

Exogenous spermidine enhances salt-stressed rice photosynthetic performance by stabilizing structure and function of chloroplast and thylakoid membranes

D.X. JIANG, X. CHU, M. LI, J.J. HOU, X. TONG, Z.P. GAO⁺, and G.X. CHEN⁺

Jiangsu Key Laboratory of Biodiversity and Biotechnology, Life Sciences College, Nanjing Normal University, 210023 Nanjing, China

Abstract

Spermidine (Spd) is a ubiquitous low-molecular-mass aliphatic amine that acts in abiotic stress tolerance in plants. We investigated how the treatment with exogenous Spd contributed to the protection against salt stress in rice chloroplasts. Analysis of the chlorophyll (Chl) fluorescence showed that there were many negative effects of salinity on different sites of the photosynthetic machinery; however, these were alleviated by adding Spd. Spd prevented the damage of structure and function of chloroplasts under salt stress; thylakoid membrane protein components and photosynthetic pigments were not severely affected by salinity in Spd-treated plants. Spd enhanced the activities of antioxidant enzymes and decreased contents of reactive oxygen species and malondialdehyde accumulation, suggesting that Spd may participate in the redox homeostasis in chloroplasts under salt stress. These results highlighted the positive effects of Spd on rice chloroplasts under salt stress through maintaining chloroplast structure stability.

Additional key words: growth regulator; OJIP transient; *Oryza sativa*; redox balance.

Introduction

Due to the generation of hyperosmotic effect by reducing the soil water potential, salt stress is one of the most deleterious environmental factors limiting crop growth and yield worldwide (Liu *et al.* 2008). Salinity causes osmotic stress, free radical production, and nutritional stress, which induces structural damage and a loss of turgidity in plant cells, thereby inhibiting plant normal growth (Munns and Tester 2008, Li *et al.* 2016). Notably, salinity-induced inhibition of the photosynthetic electron transport results in excessive accumulation of toxic reactive oxygen species (ROS) and damage of membranes through lipid peroxidation. Previous studies have demonstrated that salinity often leads to a decline of photosynthesis in plants (Mittal *et al.* 2012, Shu *et al.* 2013). Salt stress causes the damage to semipermeability of the plasma membrane resulting from changes in K^+/Na^+ ratios, which breaks the interstitial ionic homeostasis. Salt stress induces the inhibition of photosynthesis through stomatal and nonstomatal limitations including stomatal closure, damage of chloroplast structure, loss of photosynthetic pigments, inhibition of carbon assimilation, and degradation of

membrane proteins in the photosynthetic apparatus (Mittal *et al.* 2012, Gil *et al.* 2013, Shu *et al.* 2013, Li *et al.* 2015a). However, stressed plants possess a pool of antioxidant enzymes including superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX), and glutathione reductase (GR) to prevent accumulation of ROS and hence the damage of lipids, proteins, and DNA (Noctor and Foyer 1998).

Polyamines (PAs), including putrescine (Put), spermidine (Spd), and spermine (Spm), are ubiquitously distributed in living organism, which has been regarded as a new class of growth substances in improving plant stress resistance ability (He *et al.* 2002, Pál *et al.* 2015). These three main PAs were found to be closely related to thylakoid membranes and photosynthetic subcomplexes (Kotzabasis *et al.* 1993). It is well known that PAs act as the stabilizing factors on biomembranes and macromolecules (Watanabe *et al.* 1991, Navakoudis *et al.* 2007). Much evidence has shown that application of exogenous PAs can reduce the loss of pigments, inhibit destruction of the thylakoids, and enhance the photosynthetic rate (Besford *et al.* 1993, Beigbeder *et al.* 1995). Salinity damage to PSI and PSII activity is also effectively alleviated by the addition of

Received 9 October 2019, accepted 19 November 2019.

⁺Corresponding author; e-mail: guoxiangchennjnu@163.com (G.X. Chen), ketty-1982@163.com (Z.P. Gao)

Abbreviations: APX – ascorbate peroxidase; BN-PAGE – blue native-polyacrylamide gel electrophoresis; CAT – catalase; Cyt b_6/f – cytochrome b_6/f complex; DM – n-dodecyl- β -D-maltoside; DTNB – 5,5'-dithiobis-(2-nitrobenzoic acid); GR – glutathione reductase; K_a – nonphotochemical rate constant; K_p – photochemical rate constant; MDA – malondialdehyde; OEC – oxygen-evolving complex; PAs – polyamines; RC – reaction center; ROS – reactive oxygen species; SOD – superoxide dismutase; Spd – spermidine; TBA – thiobarbituric acid; TEM – transmission electron microscopy.

Acknowledgements: This work was supported by the Jiangsu Agricultural Science and Technology Independent Innovation Fund (CX173022, CX181001) and Natural Science Foundation of Jiangsu province (BK20171034).

exogenous PAs through the roots (Chattopadhyay *et al.* 2002). Additionally, it has been reported that PAs reverse malondialdehyde (MDA) accumulation, scavenge ROS, and enhance the activities of antioxidant enzymes (Igarashi and Kashiwagi 2000, Kuznetsov *et al.* 2006, Li *et al.* 2015a). Among the three major forms of PAs, Spd is closely associated with stress tolerance in plants (Kasukabe *et al.* 2004). Exogenous application of Spd confers short-term salinity tolerance in creeping bentgrass through activating antioxidant enzymes and affecting the activities of enzymes involved in PA metabolism (Li *et al.* 2015b). Duan *et al.* (2008) described the improvement of net photosynthetic rate and actual photochemical efficiency of PSII in salt-stressed cucumber seedlings treated with Spd. However, Beauchemin *et al.* (2007) noted that high concentration of Spd was shown to elicit strong inhibitory effects on PSII submembrane fractions of spinach thylakoids, resulting in a decrease of plastoquinone reduction. Exogenous Spd plays vital roles in preventing the electrolyte and amino acid leakage, photosynthetic pigment loss, inhibition of photochemical reactions, or recovering the plasma membrane damage in rice in response to salinity (Chattopadhyay *et al.* 2002, Roychoudhury *et al.* 2011).

Chloroplast is the essential organelle of green plants, where photosynthesis takes place and is very sensitive to the changes of external environment (Kaiser *et al.* 2015). The objective of the present work was to assess the effect of exogenous Spd treatment on the acclimation of rice chloroplast to salt stress. This study was undertaken to test the hypothesis that exogenous Spd acts *via* the improvement of the antioxidant defense systems in chloroplasts. Spd may play a significant protective role in maintaining the redox balance in chloroplasts and the integrity of photosynthesis apparatus in response to salt stress. The knowledge of the chloroplast adaptive mechanisms induced by Spd helps further reveal how this plant growth regulator provides tolerance against salinity to rice.

Materials and methods

Plant materials, growth conditions, and experimental design: Seeds of rice cultivars, *Oryza sativa* L. cv. Nipponbare, were kindly gifted from Prof. Chuangen Lv (Jiangsu Academy of Agricultural Sciences, China). Uniform and healthy seeds of rice were surface-sterilized with 5% sodium hypochlorite for 30 min followed by washing several times. The seeds were then placed in Petri plates containing several layers of filter paper moistened with distilled water for germination. The rice seedlings were kept at a plant growth chamber [irradiance of 500 $\mu\text{mol}(\text{photon}) \text{m}^{-2} \text{s}^{-1}$; temperature of 30/25°C day/night; relative humidity of 80%] for 30 d. Hoagland's nutrient solution was applied as nutrient. A preliminary experiment, employing 100 mM NaCl to simulate moderate salt stress and different Spd concentrations (0, 0.2, 0.6, 1.0, 1.5 mM) were used as a salt-stress tolerance test. It showed that 1 mM Spd exhibited the best effect on reducing malondialdehyde (MDA) content in the chloroplasts of rice seedlings under salt stress. Four general conditions were used in the experiments:

Treatment	
CK (control)	Hoagland's nutrient solution + 0 mM Spd
Spd	0 mM NaCl + 1 mM Spd
NaCl	100 mM NaCl + 0 mM Spd
NaCl + Spd	100 mM NaCl + 1 mM Spd

Hoagland nutrient solution was renewed every 3 d, and the pH of nutrient solutions was adjusted every day to 6.5 with 0.05 M HCl or NaOH supplementation. After 7-d treatment, the fourth fully expanded leaves of each treatment were randomly selected to measure Chl fluorescence parameters, detect chloroplast ultrastructure, and then were frozen in liquid nitrogen immediately after collection and stored at -80°C for other experiments. Treatments were carried out using a completely randomized design with three replications and were analyzed independently.

Photosynthetic pigments: The Chl *a*, Chl *b*, and total Chl concentration were determined according to the method of Lichtenthaler (1987). Chl was extracted from the fourth fully expanded leaves (100 mg) with 80% acetone in darkness for 24 h at 25°C. The concentration of photosynthetic pigment was estimated at 663 and 644 nm by using a spectrophotometer (GENESYS 10, Thermo Electron, USA).

Chl *a* fluorescence transient and JIP-test: The Chl *a* fluorescence induction kinetics was measured at room temperature using a pocket fluorometer (Handy PEA, Hansatech, UK). A typical Chl fluorescence transient shows a sequence of phases from the initial (F_0) to the maximal (F_M) fluorescence value, which have been labeled as O-step (20 μs), J-step (2 ms), I-step (30 ms), and P-step (0.5–1 s) (Strauss *et al.* 2006). The leaves were dark-adapted for at least 30 min before measurements. For each treatment, 12 plants were randomly selected for measurement. The transients in leaves were induced by red light (peak at 650 nm) of 3,000 $\mu\text{mol}(\text{photon}) \text{m}^{-2} \text{s}^{-1}$ provided by an array of three light-emitting diodes. The fluorescence signal induced between 10 s and 1 s is received by the sensor head and is digitized in the control unit using a fast digital converter. Original fluorescence transients were transferred with the professional PEA Plus and BioLyzer HP3 software to a spreadsheet. The JIP-test defines the maximal (subscript '0') energy fluxes in the energy cascade for the events absorption (ABS), trapping (TR_0), electron transport (ET_0), and reduction of end acceptors (RE_0). PSII behavior was characterized on the basis of several parameters derived from the OJIP transients, which are based on the Theory of Energy Fluxes in Biomembranes (Strasser 1981). Therefore, the definitions and equations of the JIP-test are shown in Fig. 1S (*supplement*) by using a scheme (modified after Tsimilli-Michael and Strasser 2008), which is the well-known Z-scheme of photosynthesis. The energy fluxes and energy bifurcations (wide arrows) represent the outfluxes for energy conservation (grey arrows) and the outfluxes for dissipation (white arrows), respectively. Further details of formulae and glossary of terms used by the JIP-test are listed in Table 1S (*supplement*).

Transmission electron microscopy (TEM) analysis: Fresh unfolded leaf tissues (*ca.* 2 mm²) of rice seedlings were fixed in 1% (v/v) glutaraldehyde for 3 d at 4°C. Samples were postfixed in 1% (v/v) osmium tetroxide for 2 h at 4°C followed by dehydration in a gradient series of increasing acetone concentrations. Then, the dehydrated samples were progressively embedded in Epon-araldite, followed by polymerization in the resin. Ultrathin sections (about 80 nm) were prepared and mounted on 300-mesh coated copper grids for staining with uranyl acetate for 30 min and lead citrate for 7 min and finally examined using a *Hitachi 600A-2* transmission electron microscope.

Isolation of intact chloroplasts: Intact chloroplasts were isolated according to the method of Seignur-Berny *et al.* (2008) with a slight modification. Briefly, leaves (15 g) were homogenized at 4°C with 30 ml of 2 mM ascorbate, 30 mM MES, 330 mM sorbitol, and 0.1% BSA (w/v), adjusted to pH 6.5 with Tris. The homogenate was filtered through four layers of skim gauze and centrifuged at 1,500 × *g* for 5 min. The precipitations were resuspended in 3 ml of suspension buffer (330 mM sorbitol, 30 mM Hepes and 0.2% BSA, adjusted to pH 7.5 with Tris), then put into a tube containing 10 ml of chloroplast resuspension medium in 40/80% *Percoll*, and horizontally centrifuged for 20 min at 3,000 × *g*. The intact chloroplasts were located at the interlayer between 40 and 80% *Percoll*, and washed twice by suspension buffer after aspiration in order to remove *Percoll* followed by centrifugation at 12,000 × *g* for 10 min. All procedures were carried out at 4°C. The chloroplast pellets were resuspended in ice-cold resuspension buffer containing 272 mM sucrose, 40 mM Hepes (pH 7.5) and 150 mM KCl, and adjusted to the equal Chl concentration. The intactness of chloroplasts was about 80% according to the method of Aravind and Prasad (2004).

Blue native-polyacrylamide gel electrophoresis (BN-PAGE): The supernatant enriched with chloroplasts was supplemented with an equal volume of 2% n-dodecyl-β-d-maltoside (DM) in a dark ice bath for 45 min. After 15-min centrifugation with 16,000 × *g* at 4°C, the supernatant was supplemented with 1/10 volume of 5% *Serva Blue G* [100 mM Bis-Tris-HCl (pH 7.0), 0.5 M 6-amino-n-caproic acid, and 30% glycerol], and subjected to BN-PAGE according to the method described by Schagger and von Jagow (1991). The equal volumes of supernatant equivalent to 8 μg(Chl) were subjected to BN-PAGE with a linear gradient of 5–12% (w/v) acrylamide and a stacking gel of 4.5% (w/v) acrylamide. Electrophoresis was carried out at 4°C, set at 80 V until the proteins had entered the stacking gel, and then set to 180 V for approximately 3 h. The BN-gels stained with 0.025% (w/v) *Serva Blue G* were scanned and the optical density along each lane was analyzed by quantitation software (*Quantify One*, Bio-Rad, USA).

ROS production and lipid peroxidation in chloroplasts: Superoxide radical (O₂^{•-}) content was determined by using the method of Elstner and Heupel (1976) with a minor modification. The chloroplast supernatants (0.2 ml) were

mixed with 0.75 ml of 65 mM phosphate buffer solution (pH 7.8) and 0.05 ml of 10 mM hydroxylamine hydrochloride for incubation at 25°C for 30 min. Then, the mixture was mixed with 1 ml of 17 mM sulfanilamide and 1 ml of 7 mM α-naphthylamine, and again incubated at 25°C for 20 min. The absorbance was measured at 530 nm (*GENESYS 10*, Thermo Electron, USA). O₂^{•-} generation rate was calculated by using a standard curve of NaNO₂.

Hydrogen peroxide (H₂O₂) concentration was determined according to Yu *et al.* (2003) with a minor modification. The chloroplast supernatant (0.2 ml) was added to 2 ml of a mixture of 0.1% TiCl₄ in 20% H₂SO₄ (v/v) and centrifuged at 6,000 × *g* for 15 min. The supernatants were spectrophotometrically (*GENESYS 10*, Thermo Electron, USA) measured at 410 nm and calculated by using a standard curve of H₂O₂.

Lipid peroxidation was estimated by determining the concentration of malondialdehyde (MDA) produced by thiobarbituric acid (TBA) reaction according to method of Kramer *et al.* (1991). The chloroplast supernatant (0.5 ml) reacted with 2 ml of 0.5% thiobarbituric acid in 20% TCA, followed by boiling at 95°C for 30 min, and the reaction was stopped in ice bath. The samples were centrifuged at 12,000 × *g* for 10 min, followed by determination of absorbance at 440, 532, and 600 nm.

Antioxidant enzyme activities in chloroplasts: Equal amounts of chloroplasts, by Chl concentration, were added to extract buffer consisting of 25 mM Hepes buffer (pH 7.8), 0.2 mM EDTA, and 2% PVP (w/v). The supernatants were used for antioxidant enzyme activity determination after centrifugation.

Superoxide dismutase (SOD, EC 1.15.1.1) activity was assayed by quantifying the photoreduction of nitroblue tetrazolium (NBT) according to the method described by Panda *et al.* (2017) with slight modifications. Reaction was performed using a reaction mixture consisting of 50 mM phosphate buffer (pH 7.8), 13 mM methionine, 0.1 mM EDTA-Na₂, 50 mM sodium carbonate, 9.9 mM NBT, 2.4 mM riboflavin, and 0.05 ml of enzyme extract. The absorbance was measured at 560 nm, and one unit of enzyme activity was taken as that amount of enzyme that inhibits the reduction of NBT by 50%.

Catalase (CAT, EC 1.11.1.6) activity was measured as the decrease in absorbance at 240 nm that accompanied the consumption of H₂O₂ (Panda *et al.* 2017). The reaction mixture consisted of 1 ml 0.3% H₂O₂, 1.95 ml of H₂O, and 0.05 ml of enzyme extract. The activity was calculated using the extinction coefficient (ε) of 43.6 M⁻¹ cm⁻¹ at 240 nm.

Ascorbate peroxidase (APX, EC 1.11.1.11) activity was measured by recording the change in absorbance at 290 nm as described by Jin *et al.* (2008). The reaction was carried out using a reaction mixture including 50 mM phosphate buffer (pH 7.0), 0.5 mM ascorbic acid, 0.1 mM H₂O₂, and 0.05 ml enzyme extract. The activity of APX was determined by the decrease in the absorbance at 290 nm and the amount of ascorbate oxidized using extinction coefficient (ε) of 2.8 mM⁻¹ cm⁻¹ at 290 nm.

Glutathione reductase (GR, EC 1.6.4.2) activity was estimated by measuring the rate of reduction of 5,5'-

dithiobis-(2-nitrobenzoic acid) (DTNB) as an increase in absorbance at 412 nm ($\epsilon = 14.15 \text{ mM}^{-1} \text{ cm}^{-1}$) according to the procedure described in Panda *et al.* (2017).

For CAT, APX, and GR, one unit of enzyme was defined as the amount of enzyme necessary to decompose 1 μmol of substrate per minute at 25°C.

Statistical analysis: All experiments were carried out in at least three biological replications. The means and standard deviations (SD) were calculated using *SPSS 17.0* and a *Tukey's* HSD multiple range test at a significance level of $P < 0.05$.

Results

Apparent injury and photosynthetic pigments: Salt stress caused severe inhibition of plant growth characteristics (Fig. 1A). The leaves of rice seedlings were dehydrated and shriveled upon NaCl treatment, while Spd amendment to the salt-treated plants decreased the area and the degree of shrinkage; only the edges of leaves exhibited curling. There was a slight increase but no significant difference in the photosynthetic pigment contents of rice seedlings supplemented with Spd under nonsalinity conditions (Fig. 1B–E). Compared with control, salt stress caused reduction of Chl *a* (45.1%), Chl *b* (32.2%), total Chl (44.2%), and Chl *a/b* (19.0%). However, a positive effect of Spd was observed in rice seedlings after 7-d treatment

with the high concentration of NaCl; Chl *a*, Chl *b*, total Chl, and Chl *a/b* increased by 33.0, 13.1, 27.7, and 17.6%, respectively.

Chl *a* fluorescence transients of the dark-adapted leaves of rice seedlings under different treatments were shown, on logarithmic time scale from 10 μs up to 1 s, in Fig. 2A. The following phenomena can be observed from fluorescence transient OJIP curves: (1) all curves showed the typical OJIP shape consisting of O-, J-, I-, P-steps; (2) with the NaCl treatment, K-step appeared, the major change was an increase in the J-step and a decrease in I–P phase; (3) maximum fluorescence (F_M) went down and minimum fluorescence (F_0) remained constant; (4) treatment with Spd significantly increased the intensity of fluorescence in the OJIP transient under salt stress. The insert in Fig. 2A presents the relative variable fluorescence curves normalized by F_0 and F_M : $[V_t = (F_t - F_0)/(F_M - F_0)]$. Obviously, the values of V_K and V_J significantly increased during the exposure to salt stress, and markedly declined by adding exogenous Spd.

Each transient depicted in Fig. 2A can be analyzed by the JIP-test to yield structural and functional parameters quantitating PSII behavior and activity, and the kinetics of PSII electron flow reactions (Fig. 2B). As the spider plot shows in Fig. 2B, we observed that the maximum quantum yield of primary photochemistry (ϕ_{P0}) values changed little under different treatments. In contrast to ϕ_{P0} ,

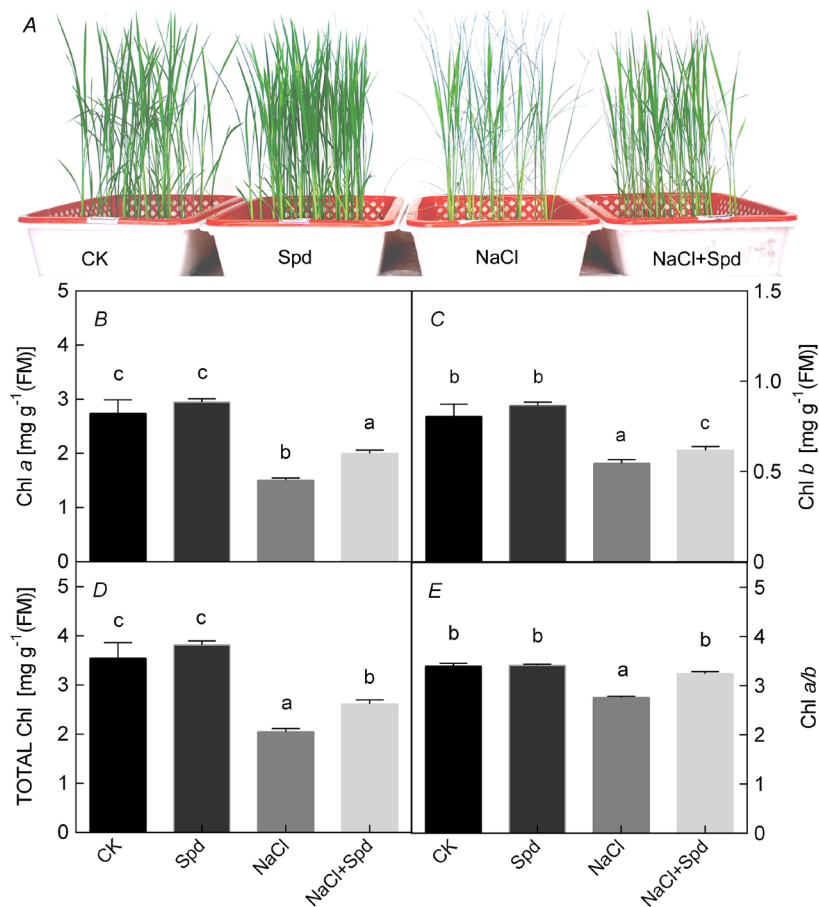


Fig. 1. Effect of exogenous spermidine (Spd) on growth performance and chlorophyll (Chl) content of rice seedlings. (A) growth performance of rice seedlings; (B) Chl *a*; (C) Chl *b*; (D) total Chl; (E) Chl *a/b* ratio. Values are means $\pm$ SD ($n = 6$). Different lowercase letters above the error bars denote significant differences at $P \leq 0.05$.

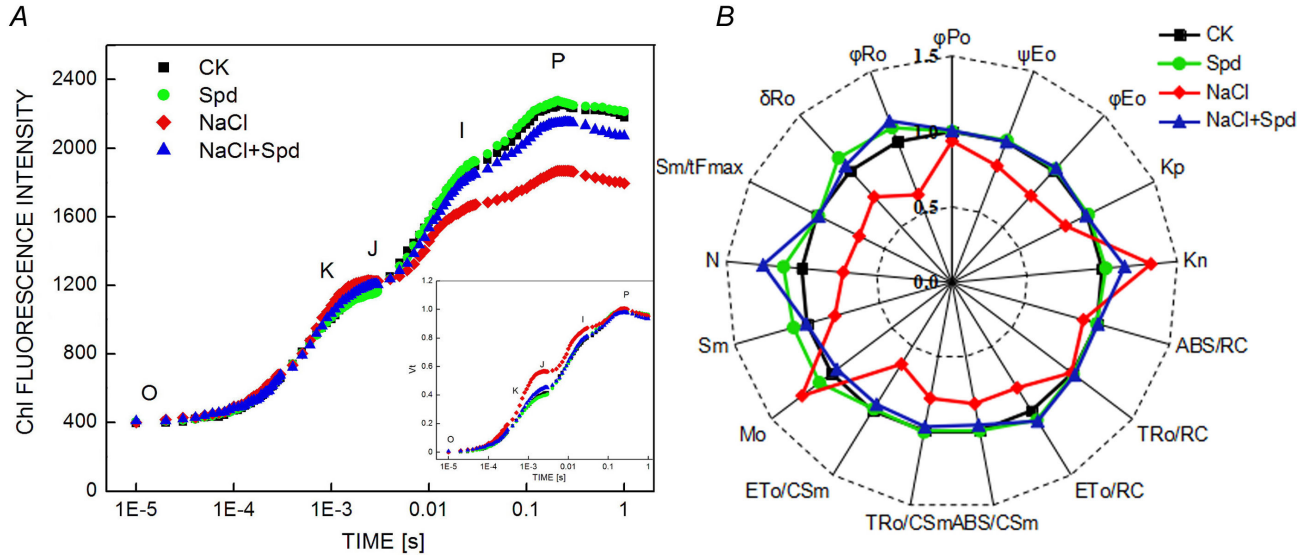


Fig. 2. Effect of exogenous spermidine (Spd) on chlorophyll fluorescence of rice seedlings. (A) raw curves without any normalization and curves normalized by F_0 and F_M [$V_t = (F_t - F_0)/(F_M - F_0)$] in the insert. (B) structural and functional photosynthetic parameters (average values of 12 replicates) derived by the JIP-test from the fluorescence transients. For each parameter, values were normalized by using as reference the control (untreated) plants, presented in the panel by squares (all parameters equal to unity). O indicates the O-step at about 20 μ s; K indicates the K-step at about 300 μ s; J indicates the J-step at about 2 ms; I indicates the I-step at about 30 ms; P indicates the P-step, the maximum fluorescence. All results are the averages of 12 biological replicates. As indicated, black, green, red, blue colors (symbols) refer to different groups and correspond to untreated (CK, squares) and those treated with 1 mM spermidine (Spd, circles), 100 mM NaCl (NaCl, diamonds) or 100 mM NaCl and 1 mM spermidine (NaCl + Spd, triangles), respectively.

the probability to move an electron further than Q_A^- (ψ_{E0}), quantum yield for electron transport (ϕ_{E0}), the probability to reduce an end electron acceptor (δ_{R0}), and quantum yield for reduction of end electron acceptors at the PSI acceptor side (ϕ_{R0}) decreased strongly under salt stress, whereas the application of exogenous Spd alleviated the decline of these parameters dramatically. The parameters referring to the sequential energy transduction include the energy fluxes for (light) absorption (ABS), trapping (TR_0), and electron transport (ET_0). These parameters are expressed per CS_m (cross section) or per RC (reaction center). The energy fluxes per RC are functional parameters (specific energy fluxes), while the energy fluxes per CS_m express the corresponding quantum yields and are structural parameters. Under the salinity conditions, ET_0/RC , ABS/CS_m , TR_0/CS_m , and ET_0/CS_m decreased strongly, but no visible changes in ABS/RC and TR_0/RC occurred. Compared with CK-treated seedlings, those subjected to salt stress exhibited significantly reduced photochemical rate constant (K_p) and increased nonphotochemical rate constant (K_n). However, all these negative effects caused by salt stress could be effectively eliminated by adding exogenous Spd. Compared with the control, rice seedlings subjected to NaCl treatment exhibited significantly increased initial slope (M_0) expressing the net rate of Q_A reduction or the RC closure, while adding Spd decreased it (Fig. 2). After salt stress, the S_m value used to quantify multiple-turnover Q_A reduction events decreased by 20% relative to the control. Meanwhile, total turnover number of Q_A reduction events (N) showed a similar trend to S_m . However, application of exogenous Spd significantly increased the

values of S_m and N of plants with NaCl (increased by 24.3 and 73.6%, respectively). S_m/tF_{max} significantly declined in plants treated only with NaCl, which confirmed the severe closure of PSII RCs induced by NaCl. The presence of Spd counteracted the effect of salt stress, and S_m/tF_{max} value returned to normal levels.

Functional indexes: Fig. 3 showed the effect of Spd on the PSII behaviors, quantified by the performance index (PI_{total}) presented with four components [RC/ABS , $\phi_{P0}/(1 - \phi_{P0})$, $\phi_{E0}/(1 - \phi_{E0})$, $\delta_{R0}/(1 - \delta_{R0})$]. The log of PI_{total} was expressed as DF_{total} called driving force. Furthermore, the product of these four independent parameters is also used as expression of the structure function index for photosynthetic systems (SFI_{total}). Under nonsalinity conditions, the value of PI_{total} significantly improved by 29.6% with adding exogenous Spd. Under salt stress, PI_{total} decreased to 77.9% that of control, while exogenous Spd strongly relieved the decline of PI_{total} . Parameters SFI_{total} and DF_{total} showed the similar trend as that of PI_{total} .

Ultrastructure of chloroplasts: No significant changes occurred in ultrastructure of chloroplasts and thylakoids of rice seedlings between the control and Spd-treated plants as shown by TEM (Fig. 4A,B). The chloroplasts in two groups were of characteristic flat elliptical shape and thylakoids were regularly arranged. Small and rarely found osmiophilic plastoglobuli were presented in these chloroplasts. In contrast to the control, the chloroplasts in salt-stressed plants appeared as seriously distorted and separated from the plasma membrane (Fig. 4C).

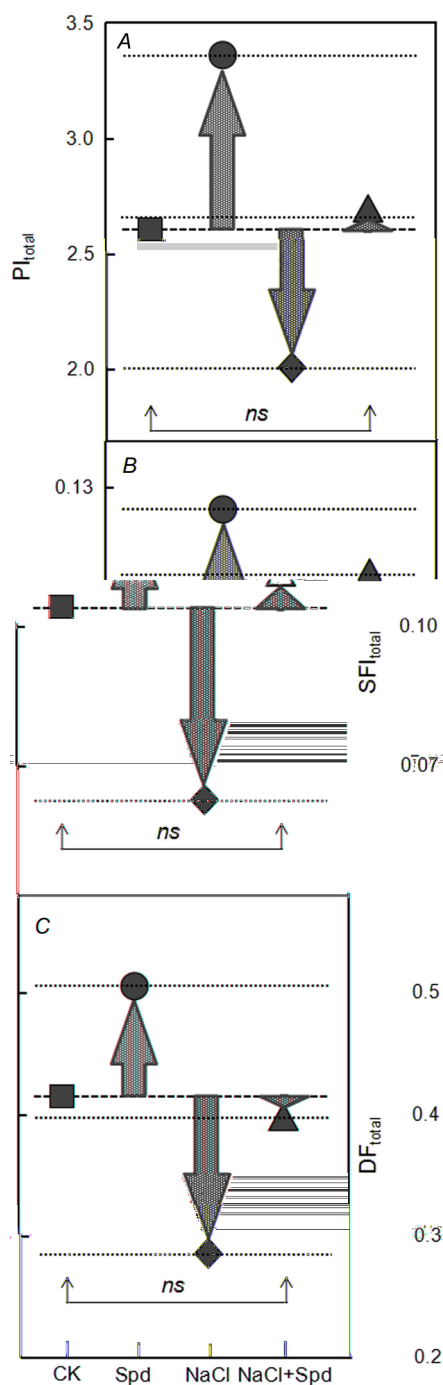


Fig. 3. Effect of exogenous spermidine (Spd) on functional indexes of rice leaves. (A) PI_{total} , performance index; (B) SFI_{total} , structure function index for photosynthetic systems; (C) DF_{total} , driving force on absorption basis. These performance parameters were derived by the JIP-test from chlorophyll fluorescence transients (shown in Fig. 2A) of dark-adapted leaves from rice seedlings that were either untreated (squares), or treated with 1 mM spermidine (circles), 100 mM NaCl (diamonds) or 100 mM NaCl + 1 mM spermidine (triangles). Downwards arrows indicate the loss, while upwards arrows indicate the gain. The values are means of 12 replicates; all the differences except those marked as 'ns' (not significant) are statistically significant ($P < 0.05$).

The internal structure in chloroplasts showed a severe dilation of the thylakoid membranes and dismantling of grana stacks, leaving diffuse membranes and numerous osmiophilic plastoglobuli. Nevertheless, the amount and organization of thylakoids in the chloroplasts of plants under salt stress treated with Spd differed remarkably (Fig. 4D). Exogenous Spd alleviated the structural changes of chloroplasts and thylakoid membranes induced by salt stress. The chloroplasts still maintained several tightly packed grana stacks consisted of at least ten thylakoids and contained less osmiophilic plastoglobuli.

Thylakoid membrane protein complexes: To confirm the chloroplast integrity and further investigate the specific changes in thylakoid membrane, we performed a BN gel separation. We observed five distinct major bands representing the PSI supercomplexes and PSII supercomplexes (PSI SC, PSII SC), PSI monomers, and PSII dimers (PSI-M, PSII-D), PSII monomers, and cytochrome b_6/f complexes (PSII-M, Cyt b_6/f), PSII LHC trimers (LHCII-T), and PSII LHC monomers (LHCII-M) (Fig. 5A). The optical density along each lane was quantified by *Quantify One* software and their relative contents were calculated (Fig. 5B). Except for LHCII-M, no significant differences in thylakoid membrane protein complexes between the control and Spd treated plants were found. However, amounts of PSII SC and PSI SC, PSI-M and PSII-D, PSII-M, and Cyt b_6/f , and LHCII-T (reduced by 35.4, 53.0, 42.1, and 55.6%, respectively) decreased under

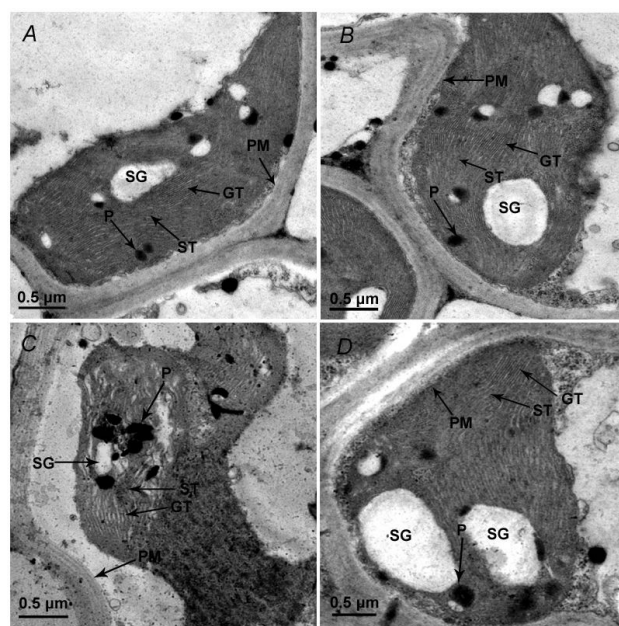


Fig. 4. Effect of exogenous spermidine (Spd) on the ultrastructure of chloroplasts and thylakoids of rice seedlings. (A) CK, 0 mM NaCl + 0 mM spermidine; (B) Spd, 0 mM NaCl + 1 mM Spd; (C) NaCl, 100 mM NaCl + 0 mM Spd; (D) NaCl + Spd, 100 mM NaCl + 1 mM Spd. The fourth fully expanded leaves of two groups were sampled for ultramicroscopic observation after 7-day treatment. ST – stromal thylakoid; GT – grana thylakoid; SG – starch grain; P – plastoglobule; PM – plasma membrane.

NaCl-induced salt stress, while increased (1.25-, 2.01-, 1.36-, and 1.97-fold, respectively) by adding exogenous Spd. Hence, Chl-pigment complexes were stable in Spd-treated leaves during salt stress.

ROS production and lipid peroxidation in chloroplasts: NaCl stress caused a significant increase in the contents of O_2^- (315.4%), H_2O_2 (152.5%), and MDA (361.4%), as compared with the controls (Fig. 6). Application of Spd alone did not notably affect ROS production and MDA content in the chloroplasts of rice seedlings; however, addition of Spd with the 100 mM NaCl prevented the effects of NaCl alone. These results indicated that exogenous Spd mitigated membrane lipid peroxidation in chloroplasts and partially alleviated the suppressed growth.

Antioxidant enzymes in chloroplasts: SOD, CAT, APX, and GR were the enzymes selected to assess the oxidative damage caused by salt stress. In general, treatment with Spd alone increased the activities of the antioxidant enzymes assayed, especially the GR activity was enhanced by 45.7% by Spd (Fig. 7). Compared with the CK, the activities of SOD, APX, and GR significantly increased (Fig. 7A,C,D) under salt stress, except for the CAT activity inhibited by 48.9% (Fig. 7B). After application of Spd, we observed that exogenous Spd further increased the activities of SOD, APX, and GR, and mitigated the activity of CAT inhibited by salinity.

Discussion

Supplementation with Spd has been shown to attenuate various abiotic stresses in plants including *Cucumis sativus*, *Zoysia japonica*, *Avena sativa*, *Oryza sativa*, and *Malus*

sieversii (Besford *et al.* 1993, Chattopadhyay *et al.* 2002, Liu *et al.* 2008, Roychoudhury *et al.* 2011, Shu *et al.* 2013, Li *et al.* 2016). Nevertheless, our knowledge regarding the mechanisms involved in Spd-mediated tolerance to salt stress still remains fragmentary. In the present study, we confirmed that addition of Spd had a protective role on the photosynthetic apparatus of rice seedlings exposed to high salt conditions.

Salt stress often led to unfavorable effects on the growth and Chl metabolisms of plants (Mittal *et al.* 2012, Li *et al.* 2015a, Jin *et al.* 2019). It is well known that Chl *b* is formed from Chl *a* by the oxidation of the methyl group on ring II of the aldehyde. As the first step of Chl degradation, the conversion of Chl *a* to Chl *b* also influences the Chl *a/b* ratio. In the present study, the dehydration and withering status in leaves of the 100 mM NaCl-treated seedlings significantly reversed when supplied with exogenous Spd (Fig. 1A). This was consistent with previous studies (Roychoudhury *et al.* 2011). Substantial enhancement of photosynthetic pigment contents occurred in Spd-treated plants under salt stress (Fig. 1B–E). In addition, Spd strongly increased the Chl *a/b* ratio, indicating that Spd effectively prevented the loss of Chl *a* and maintained the photosynthetic performance of seedlings under salt stress, because high Chl *a* as the pigment of the RCs is more conducive to plant light energy conversion and photosynthetic efficiency (Farquhar and Sharkey 1982). Our results strongly confirmed the beneficial effect of Spd on the Chl content of rice seedlings under salt stress, especially on the Chl *a* content.

The intricate relationships between fluorescence kinetics and photosynthesis are very convenient and straightforward for us to understand the changes in photosynthetic biophysical processes during plant responses

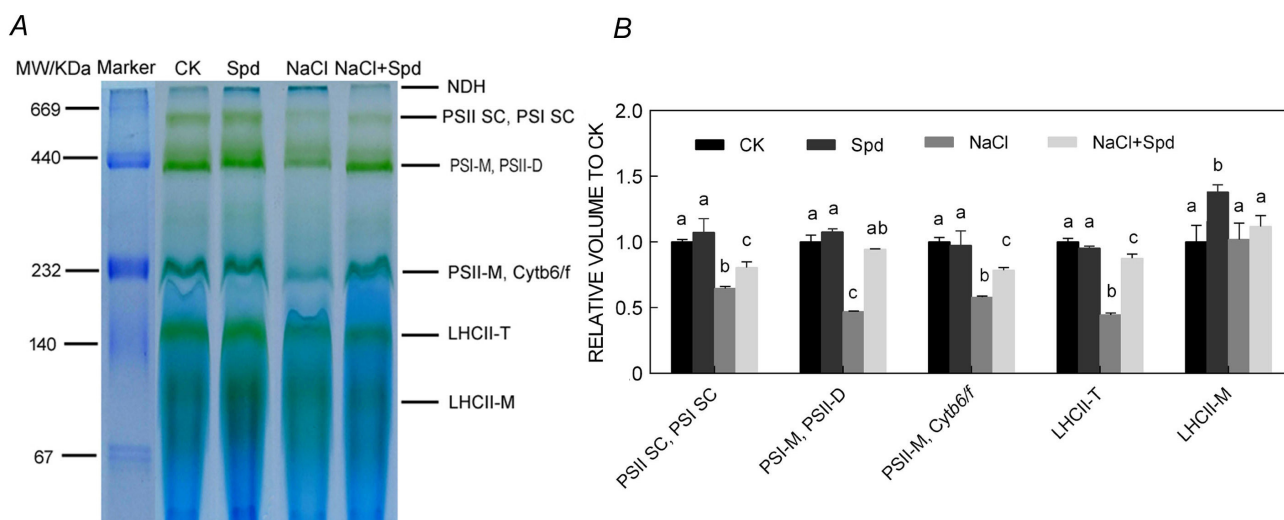


Fig. 5. Effect of exogenous Spd on thylakoid membrane protein complexes of rice seedlings. (A) Distribution of thylakoid membrane protein complexes (8 μ g chlorophylls) separated by BN-PAGE. Molecular mass (MW) of marker was labeled on the left and the nomenclature of the resolved bands was on the right. (B) Quantitation of thylakoid membrane protein complexes. The BN-gels were scanned and the optical density along each lane was quantified (*Quantify One*, Bio-Rad, USA). For each thylakoid membrane protein complex, values were normalized by using as reference the control (untreated) plants (all parameters equal to unity). Values are means $\pm$ SD ($n = 3$). LHCII-M – PSII LHC monomers; LHCII-T – PSII LHC trimers; NDH – NADPH dehydrogenase complexes; PSI-M – PSI monomers; PSI SC – PSI supercomplexes; PSII SC – PSII supercomplexes; PSII-M – PSII monomers; PSII-D – PSII dimers; Cyt *b₆/f* – cytochrome *b₆/f* complexes.

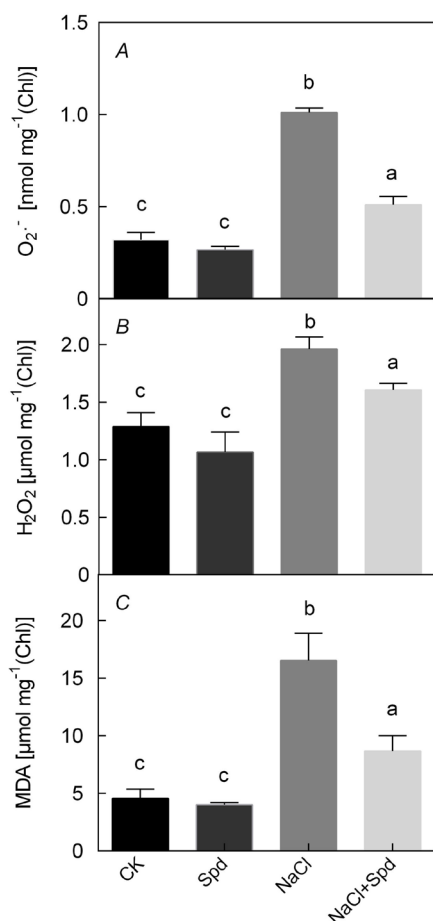


Fig. 6. Effect of exogenous spermidine (Spd) on reactive oxygen species and lipid peroxidation in chloroplasts of rice seedlings. (A) $O_2^{\cdot -}$, superoxide radical; (B) H_2O_2 , hydrogen peroxide; (C) MDA, malondialdehyde. Values are means $\pm$ SD ($n = 3$). Different lowercase letters above the error bars denote significant differences at $P \leq 0.05$.

to environmental stress (Sayed 2003). In this study, salt stress-induced damage to PSII photochemistry was demonstrated by the Chl *a* fluorescence kinetics (Fig. 2A). Single-turnover events with respect to Q_A reduction mainly occur in O–J phase (Strasser 1981, Strauss *et al.* 2006). Therefore, compared with the CK, a strong increase of the J level in stressed group reflected the accumulation of Q_A^- , which was also the evidence for the damage in PSII donor side and electron transfer chain (Fig. 2A). Although NaCl did not significantly change the F_0 , a distinct decrease in F_V and F_M indicated the lower ability of PSII to reduce the primary acceptor Q_A . The drop in the area formed vs. time between the F_I and F_M of the I–P phase manifested the decline in the size of the terminal electron acceptor pool. Obviously, the levels of V_K and V_J significantly increased during the exposure to salt stress, which suggested that NaCl treatment could result in an inhibition of oxygen-evolving complex (OEC) and a fast accumulation of Q_A^- in PSII RCs (Strasser *et al.* 1995, Lazár 2006). However, these negative impacts were markedly alleviated by applying exogenous Spd. PSII behavior/activity was

characterized on the basis of several parameters derived from the OJIP-transients, which are based on the Theory of Energy Fluxes in Biomembranes (Strasser 1981, Strasser *et al.* 1995, Sayed 2003). In this study, salt stress induced the reduction of ϕ_{E0} and ψ_{E0} , which reflected the decrease in electron transfer rate on the PSII acceptor side (Fig. 2B). Additionally, V_J and M_0 increased significantly under salt stress, further demonstrating the fact that electrons transferred from Q_A to Q_B on the PSII acceptor side were blocked. Notably, excess activation energy generated by the blockade of electron transfer may be converted into ROS, causing the oxidative damage to thylakoid membrane (Fig. 5). We also observed the drop in the absorption energy flux per RC or per excited cross section (ABS/RC, ABS/CS_m) under salt stress, suggesting PSII antenna pigments were damaged. There was no significant difference in trapping energy flux per RC (TR₀/RC), while the electron transport flux (ET₀/RC) decreased notably. Furthermore, salt stress also significantly decreased the energy fluxes per excited cross section (TR₀/CS_m and ET₀/CS_m), which may be attributed to the decrease of PSII active RCs per excited cross section (RC/CS_m). The decreases of V_J and M_0 , and the increases of ϕ_{E0} , ψ_{E0} , ABS/RC, ET₀/RC, ABS/CS_m, TR₀/CS_m, and ET₀/CS_m in exogenously Spd-treated plants indicated that Spd effectively maintained the stability of the electron transport chain and the activity of PSII.

The performance index (PI_{total}) is one of the most sensitive indicators to various stress conditions (Tsimilli-Michael and Strasser 2008). We observed that Spd enhanced the salinity resistance of rice seedlings by increasing the decreases of PI_{total} , DF_{total} , and SFI_{total} caused by NaCl (Fig. 3). The increases in these parameters indicated that Spd had the ability to alleviate the adverse effect of salinity on the PSII structure, function, and activity. PI_{total} and SFI_{total} are produced by the four independent components RC/ABS, ϕ_{P0} , ϕ_{E0} , and δ_{R0} (Tsimilli-Michael and Strasser 2008). NaCl mainly acted on ϕ_{E0} and δ_{R0} , not on ϕ_{P0} or RC/ABS, suggesting that the most direct cause of the decrease in PI_{total} and SFI_{total} was the rapid decrease in the efficiency of electron transfer redox reactions (Fig. 2B). This assumption may obtain further support from results concerning the decline of electron transport (ET₀/RC and ET₀/CS_m), a strong decrease of K_p , and a significant increase of K_n in comparison to control. From JIP-test, the decline in S_m and N caused by salt stress led to a large accumulation of Q_A^- , which inhibited PSII RCs activities. S_m/t_{Fmax} (the average redox state of the redox couple Q_A^-/Q_A) significantly declined in plants treated only with NaCl, meaning that NaCl caused the severe closure of PSII RCs. When treated with exogenous Spd, the significant increase in S_m/t_{Fmax} indicated that Spd could effectively alleviate the closure of PSII RCs. Maintaining structural and functional organization of chloroplasts is vital for the dynamics and flexibility of the photosynthetic apparatus. Photosystems involve Chl and pigment-protein complexes located in thylakoids. Therefore, a proper state of thylakoids is critical for stabilizing the function and efficiency of photosystems (Zhang *et al.* 2010). In this study, we detected the degradation of thylakoid membranes with the accumulation of plastoglobuli under

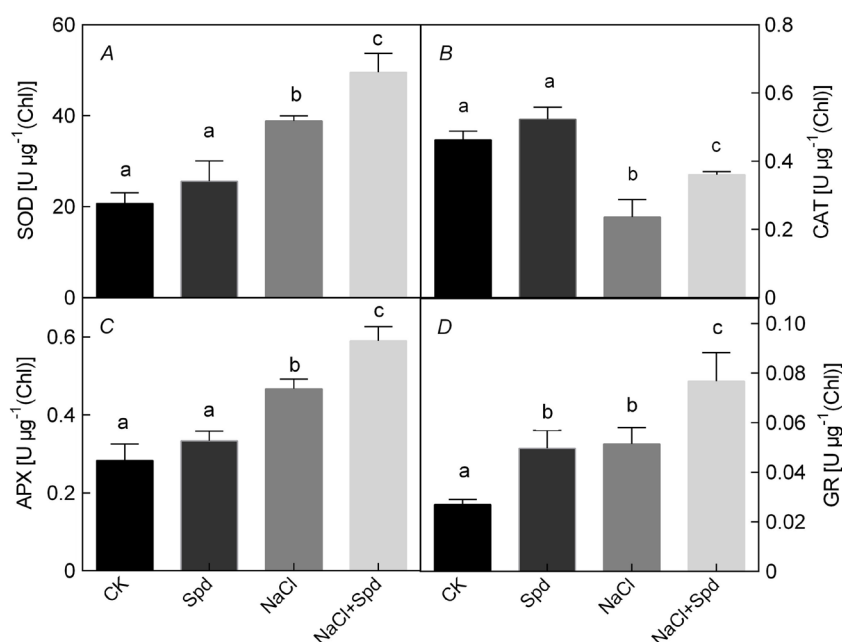


Fig. 7. Effect of exogenous Spd on anti-oxidant enzyme activities in chloroplasts of rice seedlings. (A) SOD, superoxide dismutase; (B) CAT, catalase; (C) APX, ascorbate peroxidase; (D) GR, glutathione reductase. Values are means $\pm$ SD ($n = 3$). Different lowercase letters above the error bars denote significant differences at $P \leq 0.05$.

salt stress (Fig. 4C), which was generally attributed to lipid peroxidation-mediated membrane damage. By contrast, the stressed rice seedlings treated with exogenous Spd showed less osmiophilic plastoglobuli and more grana consisting of numerous tightly packed thylakoids in chloroplasts (Fig. 4D). Many studies have focused on Chl and protein degradation during chloroplast breakdown (Hörtensteiner *et al.* 2009). We found that major thylakoid membrane protein complexes strongly decreased under salt stress (Fig. 5), while the addition of Spd prevented it. Photosystem supercomplexes, especially PSII supercomplexes as well as its two components, LHCII trimers and PSII dimers (Nelson and Yocum 2006), were significantly promoted by Spd, indicating that Spd had a significant positive effect on maintaining the integrity of the PSII structure and supported the capture and transport of light energy. Furthermore, Cyt *b₆/f* also increased under the action of exogenous Spd during salt stress, which was consistent with the higher transmission capacity of electron transfer chain (Fig. 2). Notably, content of Chl and amount of thylakoid membrane protein complexes increased by applying Spd, which may be attributed to the covalent binding of Spd to thylakoid membrane proteins to stabilize the thylakoid membrane structure or enhance the repair process of thylakoid membrane components (Dondini *et al.* 2003).

When plants are exposed to environmental stresses, loss of PSII activity causes the block of electrons in the electron transport chain, leading to an imbalance between energy production and utilization. Therefore, excess electrons are converted into ROS, such as singlet oxygen ($^1\text{O}_2$), $\text{O}_2^{\cdot-}$, $\text{OH}^{\cdot}$, and H_2O_2 in chloroplasts (Asada 2006). In turn, the overproduction of ROS can result in the damage of PSII activity by inducing the degradation of pigment-protein complexes and inhibiting the repair process of PSII (Allakhverdiev *et al.* 2008). ROS are primarily derived from chloroplasts, which can trigger the thylakoid

membrane lipid peroxidation by $^1\text{O}_2$ or oxygen reduction (Sharma *et al.* 2012). Therefore, MDA, membrane lipid peroxidation product, can substantially act as an important reflection of membrane injury caused by oxidative stress (Anjum *et al.* 2012). In the present study, we found that salt stress induced the accumulation of H_2O_2 and $\text{O}_2^{\cdot-}$ in rice chloroplasts (Fig. 6A,B). Meanwhile, MDA significantly increased under salt stress compared to normal conditions (Fig. 6C), demonstrating that ROS overproduction was responsible for the damage of thylakoid membrane induced by salt stress.

The plants have an intrinsic mechanism that comprises of enzymatic antioxidative system to counter the salinity-induced oxidative stress triggered by the overproduction of ROS. Previous studies have reported that Spd confers salinity tolerance in plants through synthesizing antioxidant metabolites, activating antioxidant enzymes, or/and affecting the activities of enzymes involved in PA metabolism to counter the salinity induced oxidative stress triggered by the excess ROS (Kuznetsov *et al.* 2006, Li *et al.* 2015a,b). As revealed in the present study, SOD, APX, and GR activities increased significantly in NaCl or NaCl + Spd-treated plants with respect to the control (Fig. 7). Among all the antioxidant enzymes, SOD acts as a primary defense enzyme scavenging $\text{O}_2^{\cdot-}$ radical. SOD initiates detoxification of $\text{O}_2^{\cdot-}$ by forming H_2O_2 , which is also toxic and is eliminated by conversion to H_2O in the subsequent reactions through a number of enzymes, such as CAT, APX, and GR. Although CAT activity in chloroplasts was significantly inhibited by salinity, it greatly increased by application of Spd (Fig. 7B). As two key enzymes in the ASA-GSH cycle, APX and GR play a central role in the regulation of the intracellular redox balance (Mittler 2002). In this cycle, AsA is catalyzed by APX to form monodehydroascorbate by consuming H_2O_2 , and GSSG is reduced by GR to form GSH (Apel and Hirt 2004). In the present study, both APX and GR

activities dramatically increased by exposure to NaCl + Spd treatment, in comparison to NaCl treatment alone (Fig. 7C,D). In addition, GSH may help stabilize enzymes of the Calvin cycle (Bradbeer 1969), which could explain that plants treated with Spd exhibited much higher biomass (Fig. 1) and showed better photosynthetic performance than that of controls (Fig. 3). These results indicated that Spd-dependent reduction of the oxidative stress caused by salinity was closely related to the induction of antioxidant enzyme activities in chloroplasts. Obviously, the application of exogenous Spd efficiently reduced ROS accumulation and lipid peroxidation levels in chloroplasts by enhancing antioxidant enzyme system, which ensured a better photosynthetic environment and increased plant tolerance to salt stress.

Conclusions: Generally, exogenous Spd treatment had significantly beneficial effects on NaCl-exposed rice seedlings and effectively alleviated NaCl-induced damage of chloroplasts. The lowered contents of supercomplexes, PSII RCs, PSII LHC, and Cyt *b₆/f* in thylakoid membranes under salt stress were promoted by applying Spd, which concomitantly improved photosynthetic pigment contents and photosynthetic performance of rice seedlings. Exogenous Spd successfully prevented salt stress which would otherwise induce the accumulation of ROS and MDA in chloroplasts. Notably, the elevated SOD, CAT, APX, and GR activities in chloroplasts play a vital role in Spd-induced alleviation of oxidative stress. Furthermore, Spd could enhance the repair process of thylakoid membranes under salt stress, conferring higher oxidative stress tolerance to rice chloroplasts.

References

- Allakhverdiev S.I., Kreslavski V.D., Klimov V.V. *et al.*: Heat stress: an overview of molecular responses in photosynthesis. – *Photosynth. Res.* **98**: 541-550, 2008.
- Anjum S.A., Saleem M.F., Wang L.C. *et al.*: Protective role of glycinebetaine in maize against drought-induced lipid peroxidation by enhancing capacity of antioxidative system. – *Aust. J. Crop Sci.* **6**: 576-583, 2012.
- Apel K., Hirt H.: Reactive oxygen species: metabolism, oxidative stress, and signal transduction. – *Annu. Rev. Plant Biol.* **55**: 373-399, 2004.
- Aravind P., Prasad M.N.V.: Zinc protects chloroplasts and associated photochemical functions in cadmium exposed *Ceratophyllum demersum* L., a fresh water macrophyte. – *Plant Sci.* **166**: 1321-1327, 2004.
- Asada K.: Production and scavenging of reactive oxygen species in chloroplasts and their functions. – *Plant Physiol.* **141**: 391-396, 2006.
- Beauchemin R., Gauthier A., Harnois J. *et al.*: Spermine and spermidine inhibition of photosystem II: disassembly of the oxygen evolving complex and consequent perturbation in electron donation from TyrZ to P680⁺ and the quinone acceptors Q_A⁻ to Q_B. – *BBA-Bioenergetics* **1767**: 905-912, 2007.
- Beigbeder A., Vavakis M., Navakoudis E., Kotzabasis K.: Influence of polyamine inhibitors on light-independent and light-dependent chlorophyll biosynthesis and on the photosynthetic rate. – *J. Photoch. Photobio. B* **28**: 235-242, 1995.
- Besford R.T., Richardson C.M., Campos J.L., Tiburcio A.F.: Effect of polyamines on stabilization of molecular complexes in thylakoid membranes of osmotically stressed oat leaves. – *Planta* **189**: 201-206, 1993.
- Bradbeer J.W.: The activities of the photosynthetic carbon cycle enzymes of greening bean leaves. – *New Phytol.* **68**: 233-245, 1969.
- Chattopadhyay M.K., Tiwari B.S., Chattopadhyay G. *et al.*: Protective role of exogenous polyamines on salinity-stressed rice (*Oryza sativa*) plants. – *Physiol. Plantarum* **116**: 192-199, 2002.
- Dondini L., Del Duca S., Dall'Agata L. *et al.*: Suborganellar localization and effect of light on *Helianthus tuberosus* chloroplast transglutaminases and their substrates. – *Planta* **217**: 84-95, 2003.
- Duan J.J., Li J., Guo S.R., Kang Y.Y.: Exogenous spermidine affects polyamine metabolism in salinity-stressed *Cucumis sativus* roots and enhances short-term salinity tolerance. – *J. Plant Physiol.* **165**: 1620-1635, 2008.
- Elstner E.F., Heupel A.: Inhibition of nitrate formation from hydroxylammonium chloride: a simple assay for superoxide dismutase. – *Anal. Biochem.* **70**: 616-620, 1976.
- Farquhar D.G., Sharkey T.D.: Stomatal conductance and photosynthesis. – *Ann. Rev. Plant Physiol.* **33**: 317-345, 1982.
- Gil R., Boscaiu M., Lull C. *et al.*: Are soluble carbohydrates ecologically relevant for salt tolerance in halophytes? – *Funct. Plant Biol.* **40**: 805-818, 2013.
- He L.X., Nada K., Kasukabe Y., Tachibana S.: Enhanced susceptibility of photosynthesis to low-temperature photo-inhibition due to interruption of chill-induced increase of S-adenosylmethionine decarboxylase activity in leaves of spinach (*Spinacia oleracea* L.). – *Plant Cell Physiol.* **43**: 196-206, 2002.
- Hörtensteiner S.: *Stay-green* regulates chlorophyll and chlorophyll-binding protein degradation during senescence. – *Trends Plant Sci.* **14**: 155-162, 2009.
- Igarashi K., Kashiwagi K.: Polyamines: Mysterious modulators of cellular functions. – *Biochem. Bioph. Res. Co.* **271**: 559-564, 2000.
- Jin X.F., Yang X., Islam E. *et al.*: Effects of cadmium on ultrastructure and antioxidative defense system in hyper-accumulator and non-hyperaccumulator ecotypes of *Sedum alfredii* Hance. – *J. Hazard. Mater.* **156**: 387-397, 2008.
- Jin X.Q., Liu T., Xu J.J. *et al.*: Exogenous GABA enhances muskmelon tolerance to salinity-alkalinity stress by regulating redox balance and chlorophyll biosynthesis. – *BMC Plant Biol.* **19**: 48, 2019.
- Kaiser E., Morales A., Harbinson J. *et al.*: Dynamic photosynthesis in different environmental conditions. – *J. Exp. Bot.* **66**: 2415-2426, 2015.
- Kasukabe Y., He L.X., Nada K. *et al.*: Overexpression of spermidine synthase enhances tolerance to multiple environmental stresses and up-regulates the expression of various stress regulated genes in transgenic *Arabidopsis thaliana*. – *Plant Cell Physiol.* **45**: 712-722, 2004.
- Kotzabasis K., Fotinou C., Roubelakis-Angelakis K.A., Ghanotakis D.: Polyamines in the photosynthetic apparatus: Photosystem II highly resolved subcomplexes are enriched in spermine. – *Photosynth. Res.* **38**: 83-88, 1993.
- Kramer G.F., Norman H.A., Krizek D.T., Mirecki R.M.: Influence of UV-B radiation on polyamines, lipid peroxidation and membrane lipids in cucumber. – *Phytochemistry* **30**: 2101-2108, 1991.
- Kuznetsov V.V., Radyukina N.L., Shevyakova N.I.: Polyamines and stress: Biological role, metabolism, and regulation. –

- Russ. J. Plant Physiol. **53**: 583-604, 2006.
- Lazár D.: The polyphasic chlorophyll *a* fluorescence rise measured under high intensity of exciting light. – *Funct. Plant Biol.* **33**: 9-30, 2006.
- Li J.M., Hu L.P., Zhang L. *et al.*: Exogenous spermidine is enhancing tomato tolerance to salinity-alkalinity stress by regulating chloroplast antioxidant system and chlorophyll metabolism. – *BMC Plant Biol.* **15**: 303, 2015a.
- Li S.C., Jin H., Zhang Q.: The Effect of exogenous spermidine concentration on polyamine metabolism and salt tolerance in zoysiagrass (*Zoysia japonica* Steud) subjected to short-term salinity stress. – *Front. Plant Sci.* **7**: 1221, 2016.
- Li Z., Zhou H., Peng Y. *et al.*: Exogenously applied spermidine improves drought tolerance in creeping bentgrass associated with changes in antioxidant defense, endogenous polyamines and phytohormones. – *Plant Growth Regul.* **76**: 71-82, 2015b.
- Lichtenthaler H.K.: Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. – *Method. Enzymol.* **148**: 350-382, 1987.
- Liu J.H., Inoue H., Moriguchi T.: Salt stress-mediated changes in free polyamine titers and expression of genes responsible for polyamine biosynthesis of apple *in vitro* shoots. – *Environ. Exp. Bot.* **62**: 28-35, 2008.
- Mittal S., Kumari N., Sharma V.: Differential response of salt stress on *Brassica juncea*: Photosynthetic performance, pigment, proline, D1 and antioxidant enzymes. – *Plant Physiol. Bioch.* **54**: 17-26, 2012.
- Mittler R.: Oxidative stress, antioxidants and stress tolerance. – *Trends Plant Sci.* **7**: 405-410, 2002.
- Munns R., Tester M.: Mechanisms of salinity tolerance. – *Annu. Rev. Plant Biol.* **59**: 651-681, 2008.
- Navakoudis E., Vrentzou K., Kotzabasis K.: A polyamine- and LHCII protease activity-based mechanism regulates the plasticity and adaptation status of the photosynthetic apparatus. – *BBA-Bioenergetics* **1767**: 261-271, 2007.
- Nelson N., Yocum C.F.: Structure and function of photosystems I and II. – *Annu. Rev. Plant Biol.* **57**: 521-565, 2006.
- Noctor G., Foyer C.H.: Ascorbate and glutathione: Keeping active oxygen under control. – *Annu. Rev. Plant Phys.* **49**: 249-279, 1998.
- Pál M., Szalai G., Janda T.: Speculation: Polyamines are important in abiotic stress signaling. – *Plant Sci.* **237**: 16-23, 2015.
- Panda A., Rangani J., Kumari A., Parida A.K.: Efficient regulation of arsenic translocation to shoot tissue and modulation of phytochelatin levels and antioxidative defense system confers salinity and arsenic tolerance in the halophyte *Suaeda maritima*. – *Environ. Exp. Bot.* **143**: 149-171, 2017.
- Roychoudhury A., Basu S., Sengupta D.N.: Amelioration of salinity stress by exogenously applied spermidine or spermine in three varieties of indica rice differing in their level of salt tolerance. – *J. Plant Physiol.* **168**: 317-328, 2011.
- Sayed O.H.: Chlorophyll fluorescence as a tool in cereal crop research. – *Photosynthetica* **41**: 321-330, 2003.
- Schägger H., von Jagow G.: Blue native electrophoresis for isolation of membrane protein complexes in enzymatically active form. – *Anal. Biochem.* **199**: 223-231, 1991.
- Seigneurin-Berny D., Salvi D., Dorne A.J. *et al.*: Percoll-purified and photosynthetically active chloroplasts from *Arabidopsis thaliana* leaves. – *Plant Physiol. Bioch.* **46**: 951-955, 2008.
- Sharma P., Jha A.B., Dubey R.S., Pessarakli M.: Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. – *J. Bot.* **2012**: 217037, 2012.
- Shu S., Yuan L.Y., Guo S.R. *et al.*: Effects of exogenous spermine on chlorophyll fluorescence, antioxidant system and ultrastructure of chloroplasts in *Cucumis sativus* L. under salt stress. – *Plant Physiol. Bioch.* **63**: 209-216, 2013.
- Strasser R.J.: The grouping model of plant photosynthesis: Heterogeneity of photosynthetic units in thylakoids. – In: Akoyunoglou G. (ed.): *Photosynthesis III. Structure and Molecular Organisation of the Photosynthetic Apparatus*. Pp. 727-737. Balaban International Science Services, Philadelphia 1981.
- Strasser R.J., Srivastava A., Govindjee: Polyphasic chlorophyll *a* fluorescence transient in plants and cyanobacteria. – *Photochem. Photobiol.* **61**: 342, 1995.
- Strauss A.J., Krüger G.H.J., Strasser R.J., Van Heerden P.D.R.: Ranking of dark chilling tolerance in soybean genotypes probed by the chlorophyll *a* fluorescence transient O-J-I-P. – *Environ. Exp. Bot.* **56**: 147-157, 2006.
- Tsimilli-Michael M., Strasser R.J.: *In vivo* assessment of stress impact on plants' vitality: Applications in detecting and evaluating the beneficial role of mycorrhization on host plants. – In: Varma A. (ed.): *Mycorrhiza. State of the Art, Genetics and Molecular Biology, Eco-Function, Biotechnology, Eco-Physiology, Structure and Systematics*. 3rd Edition. Pp. 679-703. Springer, Berlin-Heidelberg 2008.
- Watanabe S., Kusama-Eguchi K., Kobayashi H., Igarashi K.: Estimation of polyamine binding to macromolecules and ATP in bovine lymphocytes and rat liver. – *J. Biol. Chem.* **266**: 20803-20809, 1991.
- Yu C.W., Murphy T.M., Lin C.H.: Hydrogen peroxide-induces chilling tolerance in mung beans mediated through ABA-independent glutathione accumulation. – *Funct. Plant Biol.* **30**: 955-963, 2003.
- Zhang M.P., Zhang C.J., Yua G.H. *et al.*: Changes in chloroplast ultrastructure, fatty acid components of thylakoid membrane and chlorophyll *a* fluorescence transient in flag leaves of a super-high-yield hybrid rice and its parents during the reproductive stage. – *J. Plant Physiol.* **167**: 277-285, 2010.