 (1.3.1) by one substitution on TP0136. ST1(1.3.1) was frequently detected in Czech Republic, the Netherlands, France, and Cuba [9, 19–21]. Vrbová et al. conducted a large survey in the Czech Republic from 2004 to 2022, nding that SS14-like strains were predominant [22]. e rst group of isolates included pro les ST 1 (1.3.1) and ST 25 (1.26.1), while the second group comprised ST 3 (1.1.8), ST 2 (1.1.1), and e two groups accounted for 57.5% and ST 11 (1.1.3). 25.3% of the total isolates, respectively. In our study, ST 3 (1.1.8), which represented 45% of the isolates, belongs to the second group found in the Czech Republic, suggesting a somewhat di erent predominant strain. most common Nichols-like strain in the Czech Republic, ST 6 (9.7.3), accounted for only about 6% of the total in the Czech Republic [22], whereas in our study, the major Nichols-like strain, which is also ST 6 (9.7.3), represented 40% of the total, highlighting a signi cant di erence in strain distribution. In summary, our results indicate an MLST pro le similar to that of China, considering the geographical proximity to Japan. However, it's important to note that our typing utilized only three loci, and there might be additional diversity within the SS14-lineage. Limited data for the Asian region in the database highlights the need for a more convenient method to collect data for detailed analysis.

We identi ed three cases strongly suspected to be caused by TEN, the pathogen associated with bejel. A recent study by Kawabata et al. conducted a sequencebased molecular epidemiological analysis of TPA, which is prevalent in Japan. ey encountered instances of TPA that are not easily typed from specimens derived from MSM. Utilizing phylogenetic tree analysis with sequences from TP0548 [23] and TP0856 genes [24], which exhibit relatively low homology between TPA/TPE/TEN, they concluded that seven out of 70 cases were infected with TEN [25]. is report covers specimens collected from 2014 to 2019. When combined with the results of our study from 2019 to 2022, it strongly suggests that TEN, traditionally considered a non-sexually transmitted treponema, has been prevalent among Japanese MSM for at least a decade, following a transmission pattern similar to that of TPA in sexually transmitted syphilis.

We were able to detect TP0136, TP0548, and TP0705 in approximately one-third of the RPR-positive cases. According to a previous study, the PCR-positive rate when using blood samples was lower than that of swab samples [8]. However, in this study the TP0136 detection rate, encompassing the partial pro le, was 32.9% among TPA. Importantly, this doesn't imply that whole blood is unsuitable for molecular epidemiological analysis of syphilis. It's essential to recognize that the gene detection rate serves as a clinical diagnosis within the population of RPR positives. Whole blood samples can easily be collected from all people with syphilis, and the higher sensitivity of PCR positivity in participants with elevated CRP levels suggests an e cient molecular epidemiological analysis by selectively choosing patients for whole blood samples. In this study, we used TP0136 to screen for PCR positivity as part of an e cient MLST method. However, some studies have utilized multiple alleles or shorter amplicons, such as *polA*, for the genetic diagnosis of syphilis [26, 27]. Vrbova et al. performed PCR on multiple alleles simultaneously and reported positive results in 34.8% of cases [28], which is comparable to the sequencing success rate for three alleles in this study. Wang et al. reported PCR positivity rates of 7.4% in latent syphilis and 62.9% in secondary syphilis [27]. In contrast, our study found no signi cant di erence in detection rates between latent and secondary syphilis; however, di erences in HIV status, MSM status, and other background factors between the study populations may have in uenced the results.

Moreover, diagnosing and treating infectious diseases based on antibody titers, monitored through beforeand-after comparisons, relies on relative evaluations. However, antibody titers take time to decline, posing challenges in determining whether the decline is a treatment e ect or a natural process. Given that many syphilis cases are asymptomatic, and individuals who test RPR-positive may have already healed spontaneously, relying solely on antibody titers might not accurately re ect the disease status. erefore, T. pallidum DNA detection rate may o er insights into some etiologies of the disease beyond the sensitivity of the test. A previous study explored the correlation between RPR titer and T. pallidum DNA detection rate, considering the syphilis stage [27]. In addition to RPR, we examine the association between PCR-positives and CRP values. CRP, as an indicator of in ammation [29], a typically remains below 0.3 mg/dL in most healthy individuals, with normal or minor elevations falling within the range of 0.3-1.0 mg/ dL [30]. is observation may be linked to the bacterial load in the blood during early syphilis, serving as a potential pathogenetic factor alongside technical detection sensitivity. PCR-negatives may not necessarily indicate the absence of the pathogen, and therefore, the need for treatment cannot be ruled out. However, it's important to note that antibody titers do not necessarily imply the presence of the pathogen. While pathogen testing is fundamental for treating infectious diseases, it has proven insu cient in syphilis. At this stage, it is di cult to use the results of genetic testing of blood samples to make clinical decisions, and further research is needed. However, in the eld of syphilis care, there is a need for diagnosis and treatment based on antigen testing rather than antibody titers, as is commonly done for other infectious diseases.

e small sample size is one of the limitations of this e uniqueness of this study lies in its use of blood study. specimens from all stages of the disease, not limited to primary syphilis. In MSM, syphilis often presents only as a rosacea or remains asymptomatic, with only a small percentage exhibiting the skin symptoms characteristic of primary syphilis. However, this study demonstrates the feasibility of conducting molecular epidemiological analysis with blood specimens from any stage of the disease. With a larger sample size, statistical signi cance could have been established in the relationship between the RPR and TP0136 detection rate. Nonetheless, numerous cases exhibited low RPR levels with positive PCR results, enabling MLST. e observed discrepancy between the antibody response and the onset of clinical symptoms is a common phenomenon in infectious diseases. erefore. it remains unclear whether the RPR and TP0136 detection rate hold clinical signi cance. Another limitation is the lack of comparison between WGS and MLST. However, given the challenges of applying WGS to TPA, a simpler method is required. is study establishes that, at the very least, MLST possesses the discriminatory power to identify novel epidemic strains and outbreaks of bejel among MSM.

Conclusion

Our study unveiled the dissemination of SS14-like TPA in Japan, with the concurrent detection of Nichols-like strains. CRP emerged as a potential marker for evaluating PCR performance. Individuals diagnosed with syphilis may include those with bejel, and TEN could have been sexually transmitted among Japanese MSM. e MLST scheme, particularly when utilizing blood samples, proves to be valuable for broader research endeavors.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12981-024-00663-y.

Supplementary Material 1

Author contributions

WS primarily conducted the experiments and genome analysis, drafting the initial manuscript. EA designed the project, conducted statistical analysis, and drafted the nal version of the manuscript. EA was also responsible for preparing the nal version of the manuscript. EA and YK conceptualized the project. AS, YN and YK developed the original MLST protocol. EA, MK, and HY were responsible for patient care and specimen collection. All authors have reviewed and approved the nal manuscript.

Funding

This work was supported by JSPS KAKENHI Grant Number JP17K16222.

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval

Ethics approval was granted by the ethics board of the Institute of Medical Science, University of Tokyo (2020-79-0324).

Competing interests

The authors declare no competing interests.

Received: 2 July 2024 / Accepted: 9 October 2024 Published online: 16 October 2024

References

- Newman L, Rowley J, Vander Hoorn S, Wijesooriya NS, Unemo M, Low N, et al. Global estimates of the prevalence and incidence of four curable sexually transmitted infections in 2012 based on systematic review and global reporting. PLoS ONE. 2015;10(12):e0143304. https://doi.org/10.1371/journal. pone.0143304.
- Takahashi T, Arima Y, Yamagishi T, Nishiki S, Kanai M, Ishikane M, et al. Rapid increase in reports of Syphilis Associated with men who have sex with women and women who have sex with men, Japan, 2012 to 2016. Sex Transm Dis. 2018;45(3):139–43. https://doi.org/10.1097/ olg.000000000000768.
- Edmondson DG, Hu B, Norris SJ. Long-term in Vitro Culture of the Syphilis Spirochete Treponema pallidum subsp. pallidum. mBio. 2018;9(3). https://doi. org/10.1128/mBio.01153-18.
- Ratnam S. The Laboratory diagnosis of Syphilis. Can J Infect Dis Med Microbiol. 2005;16:597580. https://doi.org/10.1155/2005/597580.
- Janier M, Unemo M, Dupin N, Tiplica GS, Potocnik M, Patel R. 2020 European guideline on the management of syphilis. J Eur Acad Dermatol Venereol. 2021;35(3):574–88. https://doi.org/10.1111/jdv.16946.
- 6. Rodes B, Liu H, Johnson S, George R, Steiner B. Molecular cloning of a gene (poIA) coding for an unusual DNA polymerase I from

- Kouznetsov AV, Weisenseel P, Trommler P, Multhaup S, Prinz JC. Detection of the 47-kilodalton membrane immunogen gene of Treponema pallidum in various tissue sources of patients with syphilis. Diagn Microbiol Infect Dis. 2005;51(2):143–5. https://doi.org/10.1016/j.diagmicrobio.2004.10.003.
- Theel ES, Katz SS, Pillay A. Molecular and direct detection tests for Treponema pallidum subspecies pallidum: a review of the literature, 1964–2017. Clin Infect Dis. 2020;71(Suppl 1):S4–12.

- 24. P trošová H, Zobaníková M, ejková D, Mikalová L, Pospíšilová P, Strouhal M, et al. Whole genome sequence of Treponema pallidum ssp. pallidum, strain Mexico A, suggests recombination between yaws and syphilis strains. PLoS Negl Trop Dis. 2012;6(9):e1832. https://doi.org/10.1371/journal.pntd.0001832.
- Kawahata T, Kojima Y, Furubayashi K, Shinohara K, Shimizu T, Komano J, et al. Bejel, a nonvenereal treponematosis, among men who have sex with men, Japan. Emerg Infect Dis. 2019;25(8):1581–3. https://doi.org/10.3201/ eid2508.181690.
- Grillová L, P. trošová H, Mikalová L, Strnadel R, Dastychová E, Kuklová I, et al. Molecular typing of Treponema pallidum in the Czech Republic during 2011 to 2013: increased prevalence of identi ed genotypes and of isolates with macrolide resistance. J Clin Microbiol. 2014;52(10):3693–700. https://doi. org/10.1128/jcm.01292-14.
- Wang C, Cheng Y, Liu B, Wang Y, Gong W, Qian Y, et al. Sensitive detection of Treponema pallidum DNA from the whole blood of patients with syphilis by the nested PCR assay. Emerg Microbes Infect. 2018;7(1):83. https://doi. org/10.1038/s41426-018-0085-2.
- Vrbová E, Mikalová L, Grillová L, Pospíšilová P, Strnadel R, Dastychová E, et al. A retrospective study on nested PCR detection of syphilis treponemes

in clinical samples: PCR detection contributes to the diagnosis of syphilis in patients with seronegative and serodiscrepant results. PLoS ONE. 2020;15(8):e0237949. https://doi.org/10.1371/journal.pone.0237949.

- 29. Kushner I. The phenomenon of the acute phase response. Ann N Y Acad Sci. 1982;389:39–48. https://doi.org/10.1111/j.1749-6632.1982.tb22124.x.
- Nehring SM, Goyal A, Patel BC. C reactive protein. StatPearls. Treasure Island (FL): StatPearls Publishing Copyright © 2024. StatPearls Publishing LLC.; 2024.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional a liations.

Wakana Sato Wakana Sato is a researcher working at the University of Tokyo as a clinical laboratory technician while conducting research on the molecular epidemiology of infectious diseases.